ATENT COOPERATION TR. TY

From the INTERNATIONAL BUREAU

PCT

NOTIFICATION OF THE RECORDING OF A CHANGE

(PCT Rule 92bis.1 and Administrative Instructions, Section 422)

To:

DUGAN, Deborah, A.
Genzyme Corporation
Metrowest Place
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P.O. Box 9322
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ETATS-UNIS D'AMERIQUE

Date of mailing (day/month/year)
13 July 2000 (13.07.00)

Applicant's or agent's file reference
126881206140

IMPORTANT NOTIFICATION

International application No.
PCT/US99/25091

International filing date (day/month/year)
25 October 1999 (25.10.99)

1. The following indications appeared on record concerning:

[ ] the applicant [ ] the inventor [X] the agent [ ] the common representative

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Baker & McKenzie
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State of Nationality:
State of Residence:
Telephone No.: 650 856 2400
Facsimile No.: 650 856 9299
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2. The International Bureau hereby notifies the applicant that the following change has been recorded concerning:

[X] the person [X] the name [X] the address [ ] the nationality [ ] the residence

Name and Address
DUGAN, Deborah, A.
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Teleprinter No.:

3. Further observations, if necessary:
The indication of the above new agent in the Demand Form has been taken by the International Bureau as a request for the recording of a change under PCT Rule 92bis.

4. A copy of this notification has been sent to:

[X] the receiving Office
[X] the International Searching Authority
[X] the International Preliminary Examining Authority
[ ] the designated Offices concerned
[X] the elected Offices concerned
[ ] other:

The International Bureau of WIPO
34, chemin des Colombettes
1211 Geneva 20, Switzerland

Facsimile No.: (41-22) 740.14.35

Authorized officer
Ellen Moyse

Telephone No.: (41-22) 338.83.38

Form PCT/IB/306 (March 1994)

003409754

PATENT COOPERATION TREATY

From the INTERNATIONAL BUREAU

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NOTIFICATION OF ELECTION

(PCT Rule 61.2)

To:

Assistant Commissioner for Patents
United States Patent and Trademark Office
Box PCT
Washington, D.C.20231
ETATS-UNIS D'AMERIQUE

in its capacity as elected Office

Date of mailing (day/month/year) 13 July 2000 (13.07.00)	
International application No. PCT/US99/25091	**Applicant's or agent's file reference** 126881206140
International filing date (day/month/year) 25 October 1999 (25.10.99)	**Priority date** (day/month/year) 26 October 1998 (26.10.98)
Applicant IBRAGHIMOV-BESKROVNAYA, Oxana et al	

1. The designated Office is hereby notified of its election made:

[X] in the demand filed with the International Preliminary Examining Authority on:

26 May 2000 (26.05.00)

[ ] in a notice effecting later election filed with the International Bureau on:

2. The election [X] was

[ ] was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO **34, chemin des Colombettes** **1211 Geneva 20, Switzerland**	Authorized officer Ellen Moyse
Facsimile No.: (41-22) 740.14.35	Telephone No.: (41-22) 338.83.38

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International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 7 :
C07K 16/28, 14/47, C12N 5/12, A61K 39/395, 38/17, C12N 15/866, 5/10, G01N 33/577, 33/68, C12N 15/11, A61P 13/12

A3

(11) International Publication Number: WO 00/24780

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(21) International Application Number: PCT/US99/25091

(22) International Filing Date: 25 October 1999 (25.10.99)

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60/105,731	26 October 1998 (26.10.98)	US
60/105,876	27 October 1998 (27.10.98)	US
60/141,175	25 June 1999 (25.06.99)	US

(71) Applicant *(for all designated States except US)*: GENZYME CORPORATION [US/US]; Metrowest Place, 15 Pleasant Street Connector, Framingham, MA 01701-9322 (US).

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(75) Inventors/Applicants *(for US only)*: IBRAGHIMOV-BESKROVNAYA, Oxana [US/US]; Framingham, MA 01701 (US). VAN-DELLEN, K. [US/US]; Framingham, MA 01701 (US). PETRY, Linda, R. [US/US]; Framingham, MA 01701 (US).

(74) Agents: KONSKI, Antoinette, F. et al.; Baker & McKenzie, 660 Hansen Way, Palo Alto, CA 94304 (US).

(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

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9 November 2000 (09.11.00)

(54) Title: COMPOSITIONS AND METHODS FOR TREATING POLYCYSTIC KIDNEY DISEASE

(57) Abstract

The present invention provides an isolated antibody or fragment thereof that binds to an epitope present in the transmembrane domain of polycystin and specifically recognizes a polycystin-related polypeptide having an apparent molecular weight in the range of about 600 to about 800 kD. Polynucleotides, polypeptides, gene delivery vehicles and host cells containing the transmembrane sequences are also provided. Further provided are methods and compositions for modulating the biological activity of polycystin in a suitable cell or tissue.

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INTERNATIONAL SEARCH REPORT

Int. [redacted] Application No
PCT/US 99/25091

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C07K16/28 C07K14/47 C12N5/12 A61K39/395 A61K38/17
C12N15/866 C12N5/10 G01N33/577 G01N33/68 C12N15/11
A61P13/12

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C07K C12N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 95 34573 A (BRIGHAM AND WOMEN'S HOSPITAL) 21 December 1995 (1995-12-21) claims 1-35	1-29
X	VAN ADELSBERG J ET AL: "Polycystin expression is temporally and spatially regulated during renal development." AMERICAN JOURNAL OF PHYSIOLOGY, (1997) 272 F602-F609, XP000892142 page F602, line 37 - line 48	1-3,6
A	PALSSON R ET AL: "Characterization and cell distribution of polycystin, the product of autosomal dominant polycystc kidney disease gene 1." MOLECULAR MEDICINE, (1996) 2 702-11, XP000892141 abstract	1-29

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

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Date of the actual completion of the international search	Date of mailing of the international search report
20 April 2000	11/05/2000

Name and mading address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Le Flao, K

Form PCT/ISA/210 (second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 99/25091

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. [X] Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

Remark: Although claims 23-29 partially (as far as an in vivo method is concernerd) are directed to a method of treatment of the human/ animal body, the search has been carried out and based on the alleged effects of the compound/composition.

2. [ ] Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. [ ] As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int[redacted]nal Application No
PCT/US 99/25091

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9534573 A	21-12-1995	AU 2766195 A	05-01-1996
		US 5891628 A	06-04-1999

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CORRECTED VERSION

(19) World Intellectual Property Organization
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4 May 2000 (04.05.2000)

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(10) International Publication Number
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(51) International Patent Classification[7]: C07K 16/28, 14/47, C12N 5/12, A61K 39/395, 38/17, C12N 15/866, 5/10, G01N 33/577, 33/68, C12N 15/11, A61P 13/12

(21) International Application Number: PCT/US99/25091

(22) International Filing Date: 25 October 1999 (25.10.1999)

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(30) Priority Data:

60/105,731	26 October 1998 (26.10.1998)	US
60/105,876	27 October 1998 (27.10.1998)	US
60/141,175	25 June 1999 (25.06.1999)	US

(71) Applicant *(for all designated States except US)*: **GENZYME CORPORATION** [US/US]; Metrowest Place, 15 Pleasant Street Connector, Framingham, MA 01701-9322 (US).

(72) Inventors; and

(75) Inventors/Applicants *(for US only)*: **IBRAGHIMOV-BESKROVNAYA, Oxana** [US/US]; 3 Blendon Woods Drive, Southborough, MA 01772 (US). **VAN-DELLEN, K.** [US/US]; Framingham, MA 01701 (US). **PETRY, Linda, R.** [US/US]; Framingham, MA 01701 (US).

(74) Agent: **DUGAN, Deborah, A.**; Genzyme Corporation, Metrowest Place, 15 Pleasant Street Connector, P.O. Box 9322, Framingham, MA 01701-9322 (US).

(81) Designated States *(national)*: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States *(regional)*: ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

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see PCT Gazette No. 44/2001 of 1 November 2001, Section II

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 00/24780 A3

(54) Title: COMPOSITIONS AND METHODS FOR TREATING POLYCYSTIC KIDNEY DISEASE

(57) Abstract: The present invention provides an isolated antibody or fragment thereof that binds to an epitope present in the transmembrane domain of polycystin and specifically recognizes a polycystin-related polypeptide having an apparent molecular weight in the range of about 600 to about 800 kD. Polynucleotides, polypeptides, gene delivery vehicles and host cells containing the transmembrane sequences are also provided. Further provided are methods and compositions for modulating the biological activity of polycystin in a suitable cell or tissue.

COMPOSITIONS AND METHODS FOR TREATING POLYCYSTIC KIDNEY DISEASE

CROSS-REFERENCE TO RELATED APPLICATIONS

The present application claims priority under 35 U.S.C. § 119(e) to U.S. Provisional Application Nos. 60/105,731; 60/105,876; and 60/141,175, filed October 26, 1998, October 27, 1998 and June 25, 1999, respectively, the contents of which are hereby incorporated by reference into the present disclosure.

TECHNICAL FIELD

This invention is in the field of nephrology. The compositions and methods of the present invention are particularly useful in diagnoses and treatment of polycystic renal diseases.

BACKGROUND OF THE INVENTION

Polycystic kidney disease (PKD) is a common inherited condition for which there are no cures and few effective therapies. The disease can be transmitted as an autosomal dominant or recessive defect. The dominant form of PKD is one of the most prevalent life-threatening genetic diseases, affecting approximately 600,000 Americans and more than 12 million families worldwide. The National Institutes of Health estimates that one in 400 to 1,000 persons has autosomal dominant polycystic kidney disease (ADPKD), and one in 10,000 to 40,000 individuals has autosomal recessive polycystic kidney disease (ARPKD). More than fifty percent of the affected individuals are expected to develop renal failure by the age of 60; consequently, ADPKD currently accounts for 4 to 8 percent of the renal dialysis and transplantation cases in the United States and Europe (Robinson and Hawkins (1981) Proc. European Dialysis and Transplant Assn. 17:20).

Most forms of PKD are characterized by the development of fluid-filled cysts from the nephrons and collecting ducts of affected kidney tissue, which results in grossly enlarged kidneys with progressively weakened renal-concentration ability. Cyst development can also occur in other ductal organs such as liver, pancreas and spleen. Further systemic manifestations may include gastrointestinal, cardiovascular, and musculoskeletal abnormalities, such as colonic diverticulitis, berry aneurysms, hernias, and mitral valve prolapse (Gabow, et al. (1989) Adv. Nephrol. **18**:19-32 and Gabow et al. (1993) New Eng. J. Med. **329**:332-342). Hypertension and endocrine dysfunction are also common in ADPKD patients, appearing even before symptoms of renal insufficiency.

Recently, a few genetic attributes of PKD have been identified. Linkage studies and mutation analysis have indicated a causative gene (PKD1) located on chromosome 16p13.3, which is responsible for eighty-five percent of ADPKD cases (Reeders et al. (1985) Nature **317**:542-544; Breuning et al. (1987) Lancet **ii**:1359-1361). A large number of mutations in the PKD1 gene sequences have been found to be associated with the onset of polycystic kidney disease. Apart from large genomic deletions that eliminate PKD1, the mutations that have been defined clearly in ADPKD1 families appear to result in the transcription of a truncated or abnormal message RNA from the affected allele (The American PKD1 Consortium (1995) Human Mol. Genet. **4**:575-582). These gene sequence alterations include small in-frame deletions, deletions and missense mutations that result in premature termination, splice-site mutations and chromosomal translocations which interrupt the gene. Most of the other ADPKD cases can be attributed to PKD2 (Kimberling W.J. et al. (1993) Genomics **18**:467-472; Mochizuki T. et al. (1996) Science **272**:1339-1342), with less than one percent due to the third locus for ADPKD, which has not been mapped yet.

The wild-type PKD1 gene encodes a large protein, polycystin-1, which is predicted to be approximately 462 kD in size. The primary sequence of polycystin predicts a protein having structural features characteristic of a cell surface receptor or adhesion molecule. At the N-terminus, an extracellular

domain of about 3,000 amino acids contains a number of recognizable protein motifs known for their involvement in protein-protein interaction. At the C-terminus, a short cytosolic domain consisting of approximately 250 amino acids possess several phosphorylation sites and a potential PEST (proline, glutamic acid, serine, and threonine) sequence. Linking the two terminal regions is the transmembrane domain of about 1,000 amino acids in length that comprises a group of characteristic seven membrane segments also found in the G-protein coupled cell surface receptors.

Highly conserved motifs residing in the N-terminal extracellular domain include two leucine-rich repeats (LRRs) with cysteine-rich flanking regions, immunoglobulin (Ig)-like repeats, and a C-type lectin domain. Leucine-rich repeats (LRRs) are commonly found in the leucine-rich glycoprotein family, which takes part in a diversity of physiological events. Proteins sharing this homology include but are not limited to α2-glycoprotein, members of the GPIb.LX complex (von Willebrand factor receptor), *Drosophila chaoptin*, toll and slit (Burns et al. (1995) Human Mol. Genet. **4**:575-82). Many LRR proteins are localized in the plasma membrane or extracellular matrix and are thought to be involved in cell adhesion and developmental regulation (Kobe et al. (1994) Trends Biochem. Sci. **19**:415-21). At least half of the LRR-containing proteins identified thus far have been shown to be involved in signal transduction, as for example the receptor tyrosine kinases Trk, TrkB, and TrkC. In addition, C-type lectin domains are known to mediate calcium-dependent, carbohydrate binding in cell-cell and cell-matrix adhesion (The International Polycystic Kidney Disease Consortium (1995) Cell **81**:289-98).

The 16 Ig-like domains are linearly segmented within the sequence such that the first Ig-like domain is localized between the LRRs and the C-type lectin domain while the remaining 15 Ig-like domains are tandemly clustered in the middle part of the molecule. Originally thought to be members of the Ig superfamily, recent work suggests that while PKD domains contain an Ig-like

fold, they represent a novel family (Bycroft M. et al. (1999) EMBO J. **18**:297-305).

Elucidation of the biological functions of a gene often begins with examining the expression pattern of the gene product. Polyclonal and monoclonal antibodies directed against peptide or fusion proteins, mainly from the C-terminal region of polycystin, have been used to study the expression of polycystin in human and animal tissues (Ward et al. (1996) Proc. Natl. Acad. Sci. USA **93**:1524-1528; Griffin et al. (1996) Proc. Assoc. Am. Physicians **108**:185-197; Peters et al. (1996) Lab. Invest. **75**:221-230; Geng et al. (1996) J. Clin. Invest. **98**:2674-2682; Paulson et al. (1996) Molec. Med. **2**:702-711; Van Adelsberg et al. (1997) Am. J. Physiol. **272**:F602-F609; Ibraghimov-Beskrovnaya et al. (1997) Proc. Natl. Acad. Sci. USA **94**:6397-6402; Geng et al. (1997) Am. J. Physiol. **272**:F451-F459; Griffin et al. (1997) Kidney Int. **52**:1196-1205; Geng et al. (1997) J. Am. Soc. Nephrol. **8**:372A). These studies indicate that polycystin is expressed in many tissues in addition to the kidney and the liver. These include the epithelial cells of pancreatic and mammary ducts, intestinal crypts, urothelium and bronchioles; basal keratinocytes of the skin; neural crest, brain, neural plexuses and adrenal medulla; myocardium vascular smooth muscle of elastic and distributive arteries; and certain endothelial cells (Griffin et al. (1996) Proc. Assoc. Am. Physicians **108**:185-197; Geng et al. (1996) J. Clin. Invest. **98**:2674-2682; Ibraghimov-Beskrovnaya et al. (1997) Proc. Natl. Acad. Sci. USA **94**:6397-6402; Geng et al. (1997) Am. J. Physiol. **272**:F451-F459; Griffin et al. (1997) Kidney Int. **52**:1196-1205; Griffin et al. (1997) J. Am. Soc. Nephrol. **8**:616-626; O'Sullivan et al. (1997) J. Am. Soc. Nephrol. **8**:376A). Studies on the immunolocalization of polycystin in the kidney, however, yielded ambiguous results. For instance, there are conflicting observations as to whether polycystin is expressed in the glomeruli region of the kidney nephrons. There are also differing views as to whether polycystin is localized to basal and apical membranes of renal epithelial cells, and to what degree it is present in the cytoplasm.

There thus remains a considerable need for antibodies that specifically bind to endogenous polycystin and/or polycystin-related proteins for better characterization of their tissue distribution and intracellular localization. The generation of these antibodies would provide a significant contribution to elucidation of the basic biochemical mechanisms underlying the polycystic kidney disease; it would also greatly facilitate diagnosis, prognosis, and development of new and effective therapeutics for ADPKD. This invention satisfies these needs and provides related advantages as well.

DISCLOSURE OF THE INVENTION

This invention provides an isolated antibody or a fragment thereof that binds to an epitope present in the transmembrane domain of polycystin and specifically recognizes at least one novel, polycystin-related polypeptide(s) (referred to herein as "PRP" for polycystin-related polypeptide) having an apparent molecular weight in the range of about 600 to about 800 kD. The invention also provides polynucleotides, polypeptides, gene delivery vehicles and host cells useful for generating such antibodies, as well as methods for using the antibodies and/or polypeptides for diagnostic purposes.

In one aspect, the invention includes antibodies raised against an epitope present in the loop region of the polycystin transmembrane domain, wherein the epitope is selected from the group comprising amino acid residues 2621 to 2710, or residues 2734 to 3094, or residues 3116 to 3300, or residues 3364 to 3578, or residues 3623 to 3688, or residues 3710 to 3914, or residues 3931 to 4046, or residues 2166 to 2599, or residues 4097 to 4302, or residues 4148 to 4219, or residues 4220 to 4302, or residues 27 to 360, as shown in Figures 1 (SEQ ID NO:2) and 2.

In another aspect, the invention includes antibodies raised against an epitope outside the loop region but within the polycystin transmembrane domain, wherein the epitope comprises amino acid residues 2166 to 2599 as shown in Figure 1 (SEQ ID NO:2).

In yet another aspect, the invention provides at least one isolated antibody or a fragment thereof that specifically binds to the transmembrane domain of an integral membrane protein that is associated with polycystic kidney disease, wherein the integral membrane protein also binds to a reference antibody selected from the group consisting of anti-FP-L1, anti-FP-L2, anti-FP-L3, anti-FP-L4, anti-FP-L5, anti-FP-L6, anti-FP-L7, anti-MAL-REJ antibody, anti-MAL-BD3 antibody, anti-FP-46-2 antibody, anti-FP-46-1c antibody, or anti-FP-LRR antibody.

In a further aspect, the invention provides antibodies raised against the Ig-like domains of polycystin, and in particular, peptides comprising amino acids 843 to 1200 or 1205 to 1625 or 1626 to 2136, as shown in Figure 1 (SEQ ID NO:2).

In yet another aspect, the invention provides a hybridoma cell line that produces the monoclonal antibodies of the present invention.

In yet another aspect, the invention provides an isolated polypeptide (PRP) having an apparent molecular weight in the range of about 600 to about 800 kD that specifically binds to an antibody or a fragment thereof as described above.

In still another aspect, the invention provides a recombinant polypeptide comprising a polypeptide fragment of polycystin, wherein the fragment is a membrane-spanning segment of polycystin selected from the group consisting of loop 1, loop 2, loop 3, loop 4 and loop 7. In yet another aspect, the invention provides an isolated polypeptide comprising amino acid residues 2166 to 2599 of polycystin. In yet a further aspect, the polypeptide comprises at least one IgG like domain of polycystin. In still a further aspect, the polypeptide comprises amino acids 843 to 1200 or 1205 to 1625 or 1626 to 2136, as shown in Figure 1 (SEQ ID NO:2).

In still another aspect, the invention provides an isolated polynucleotide encoding the recombinant polypeptide of the present invention.

In other separate aspects, the invention provides an isolated polynucleotide, a gene delivery vehicle, or a cell encoding sequences comprising the polypeptides of the present invention.

An additional aspect of the invention is a method for producing the polypeptides by growing the cells of the invention under conditions favorable for the transcription and translation of the polynucleotide. The polypeptides can be further purified.

A further aspect of the invention also provides methods of generating an antibody or fragment thereof and the methods of using these antibodies for detecting polycystin-related proteins.

In an alternative aspect, the present invention further provides a diagnostic kit for detecting a polycystin-related polypeptide present in a sample, that contains an above-described antibody and instructions for the use of the antibody to detect the polypeptide.

In a yet further aspect, the present invention also provides methods for modulating cell-cell and cell-matrix adhesion in a suitable tissue by delivering to the tissue an effective amount of an agent that modulates the binding of polycystin to its ligand.

In an additional aspect, methods for modulating a pathology associated with disregulation of cell-cell or cell-matrix adhesion in a subject are provided by this invention.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is the polynucleotide sequence of the full-length PKD1 (also referred to herein as "polycystin") cDNA and the predicted amino acid sequence (SEQ ID NOS:1-2).

Figure 2 depicts a panel of 12 fusion proteins comprising the transmembrane sequences of polycystin.

Figure 3A is a schematic representation of the full-length coding region of the PKD1 gene and various deletion constructs of polycystin that were expressed in a baculovirus/insect system and COS cells. The schematic structure of several of expressed recombinant polycystin-1 constructs: FLC13 - full-length polycystin-1 molecule and truncated polycystins - HTM3 (amino acids 3070-4302) and Nhe

delta (deletion of amino acids 290 through 2960). Signal peptide (S), leucine rich repeats (LRR) Ig-like repeats (Ig-like), REJ-domain (REJ) and transmembrane regions (TM) are indicated. The epitopes recognized by antibodies are shown by black bars. Figure 3B shows expression of recombinant polycystin-1 and characterization of anti-polycystin-1 antibodies. Immunoblotting of insect Sf21 cells infected with wild-type virus (control), Nhe delta recombinant virus or HTM3 construct (HTM3) with anti-BD3 antibody. Figures 3 C and 3D show immunofluorescence staining using anti-BD3 antibody of Sf21 cells infected with Nhe-delta virus, or with wild-type virus as negative control respectively.

Figure 4 depicts a schematic representation of the full-length coding region of the PKD1 gene with an emphasis on the predicted, conserved domains that are also shared amongst other proteins.

Figure 5 depicts a panel of deletion constructs comprising various domains of polycystin.

Figure 6 depicts the expression of two truncation mutants of polycystin, Nhe delta and HTM3, in baculovirus/insect system.

Figure 7 depicts an immunoblot demonstrating the detection of the truncated polycystin, Nhe delta, by various antibodies.

Figure 8 depicts the expression of C-terminal part of polycystin in COS1 cells.

Figure 9 depicts the transient expression of HTM3 in COS1 cells.

Figure 10A depicts the subcellular distribution of a polycystin-related protein in kidney and liver tissues. Figure 10B depicts the differential expression of a polycystin-related protein in microsomal fraction of fetal brain and kidney tissue. Figure 10C depicts the membrane association of a polycystin-related protein in kidney and brain tissues. Figure 10D depicts the expression of a polycystin-related protein in various cell lines.

Figure 11 shows subcellular localization of polycystin-1 in MDCK cells. Immunofluorescence staining with the different anti-polycystin-1 antibodies, anti-

LRR, anti-L2 and anti-BD3, each demonstrate intercellular membrane localization of polycystin-1.

Figure 12 shows *in vitro* binding analysis. In Figure 12A, a schematic structure of the full-length polycystin-1 is indicated with structural motifs. Shown are the fusion protein constructs of Ig-like regions which were immobilized on beads (GST-Ig^a, GST-Ig^b and GST-Ig^c) and the *in vitro* translated probes (^{35}S-Ig^a, ^{35}S-Ig^b, ^{35}S-Ig^c) used for the *in vitro* binding assays. Figure 12B shows homophilic interactions of Ig-like clusters. Autoradiograms of *in vitro* translated ^{35}S-labeled probes of Ig-like regions (shown on top of each panel) specifically bound to bead-immobilized fusion proteins (indicated on the bottom of each panel as GST-Ig^a, GST-Ig^b,GST-Ig^c and GST, respectively). The first lane of each panel contains total input of ^{35}S-labeled probe used for each binding experiment. In Figure 12C, the left panel shows an autoradiogram of *in vitro* binding assay for p53 - T-antigen. ^{35}S-T-antigen probe input is shown in lane 1. Lanes 2 and 3 show probe bound to immobilized fusion proteins GST-p53 and GST carrier, respectively. The right panel represents binding of the c-terminal region of the polycystin-2 probe (input shown in the first lane) to immobilized polycystin-1 c-terminal fusion protein (lane 2, MBP-PKD1). Binding of the probe to MBP-lacZ fusion protein was used as negative control (lane 3).

Figure 13 depicts quantitative analysis of Ig-like homophilic interactions. Sepharose beads with immobilized fusion proteins (indicated as immobilized protein) were incubated with ^{35}S-labeled *in vitro* translated probes (shown below). The percentage of bound probe calculated as described in experimental procedures is plotted on the y axis. Beads with corresponding fusion protein carriers (GST or MBP-lacZ) were used as controls for background binding.

Figure 14 shows the disruption of intercellular adhesion. In Figure 14A, the effect of soluble Ig-like domains of polycystin-1 on cell-cell adhesion in MDCK cell monolayers are shown. Cell monolayers were incubated with GST-Ig^a, GST-Ig^b and GST-Ig^c fusion proteins (media+GST-Ig^{abc}). Note the separation of cells from one another and the fibroblastic morphology of cells at the edge of

the island. Cell monolayers incubated with GST protein (media+GST) or grown in the media alone show a compact regularly packed monolayer. Figure 14B shows disruption of aggregate formation by soluble Ig-like domains of polycystin-1. Single MDCK cell suspensions were assayed for their ability to form aggregates in the presence of GST-Ig^a, GST-Ig^b and GST-Ig^c (media+GST-Ig^{abc}). Note the loss of large aggregates in this sample. Formation of large aggregates can be detected easily in the media alone or in the presence of the GST carrier (media+GST) as control.

MODE(S) FOR CARRYING OUT THE INVENTION

Throughout this disclosure, various publications, patents and published patent specifications are referenced by an identifying citation. The disclosures of these publications, patents and published patent specifications are hereby incorporated by reference into the present disclosure to more fully describe the state of the art to which this invention pertains.

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of immunology, molecular biology, microbiology, cell biology and recombinant DNA, which are within the skill of the art. See, e.g., Sambrook et al. MOLECULAR CLONING: A LABORATORY MANUAL, 2nd edition (1989); CURRENT PROTOCOLS IN MOLECULAR BIOLOGY (F. M. Ausubel et al. eds., (1987)); the series METHODS IN ENZYMOLOGY (Academic Press, Inc.): PCR 2: A PRACTICAL APPROACH (M.J. MacPherson, B.D. Hames and G.R. Taylor eds. (1995)), Harlow and Lane, eds. (1988) ANTIBODIES, A LABORATORY MANUAL, and ANIMAL CELL CULTURE (R.I. Freshney ed. (1987)).

As used in the specification and claims, the singular form "a", "an" and "the" include plural references unless the context clearly dictates otherwise. For example, the term "a cell" includes a plurality of cells, including mixtures thereof.

The term "comprising" is intended to mean that the compositions and methods include the recited elements, but not excluding others. "Consisting essentially of" when used to define compositions and methods, shall mean

excluding other elements of any essential significance to the combination. Thus, a composition consisting essentially of the elements as defined herein would not exclude trace contaminants from the isolation and purification method and pharmaceutically acceptable carriers, such as phosphate buffered saline, preservatives, and the like. "Consisting of" shall mean excluding more than trace elements of other ingredients and substantial method steps for administering the compositions of this invention. Embodiments defined by each of these transition terms are within the scope of this invention.

The term "polynucleotide" refers to a polymeric form of nucleotides of any length, either deoxyribonucleotides or ribonucleotides, or analogs thereof. Polynucleotides may have any three-dimensional structure, and may perform any function, known or unknown. The following are non-limiting examples of polynucleotides: a gene or gene fragment, exons, introns, messenger RNA (mRNA), transfer RNA, ribosomal RNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers. A polynucleotide may comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs. If present, modifications to the nucleotide structure may be imparted before or after assembly of the polymer. The sequence of nucleotides may be interrupted by non-nucleotide components. A polynucleotide may be further modified after polymerization, such as by conjugation with a labeling component.

A "gene" refers to a polynucleotide containing at least one open reading frame that is capable of encoding a particular protein after being transcribed and translated.

The terms "polypeptide," "peptide" and "protein" are used interchangeably herein to refer to polymers of amino acids of any length. The polymer may be linear or branched, it may comprise modified amino acids, and it may be interrupted by non-amino acids. The terms also encompass an amino acid polymer that has been modified; for example, disulfide bond formation,

glycosylation, lipidation, acetylation, phosphorylation, or any other manipulation, such as conjugation with a labeling component. As used herein the term "amino acid" refers to either natural and/or unnatural or synthetic amino acids, including glycine and both the D or L optical isomers, and amino acid analogs and peptidomimetics.

A protein is associated with polycystic kidney disease when it is present at a substantially altered level or in a substantially altered form in the cells derived from a PDK-affected tissue compared with cells of a control tissue. Such protein may also play a role in renal cystogenesis.

An "integral membrane protein" is a transmembrane protein that extends across the lipid bilayer of the plasma membrane of a cell. A typical integral membrane protein consists of at least one "membrane spanning segment" that generally comprises hydrophobic amino acid residues. Unlike peripheral membrane proteins that can be released from the membrane by relatively gentle extraction procedures, such as exposure to solutions of very high or low ionic strength or extreme pH, integral membrane protein may be linked to the phosphatidylinositols of the bilayer, or be held in the bilayer by a fatty acid chain, and thus can be released only by disrupting the lipid bilayer with detergents or organic solvents. As used herein, "membrane associated" polypeptides include peripheral and integral membrane polypeptides that are bound to any cellular membranes including plasma membranes and membranes of intracellular organelles.

As used herein, the term "antibody" refers to a polypeptide or group of polypeptides which are comprised of at least one antibody combining site. An "antibody combining site" or "binding domain" is formed from the folding of variable domains of an antibody molecule(s) to form three-dimensional binding spaces with an internal surface shape and charge distribution complementary to the features of an epitope of an antigen, which allows an immunological reaction with the antigen. An antibody combining site may be formed from a heavy and/or a light chain domain (VH and VL, respectively), which form hypervariable loops

which contribute to antigen binding. The term "antibody" includes, for example, vertebrate antibodies, hybrid antibodies, chimeric antibodies, altered antibodies, univalent antibodies, the Fab proteins, and single domain antibodies.

An antibody "specifically binds to" or "specifically recognizes" a polypeptide if it binds with greater affinity or avidity than it binds to other reference polypeptides or substances.

"Antigen" as used herein means a substance that is recognized and bound specifically by an antibody or by a T cell antigen receptor. Antigens can include peptides, proteins, glycoproteins, polysaccharides and lipids; portions thereof and combinations thereof. The antigens can be those found in nature or can be synthetic.

As used herein, the term "epitope" is meant to include any antigenic determinant having specific affinity for the antibodies of the invention. Epitopic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and usually have specific three dimensional structural characteristics, as well as specific charge characteristics. Whereas an epitope can comprise 3 amino acids in a spatial conformation which is unique to the epitope, it generally consists of at least 5 such amino acids, and more usually, consists of at least 8-10 such amino acids. Methods of determining the spatial conformation of amino acids are known in the art, and include, for example, x-ray crystallography and 2-dimensional nuclear magnetic resonance. "Immunological reactivity" as applied to a polypeptide refers to the ability of the polypeptide to specifically bind to an antibody of the present invention. It also refers to the ability of the polypeptide to elicit a specific immune response resulting in the production of antibodies of the present invention.

As used herein, the term "isolated" means separated from constituents, cellular and otherwise, in which the polynucleotide, peptide, polypeptide, protein, antibody, or fragments thereof, are normally associated with in nature. As is apparent to those of skill in the art, a non-naturally occurring the polynucleotide, peptide, polypeptide, protein, antibody, or fragments thereof, does not require

"isolation" to distinguish it from its naturally occurring counterpart. In addition, a "concentrated", "separated" or "diluted" polynucleotide, peptide, polypeptide, protein, antibody, or fragments thereof, is distinguishable from its naturally occurring counterpart in that the concentration or number of molecules per volume is greater than "concentrated" or less than "separated" than that of its naturally occurring counterpart.

Enrichment can be measured on an absolute basis, such as weight per volume of solution, or it can be measured in relation to a second, potentially interfering substance present in the source mixture. Increasing enrichments of the embodiments of this invention are increasingly more preferred. Thus, for example, a 2-fold enrichment is preferred, 10-fold enrichment is more preferred, 100-fold enrichment is more preferred, 1000-fold enrichment is even more preferred. A substance can also be provided in an isolated state by a process of artificial assembly, such as by chemical synthesis or recombinant expression.

The "expression" refers to the process by which polynucleotides are transcribed into mRNA and/or the process by which the transcribed mRNA (also referred to as "transcript") is subsequently being translated into peptides, polypeptides, or proteins. The transcripts and the encoded polypeptides are collectedly referred to as gene product. If the polynucleotide is derived from genomic DNA, expression may include splicing of the mRNA in a eukaryotic cell.

"Differentially expressed", as applied to nucleotide sequence or polypeptide sequence in a cell or a tissue, refers to over-expression or under-expression of that sequence when compared to that detected in a control cell or tissue. Underexpression also encompass absence of expression of a particular sequence as evidenced by the absence of detectable expression in a tested sample when compared to a control.

The term "PKD-associated gene" refers to any gene which is yielding transcription or translation products at a substantially altered level or in a substantially altered form in cells derived from PDK-affected tissues compared

with cells of a control tissue, and which may play a role in renal cystogenesis. It may be a normally quiescent gene that becomes activated (such as a dominant cyst-causing gene); it may be a gene that becomes expressed at an abnormally high; it may be a gene that becomes mutated to produce a variant phenotype; it may be a gene that becomes expressed at an abnormally low level (such as a cyst suppresser gene); or it may be a gene exhibiting differential expression, in which the differential expression correlates with cyst formation or growth.

The term "hybridize" as applied to a transcript refers to the ability of the transcript to form a complex that is stabilized via hydrogen bonding between the bases of the nucleotide residues in a hybridization reaction. The hydrogen bonding may occur by Watson-Crick base pairing, Hoogstein binding, or in any other sequence-specific manner. The complex may comprise two strands forming a duplex structure, three or more strands forming a multi-stranded complex, a single self-hybridizing strand, or any combination of these. The hybridization reaction may constitute a step in a more extensive process, such as the initiation of a PCR reaction, or the enzymatic cleavage of a polynucleotide by a ribozyme.

A "gene delivery vehicle" is defined as any molecule that can carry inserted polynucleotides into a host cell. Examples of gene delivery vehicles are liposomes, viruses, such as baculovirus, adenovirus and retrovirus, bacteriophage, cosmid, plasmid, fungal vectors and other recombination vehicles typically used in the art which have been described for expression in a variety of eukaryotic and prokaryotic hosts, and may be used for gene therapy as well as for simple protein expression.

A "viral vector" is defined as a recombinantly produced virus or viral particle that comprises a polynucleotide to be delivered into a host cell, either *in vivo*, ex vivo or *in vitro*. Examples of viral vectors include retroviral vectors, adenovirus vectors, adeno-associated virus vectors and the like. In aspects where gene transfer is mediated by a retroviral vector, a vector construct refers to the polynucleotide comprising the retroviral genome or part thereof, and a therapeutic gene. As used herein, "retroviral mediated gene transfer" or "retroviral

transduction" carries the same meaning and refers to the process by which a gene or nucleic acid sequences are stably transferred into the host cell by virtue of the virus entering the cell and integrating its genome into the host cell genome. The virus can enter the host cell via its normal mechanism of infection or be modified such that it binds to a different host cell surface receptor or ligand to enter the cell. As used herein, retroviral vector refers to a viral particle capable of introducing exogenous nucleic acid into a cell through a viral or viral-like entry mechanism.

Retroviruses carry their genetic information in the form of RNA; however, once the virus infects a cell, the RNA is reverse-transcribed into the DNA form which integrates into the genomic DNA of the infected cell. The integrated DNA form is called a provirus.

In aspects where gene transfer is mediated by a DNA viral vector, such as an adenovirus (Ad) or adeno-associated virus (AAV), a vector construct refers to the polynucleotide comprising the viral genome or part thereof, and a therapeutic gene. Adenoviruses (Ads) are a relatively well characterized, homogenous group of viruses, including over 50 serotypes. (see, e.g., WO 95/27071) Ads are easy to grow and do not require integration into the host cell genome. Recombinant Ad-derived vectors, particularly those that reduce the potential for recombination and generation of wild-type virus, have also been constructed. (see, WO 95/00655; WO 95/11984). Wild-type AAV has high infectivity and specificity integrating into the host cells genome. (Hermonat and Muzyczka (1984) Proc. Natl. Acad. Sci. USA **81**:6466-6470; Lebkowski et al. (1988) Mol. Cell. Biol. **8**:3988-3996).

Gene delivery vehicles also include several non-viral vectors, including DNA/liposome complexes, and targeted viral protein DNA complexes. Liposomes that also comprise a targeting antibody or fragment thereof can be used in the methods of this invention. To enhance delivery to a cell, the nucleic acid or proteins of this invention can be conjugated to antibodies or binding fragments thereof which bind cell surface antigens, e.g., TCR, CD3 or CD4.

A "subject," "individual" or "patient" is used interchangeably herein, which refers to a vertebrate, preferably a mammal, more preferably a human.

Mammals include, but are not limited to, rabbits, murines, simians, humans, farm animals, sport animals, and pets.

A "control" is an alternative subject or sample used in an experiment for comparison purpose. A control can be "positive" or "negative." For example, where the purpose of the experiment is to detect a differentially expressed transcript or polypeptide in cell or tissue affected by a disease of concern, it is generally preferable to use a positive control (a subject or a sample from a subject, exhibiting such differential expression and syndromes characteristic of that disease), and a negative control (a subject or a sample from a subject lacking the differential expression and clinical syndrome of that disease).

The term "modulate" shall mean upregulate or downregulate as compared to a control response or wild-type response.

Antibodies and their preparation

An aspect of the present invention is the generation of an antibody capable of binding to the transmembrane domain of polycystin and which specifically recognizes at least one polycystin-related polypeptide having an apparent molecular weight of about 600 or about 800 kD. Unlike previously characterized antibodies that bind to a PKD1 polypeptide(s) of approximately 465 kD, which is consistent with the calculated molecular weight of polycystin, the antibodies of the instant invention specifically recognize an endogenous polycystin-related polypeptide having a much higher molecular weight. Such polypeptide has not been previously identified. The polypeptide is expressed in a variety of adult and fetal tissues including but not limited to kidney, liver, brain and neuronal tissues.

In one embodiment, the invention includes antibodies raised against an epitope present in the loop region of the polycystin transmembrane domain, wherein the epitope comprises amino acid residues 2621 to 2710, or residues 2734 to 3094, or residues 3116 to 3300, or residues 3364 to 3578, or residues 3623 to 3688, or residues 3710 to 3914, or residues 3931 to 4046, or residues 2166 to 2599, or residues 4097 to 4302, or residues 4148 to 4219, or residues 4220 to

4302, or residues 27 to 360, as shown in Figures 1 and 2. In another embodiment, the invention includes antibodies raised against an epitope outside the loop region but within the polycystin transmembrane domain, wherein the epitope comprises amino acid residues 2166 to 2599 as shown in Figure 1. Also within the scope of the invention are functionally equivalent epitopes, e.g. epitopes having substantially identical amino acid sequences but for a conservative amino acid substitution. Antibodies raised against these functionally equivalent epitopes are also provided in the invention.

In yet another embodiment, the invention provides an isolated antibody or fragment thereof that specifically binds to the transmembrane domain of an integral membrane protein that is associated with polycystic kidney disease, wherein the integral membrane protein also binds to a reference antibody selected from the group consisting of anti-FP-L1, anti-FP-L2, anti-FP-L3, anti-FP-L4, anti-FP-L5, anti-FP-L6, anti-FP-L7, anti-MAL-REJ antibody, anti-MAL-BD3 antibody, anti-FP-46-2 antibody, anti-FP-46-1c antibody, or anti-FP-LRR antibody (see Figure 2). Also within the scope of the invention are functionally equivalent epitopes, e.g. epitopes having substantially identical amino acid sequences but for a conservative amino acid substitution. Antibodies raised against these functionally equivalent epitopes are also provided in the invention.

Further encompassed by this invention are antibodies raised against the Ig-like domains of polycystin. Examples of such antibodies include, but are not limited to antibodies raised against peptides comprising amino acids 843 to 1200 or 1205 to 1625 or 1626 to 2136, as shown in Figure 1 (SEQ ID NO:2). Also within the scope of the invention are functionally equivalent epitopes, e.g. epitopes having substantially identical amino acid sequences but for a conservative amino acid substitution. Antibodies raised against these functionally equivalent epitopes are also provided in the invention.

The antibodies of the present invention encompass polyclonal antibodies and monoclonal antibodies. They include but are not limited to mouse, rat, and rabbit or human antibodies. This invention also encompasses functionally

equivalent antibodies and fragments thereof. As used herein with respect to the exemplified antibodies, the phrase "functional equivalent" means an antibody or fragment thereof, or any molecule having the antigen binding site (or epitope) of the antibody that cross-blocks an exemplified antibody when used in an immunoassay such as immunoblotting or immunoprecipitation.

Antibody fragments include the Fab, Fab', $F(ab')_2$, and Fv regions, or derivatives or combinations thereof. Fab, Fab', and $F(ab')_2$ regions of an immunoglobulin may be generated by enzymatic digestion of the monoclonal antibodies using techniques well known to those skilled in the art. Fab fragments may be generated by digesting the monoclonal antibody with papain and contacting the digest with a reducing agent to reductively cleave disulfide bonds. Fab' fragments may be obtained by digesting the antibody with pepsin and reductive cleavage of the fragment so produce with a reducing agent. In the absence of reductive cleavage, enzymatic digestion of the monoclonal with pepsin produces $F(ab')_2$ fragments.

It will further be appreciated that encompassed within the definition of antibody fragment is single chain antibody that can be generated as described in U.S. Pat. No. 4,704,692, as well as chimeric antibodies and humanized antibodies (Oi et al. (1986) BioTechniques **4(3):**214). Chimeric antibodies are those in which the various domains of the antibodies' heavy and light chains are coded for by DNA from more than one species.

As used herein with regard to the monoclonal antibody, the "hybridoma cell line" is intended to include all derivatives, progeny cells of the parent hybridoma that produce the monoclonal antibodies specific for the polycystin related proteins, regardless of generation of karyotypic identity.

Laboratory methods for producing polyclonal antibodies and monoclonal antibodies, as well as deducing their corresponding nucleic acid sequences, are known in the art, see Harlow and Lane (1988) *supra* and Sambrook et al. (1989) *supra*. For production of polyclonal antibodies, an appropriate host animal is selected, typically a mouse or rabbit. The substantially purified antigen, whether

the whole transmembrane domain, a fragment thereof, or a polypeptide corresponding to a segment of or the entire specific loop region within the transmembrane domain, coupled or fused to another polypeptide, is presented to the immune system of the host by methods appropriate for the host. The antigen is introduced commonly by injection into the host footpads, via intramuscular, intraperitoneal, or intradermal routes. Peptide fragments suitable for raising antibodies may be prepared by chemical synthesis, and are commonly coupled to a carrier molecule (e.g., keyhole limpet hemocyanin) and injected into a host over a period of time suitable for the production of antibodies. Alternatively, the antigen can be generated recombinantly as a fusion protein. Examples of components for these fusion proteins include, but are not limited to myc, HA, FLAG, His-6, glutathione S-transferase, maltose binding protein or the Fc portion of immunoglobulin.

The monoclonal antibodies of this invention refer to antibody compositions having a homogeneous antibody population. It is not intended to be limited as regards to the source of the antibody or the manner in which it is made. Generally, monoclonal antibodies are biologically produced by introducing protein or a fragment thereof into a suitable host, e.g., a mouse. After the appropriate period of time, the spleens of such animal is excised and individual spleen cells fused, typically, to immortalized myeloma cells under appropriate selection conditions. Thereafter the cells are clonally separated and the supernatants of each clone are tested for their production of an appropriate antibody specific for the desired region of the antigen using methods well known in the art.

The isolation of other hybridomas secreting monoclonal antibodies with the specificity of the monoclonal antibodies of the invention can also be accomplished by one of ordinary skill in the art by producing anti-idiotypic antibodies (Herlyn et al. (1986) Science **232**:100). An anti-idiotypic antibody is an antibody which recognizes unique determinants present on the monoclonal antibody produced by the hybridoma of interest.

Idiotypic identity between monoclonal antibodies of two hybridomas demonstrates that the two monoclonal antibodies are the same with respect to their recognition of the same epitopic determinant. Thus, by using antibodies to the epitopic determinants on a monoclonal antibody it is possible to identify other hybridomas expressing monoclonal antibodies of the same epitopic specificity.

It is also possible to use the anti-idiotype technology to produce monoclonal antibodies which mimic an epitope. For example, an anti-idiotypic monoclonal antibody made to a first monoclonal antibody will have a binding domain in the hypervariable region which is the mirror image of the epitope bound by the first monoclonal antibody. Thus, in this instance, the anti-idiotypic monoclonal antibody could be used for immunization for production of these antibodies.

Other suitable techniques of antibody production include, but are not limited to, *in vitro* exposure of lymphocytes to the antigenic polypeptides or selection of libraries of antibodies in phage or similar vectors. See Huse et al. (1989) Science **246**:1275-1281. Genetically engineered variants of the antibody can be produced by obtaining a polynucleotide encoding the antibody, and applying the general methods of molecular biology to introduce mutations and translate the variant. The above described antibody "derivatives" are further provided herein.

Sera harvested from the immunized animals provide a source of polyclonal antibodies. Detailed procedures for purifying specific antibody activity from a source material are known within the art. Undesired activity cross-reacting with other antigens, if present, can be removed, for example, by running the preparation over adsorbants made of those antigens attached to a solid phase and eluting or releasing the desired antibodies off the antigens. If desired, the specific antibody activity can be further purified by such techniques as protein A chromatography, ammonium sulfate precipitation, ion exchange chromatography, high-performance liquid chromatography and immunoaffinity

chromatography on a column of the immunizing polypeptide coupled to a solid support.

The specificity of an antibody refers to the ability of the antibody to distinguish polypeptides comprising the immunizing epitope from other polypeptides. If an antibody or fragment thereof being tested binds to an epitope in the transmembrane domain of polycystin and recognizes a related protein having a molecular weight of about 600 or about 800 kD, then the antibody being tested and the antibodies provided by this invention have the same specificity. An ordinary skill in the art can readily determine without undue experimentation whether an antibody shares the same specificity as an antibody of this invention by determining whether the antibody being tested prevents an antibody of this invention from binding the polypeptide(s) with which the antibody is normally reactive. If the antibody being tested competes with the antibody of the invention as shown by a decrease in binding by the antibody of this invention, then it is likely that the two antibodies bind to the same or a closely related epitope. Alternatively, one can pre-incubate the antibody of this invention with the polypeptide(s) with which it is normally reactive, and determine if the antibody being tested is inhibited in its ability to bind the antigen. If the antibody being tested is inhibited, then, in all likelihood, it has the same, or a closely related, epitopic specificity as the antibody of this invention.

The antibodies of the invention can be bound to many different carriers. Thus, this invention also provides compositions containing antibodies and a carrier. Carriers can be active and/or inert. Examples of well-known carriers include polypropylene, polystyrene, polyethylene, dextran, nylon, amylases, glass, natural and modified celluloses, polyacrylamides, agaroses and magnetite. The nature of the carrier can be either soluble or insoluble for purposes of the invention. Those skilled in the art will know of other suitable carriers for binding antibodies, or will be able to ascertain such, using routine experimentation.

The antibodies of this invention can also be conjugated to a detectable agent or a hapten. The complex is useful to detect the polypeptide(s) (or

polypeptide fragments) to which the antibody specifically binds in a sample, using standard immunochemical techniques such as immunohistochemistry as described by Harlow and Lane (1988), *supra*. There are many different labels and methods of labeling known to those of ordinary skill in the art. Examples of the types of labels which can be used in the present invention include radioisotopes, enzymes, colloidal metals, fluorescent compounds, bioluminescent compounds, and chemiluminescent compounds. Those of ordinary skill in the art will know of other suitable labels for binding to the antibody, or will be able to ascertain such, using routine experimentation. Furthermore, the binding of these labels to the antibody of the invention can be done using standard techniques common to those of ordinary skill in the art.

Another technique which may also result in greater sensitivity consists of coupling the antibodies to low molecular weight haptens. These haptens can then be specifically detected by means of a second reaction. For example, it is common to use such haptens as biotin, which reacts avidin, or dinitrophenyl, pyridoxal, and fluorescein, that can react with specific anti-hapten antibodies. See Harlow and Lane (1988), *supra*.

Polypeptides of the present invention

This present invention encompasses polypeptides separately comprising the transmembrane and Ig-like domains of a PKD1 gene product. These transmembrane domain specific polypeptides are characterized by their ability to elicit a humoral and/or cellular immune response in a host that results in production of antibodies capable of detecting novel polypeptides related to the polycystin protein family. The antibodies bind to the Ig-like domains of polycystin and block binding of polycystin to its ligand. These antibodies also useful to modulate cell-cell and cell-tissue adhesion in a suitable tissue.

The polypeptides of this invention also comprise fragments of the PKD protein comprising the Ig-like domains. In one embodiment, the polypeptide comprises regions II-V (Figure 1, amino acids 843 to 1200). In a separate

embodiment, the polypeptide comprises regions VI to X (Figure 1, amino acids 1205 to 1625). In a further embodiment, the polypeptide comprises regions XI to XVI (Figure 1, amino acids 1626 to 2136). The Ig-like polypeptides of this invention are useful to enhance or promote cell-cell or cell-matrix adhesion in a suitable tissue because they are shown to mediate interactions between these domains. In some situations, where due to mutation, a soluble form of extracellular domains, including Ig-like domains, can be produced. The soluble proteins can disrupt the cell-cell adhesion. The antibodies of this invention are useful to bind and/or remove the soluble, mutated polycystin thereby restoring normal adhesion to tissue. The antibodies are further useful in screens to identify agents that may prevent or treat pathologies related to the disregulation of the PKD gene in a subject as described above.

Such tissue includes, but is not limited to kidney, brain, liver or neuronal. Additional suitable tissues can be screened using the antibodies that specifically recognize and bind the loop domains. If the antibody binds to the tissue, the tissue expresses polycystin.

This invention also provides a novel polypeptide that differs from the previously characterized polycystin polypeptides in that they contain additional amino acid sequences and/or post-translationally modified motifs, and exhibit a mobility on a SDS-PAGE gel of about 600 kD or about 800 kD, that are approximately 200 to 400 kD higher than that predicted for polycystin.

In one embodiment, a polypeptide comprises transmembrane sequences of polycystin corresponding to a specific loop region. According to the predicted structure, loops 1, 3, 4, 5 and 7 reside on the intracellular side of the plasma membrane, whereas loops 2 and 6 extend primarily to the extracellular side of the plasma membrane (see Fig. 2). The predicted amino acid sequence of full-length polycystin is shown in Figure 1 (SEQ ID NO:2). Accordingly, the invention includes a polypeptide comprising the transmembrane domain sequences selected from the group consisting of loop 1, loop 2, loop 3, loop 4, and loop 7 (see Fig. 2, and the description in U.S. Patent No. 5,654,170).

In another embodiment, a polypeptide comprises sequences residing outside the seven loop regions but within the transmembrane domain. For example, polypeptides comprise residues 2166 to 2599 or residues 27-360, of polycystin as shown in Figures 1 and 2.

In yet another embodiment, the present invention provides an isolated polypeptide having an apparent molecular weight of about 600 or about 800 kD, which specifically binds to an above-described antibody or fragment thereof. The polypeptide exhibits sequence homology with polycystin, as it binds to the antibodies raised against epitopes present in the transmembrane domain of polycystin. It can be isolated from cellular constituents with which it is normally associated by conventional protein purification techniques. Non limiting examples include ammonium sulfate precipitation, gel electrophoresis, ion exchange chromatography, and high-performance liquid chromatography. A preferred method is immunoaffinity chromatography using antibodies to which the polypeptide binds. Where desired, the amino acid sequences of the 600 kD and 800 kD protein and fragments thereof can be determined by methods well established in the art.

In one embodiment, the polypeptide is expressed in a tissue selected from the group consisting of kidney, brain, liver and neuronal tissues. In another embodiment, the polypeptide is associated with cellular membranes including the plasma membrane and membranes of cellular organelles. Non limiting examples of cellular organelles include Golgi, endoplasmic reticulum, lysosome, and mitochondria. In yet another embodiment, the polypeptide is an integral membrane protein. In still another embodiment, the polypeptide is a cytosolic protein (i.e., distributed predominantly or about equally in the membrane and cytosolic fractions). Such polypeptide may be an isoform of polycystin that is unprocessed, variably spliced, or differentially expressed in cells or tissues, such as those affected by polycystic kidney disease. The polypeptide may also be a mutated variant that is involved in pathogenic events leading to kidney cyst formation.

It is understood that biological or functional equivalents or derivatives of the exemplified polypeptides are also encompassed by this invention. A "functionally equivalent" varies from the native sequence disclosed herein by any combination of additions, deletions, or substitutions while preserving at least one functional property of the fragment relevant to the context in which it is being used. A functional equivalent of a polypeptide of the invention typically has the ability to elicit an immune response with a similar antigen specificity as that elicited by exemplified polypeptides or to mediate cell-cell or cell-matrix adhesion. For example, the size of the polypeptide fragments useful for immunizing a host may vary widely, as the length required to affect an immune response could be as small as, for example, a 3-mer amino acid sequence. The maximum length generally is not detrimental to effecting activity. The minimum size must be sufficient to provide a desired function. Thus, the invention includes polypeptide fragments comprising a portion of the transmembrane amino acid sequences exemplified herein, in which the polypeptide is at least about 3, more preferably about 50, more preferably about 75, more preferably 100, more preferably 200 or more, amino acids in length. As is apparent to one skilled in the art, these polypeptides, regardless of their size, may also be associated with, or conjugated with, other substances or agents to facilitate, enhance, or modulate their function.

The invention includes modified polypeptides containing conservative or non-conservative substitutions that do not significantly affect their properties, such as the immunogenicity of the peptides. Modification of polypeptides is routine practice in the art. Amino acid residues which can be conservatively substituted for one another include but are not limited to: glycine/alanine; valine/isoleucine/leucine; asparagine/glutamine; aspartic acid/glutamic acid; serine/threonine; lysine/arginine; and phenylalanine/tryosine. These polypeptides also include glycosylated and nonglycosylated polypeptides, as well as polypeptides with other post-translational modifications, such as, for example, glycosylation with different sugars, acetylation, and phosphorylation.

The polypeptides of the invention can also be conjugated to a chemically functional moiety. Typically, the moiety is a label capable of producing a detectable signal. These conjugated polypeptides are useful, for example, in detection systems such as imaging of renal cysts. Such labels are known in the art and include, but are not limited to, radioisotopes, enzymes, fluorescent compounds, chemiluminescent compounds, bioluminescent compounds substrate cofactors and inhibitors. See, for examples of patents teaching the use of such labels, U.S. Patent Nos. 3,817,837; 3,850,752; 3,939,350; 3,996,345; 4,277,437; 4,275,149; and 4,366,241. The moieties can be covalently linked to the polypeptides, recombinantly linked, or conjugated to the polypeptides through a secondary reagent, such as a second antibody, protein A, or a biotin-avidin complex.

Other functional moieties include agents that enhance immunological reactivity, agents that facilitate coupling to a solid support, vaccine carriers, bioresponse modifiers, paramagnetic labels and drugs. Agents that enhance immunological reactivity include, but are not limited to, bacterial superantigens. Agents that facilitate coupling to a solid support include, but are not limited to, biotin or avidin. Immunogen carriers include, but are not limited to, any physiologically acceptable buffers.

The invention also encompasses fusion proteins comprising polycystin transmembrane sequences and Ig-like domains and fragments thereof. Such fusion may be between two or more polycystin transmembrane or Ig-like sequences or between the sequences of polycystin and a related or unrelated polypeptide. Useful fusion partners include sequences that enhance immunological reactivity, or facilitate the coupling of the polypeptide to an immunoassay support or a vaccine carrier. For instance, the polycystin transmembrane sequences can be fused with a bioresponse modifier. Examples of bioresponse modifiers include, but are not limited to, cytokines or lymphokines such as interleukin-2 (IL-2), interleukin 4 (IL-4), GM-CSF, and interferon. Another useful fusion sequence is one that facilitates purification. Examples of

such sequences are known in the art and include those encoding epitopes such as Myc, HA (derived from influenza virus hemagglutinin), His-6, or FLAG. Other fusion sequences that facilitate purification are derived from proteins such as glutathione S-transferase (GST), maltose-binding protein (MBP), or the Fc portion of immunoglobulin. For immunological purposes, tandemly repeated polypeptide segments may be used as antigens, thereby producing highly immunogenic proteins.

The proteins of this invention also can be combined with various liquid phase carriers, such as sterile or aqueous solutions, pharmaceutically acceptable carriers, suspensions and emulsions. Examples of non-aqueous solvents include propyl ethylene glycol, polyethylene glycol and vegetable oils. When used to prepare antibodies, the carriers also can include an adjuvant that is useful to non-specifically augment a specific immune response. A skilled artisan can easily determine whether an adjuvant is required and select one. However, for the purpose of illustration only, suitable adjuvants include, but are not limited to Freund's Complete and Incomplete, mineral salts and polynucleotides.

The proteins and polypeptides of this invention are obtainable by a number of processes well known to those of skill in the art, which include purification, chemical synthesis and recombinant methods. Full-length proteins can be purified from a cell derived from polycystic tissue or tissue lysate by methods such as immunoprecipitation with antibody, and standard techniques such as gel filtration, ion-exchange, reversed-phase, and affinity chromatography using a fusion protein as shown herein. For such methodology, see for example, Deutscher et al. (1999) GUIDE TO PROTEIN PURIFICATION: METHODS IN ENZYMOLOGY (Vol. 182, Academic Press). Accordingly, this invention also provides the processes for obtaining these proteins and polypeptides as well as the products obtainable and obtained by these processes.

The proteins and polypeptides also can be obtained by chemical synthesis using a commercially available automated peptide synthesizer such as those manufactured by Perkin Elmer/Applied Biosystems, Inc., Model 430A or 431A,

Foster City, CA. The synthesized protein or polypeptide can be precipitated and further purified, for example by high performance liquid chromatography (HPLC). Accordingly, this invention also provides a process for chemically synthesizing the proteins of this invention by providing the sequence of the protein and reagents, such as amino acids and enzymes and linking together the amino acids in the proper orientation and linear sequence.

Alternatively, the proteins and polypeptides can be generated recombinantly by expressing polynucleotides using the vector systems and host cells as described in the section that follows.

The polypeptides or proteins embodied in the present invention can be characterized in several ways. For instance, a polycystin-related polypeptide may be tested for its ability to bind specifically to an antibody described herein, or for its ability to specifically interfere the binding between another polypeptide and an antibody of the present invention. The ability of a polypeptide to bind specific antibodies can be tested by immunoassay. In one such assay, the antibody is labeled. Suitable labels include radioisotopes such as ^{125}I, enzymes such as peroxidase, fluorescent labels such as fluorescein, and chemiluminescent labels. Typically, the other binding partner is immobilized to a solid phase, e.g., by coating onto a microtiter plate or by coupling to beads. For such solid-phase assay, the unreacted antibodies are removed by washing. In a liquid-phase assay, however, the unreacted antibodies are removed by some other separation technique, such as filtration or chromatography. After binding the polypeptides to the antibodies, the amount of bound label is determined. A variation of this technique is a competitive assay, in which the tested polypeptide is titered for its ability to decrease the binding of antibodies specific for, e.g., the 600 kD or 800 kD polycystin-related protein.

Polynucleotides, vectors and cells of the present invention

The invention provides various polynucleotides that encode the polypeptides of the invention. The polynucleotides are selected based on the

predicted transmembrane and Ig-like domain sequences of the PKD1 gene. The transmembrane polynucleotides yield proteins or polypeptides that elicit, in a suitable host, domain specific antibodies that are capable of binding to a novel polypeptide exhibiting a molecular mobility (approximately 600 kD or 800 kD on a SDS-PAGE gel) distinct from the previously characterized polycystin protein. The Ig-like domain polynucleotides yield proteins or polypeptides that mediate or facilitate cell-cell or cell-matrix adhesion.

In one embodiment, the invention encompasses an isolated polynucleotide encoding a polypeptide having immunological activity of a polypeptide comprising sequences of the transmembrane loop region 1, 2, 3, 4 or 7. In another embodiment, an isolated polynucleotide encodes a polypeptide comprising sequences corresponding to amino acid residues 2621 to 2710, or residues 2734 to 3094, or residues 3116 to 3300, or residues 3364 to 3578, or residues 3623 to 3688, or residues 3710 to 3914, or residues 3931 to 4046, or residues 2166 to 2599, or residues 4097 to 4302, or residues 4148 to 4219, or residues 4220 to 4302, or residues 27 to 360, as shown in Figures 1 and 2. In a further embodiment, an isolated polynucleotide encodes a polypeptide corresponding to the Ig-like domains in polycystin-1. Such polypeptides include, but are not limited to polypeptides comprising amino acids 843 to 1200 or 1205 to 1625 or 1626 to 2136, as shown in Figure 1.

It is understood that the polynucleotides embodied in the invention include those coding for biological equivalents and fragments of the exemplified polypeptides. Biologically equivalent polypeptides include those which do not significantly affect properties of the polypeptides encoded thereby. Biological equivalents include, but are not limited to polypeptides having conservative amino acid substitutions, analogs including fusions, and muteins.

While the length of a polynucleotide may vary widely, the polynucleotide of the present invention preferably comprises at least 15 consecutive nucleotides, preferably at least about 150 consecutive nucleotides, more preferably at least about 225 consecutive nucleotides, even more preferably at least about 300

consecutive nucleotides, still more preferably at least about 300 consecutive nucleotides, that hybridizes with a polynucleotide encoding a polypeptide comprising sequences of the transmembrane loop region 1, 2, 3, 4, or 7. A preferred polynucleotide forms a hybrid with a polynucleotide encoding residues 2621 to 2710, or residues 2734 to 3094, or residues 3116 to 3300, or residues 3364 to 3578, or residues 3623 to 3688, or residues 3710 to 3914, or residues 3931 to 4046, or residues 2166 to 2599, or residues 4097 to 4302, or residues 4148 to 4219, or residues 4220 to 4302, or residues 27 to 360, as shown in Figures 1 and 2. In an alternative embodiment, the polynucleotides hybridize under moderate or stringent conditions to the polynucleotides that encode a polypeptide comprising amino acids 843 to 1200 or 1205 to 1625 or 1626 to 2136, as shown in Figure 1.

Hybridization can be performed under conditions of different "stringency." Conditions that vary levels of stringency are well known in the art. See, for example, Sambrook et al., *supra*. Briefly, relevant conditions include temperature, ionic strength, time of incubation, the presence of additional solutes in the reaction mixture such as formamide, and the washing procedure. Higher stringency conditions are those conditions, such as higher temperature and lower sodium ion concentration, which require higher minimum complementarity between hybridizing elements for a stable hybridization complex to form. In general, a low stringency hybridization reaction is carried out at about 40°C in 10 x SSC or a solution of equivalent ionic strength/temperature. A moderate stringency hybridization is typically performed at about 50°C in 6 X SSC, and a high stringency hybridization reaction is generally performed at about 60°C in 1 X SSC. In choosing a polynucleotide most closely related to those encoding the exemplary polypeptides, stringent hybridization is preferred.

This invention also encompasses "biologically equivalent" polynucleotides that encode polypeptides having the biological activity of wild-type polypeptides, but differ in primary polypeptide or polynucleotide sequences. Biologically equivalent polynucleotides can be identified using sequence homology searches.

Several embodiments of biologically equivalent polynucleotides are within the scope of this invention, e.g., those characterized by possessing at least 75%, or at least 80%, or at least 90% or at least 95% sequence homology as determined using a sequence alignment program under default parameters correcting for ambiguities in the sequence data, changes in nucleotide sequence that do not alter the amino acid sequence because of degeneracy of the genetic code, conservative amino acid substitutions and corresponding changes in nucleotide sequence, and variations in the lengths of the aligned sequences due to splicing variants or small deletions or insertions between sequences that do not affect function.

A variety of software programs are available in the art. Non-limiting examples of these programs are BLAST family programs including BLASTN, BLASTP, BLASTX, TBLASTN, and TBLASTX (BLAST is available from the worldwide web at http://www.ncbi.nlm.nih.gov/BLAST/), FastA, Compare, DotPlot, BestFit, GAP, FrameAlign, ClustalW, and PileUp. These programs can be obtained commercially in a comprehensive package of sequence analysis software such as GCG Inc.'s Wisconsin Package. Other similar analysis and alignment programs can be purchased from various providers such as DNA Star's MegAlign, or the alignment programs in GeneJockey. Alternatively, sequence analysis and alignment programs can be accessed through the world wide web at sites such as the CMS Molecular Biology Resource at http://www.sdsc.edu/ResTools/cmshp.html. Any sequence database that contains DNA or protein sequences corresponding to a gene or a segment thereof can be used for sequence analysis. Commonly employed databases include but are not limited to GenBank, EMBL, DDBJ, PDB, SWISS-PROT, EST, STS, GSS, and HTGS. Sequence similarity can be discerned by aligning the tag sequence against a DNA sequence database. Alternatively, the tag sequence can be translated into six reading frames; the predicted peptide sequences of all possible reading frames are then compared to individual sequences stored in a protein database such as the BLASTX program.

Parameters for determining the extent of homology set forth by one or more of the aforementioned alignment programs are well established in the art. They include, but are not limited to, p value, percent sequence identity and the percent sequence similarity. P value is the probability that the alignment is produced by chance. For a single alignment, the p value can be calculated according to Karlin et al. (1990) Proc. Natl. Acad. Sci. USA 87:2246. For multiple alignments, the p value can be calculated using a heuristic approach such as the one programmed in BLAST. Percent sequence identify is defined by the ratio of the number of nucleotide or amino acid matches between the query sequence and the known sequence when the two are optimally aligned. The percent sequence similarity is calculated in the same way as percent identity except one scores amino acids that are different but similar as positive when calculating the percent similarity.. Thus, conservative changes that occur frequently without altering function, such as a change from one basic amino acid to another or a change from one hydrophobic amino acid to another are scored as if they were identical.

The polynucleotides can be conjugated to a detectable marker, e.g., an enzymatic label or a radioisotope for detection of nucleic acid and/or expression of the gene in a cell. A wide variety of appropriate detectable markers are known in the art, including fluorescent, radioactive, enzymatic or other ligands, such as avidin/biotin, which are capable of giving a detectable signal. In preferred embodiments, one will likely desire to employ a fluorescent label or an enzyme tag, such as urease, alkaline phosphatase or peroxidase, instead of radioactive or other environmental undesirable reagents. In the case of enzyme tags, colorimetric indicator substrates are known which can be employed to provide a means visible to the human eye or spectrophotometrically, to identify specific hybridization with complementary nucleic acid-containing samples.

The polynucleotides of the invention can comprise additional sequences, such as additional encoding sequences within the same transcription unit, controlling elements such as promoters, ribosome binding sites, and

polyadenylation sites, additional transcription units under control of the same or a different promoter, sequences that permit cloning, expression, and transformation of a host cell, and any such construct as may be desirable to provide embodiments of this invention.

The polynucleotides embodied in this invention can be obtained using chemical synthesis, recombinant cloning methods, PCR, or any combination thereof. Methods of chemical polynucleotide synthesis are well known in the art and need not be described in detail herein. One of skill in the art can use the sequence data provided herein to obtain a desired polynucleotide by employing a DNA synthesizer or ordering from a commercial service.

Polynucleotides comprising a desired sequence can be inserted into a suitable vector, and the vector in turn can be introduced into a suitable host cell for replication and amplification. Polynucleotides can be introduced into host cells by any means known in the art. Cells are transformed by introducing an exogenous polynucleotide by direct uptake, endocytosis, transfection, f-mating or electroporation. Once introduced, the exogenous polynucleotide can be maintained within the cell as a non-integrated vector (such as a plasmid) or integrated into the host cell genome. Amplified DNA can be isolated from the host cell by standard methods. See, e.g., Sambrook, et al. (1989) *supra*. RNA can also be obtained from transformed host cell, or it can be obtained directly from the DNA by using a DNA-dependent RNA polymerase.

The present invention further encompasses a variety of gene delivery vehicles comprising the polynucleotide of the present invention. Gene delivery vehicles include both viral and non-viral vectors such as naked plasmid DNA or DNA/liposome complexes. Vectors are generally categorized into cloning and expression vectors. Cloning vectors are useful for obtaining replicate copies of the polynucleotides they contain, or as a means of storing the polynucleotides in a depository for future recovery. Expression vectors (and host cells containing these expression vectors) can be used to obtain polypeptides produced from the polynucleotides they contain. Suitable cloning and expression vectors include any

known in the art, e.g., those for use in bacterial, mammalian, yeast and insect expression systems. The polypeptides produced in the various expression systems are also within the scope of the invention.

Cloning and expression vectors typically contain a selectable marker (for example, a gene encoding a protein necessary for the survival or growth of a host cell transformed with the vector), although such a marker gene can be carried on another polynucleotide sequence co-introduced into the host cell. Only those host cells into which a selectable gene has been introduced will grow under selective conditions. Typical selection genes either: (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate; (b) complement autotrophic deficiencies; or (c) supply critical nutrients not available from complex media. The choice of the proper marker gene will depend on the host cell, and appropriate genes for different hosts are known in the art. Vectors also typically contain a replication system recognized by the host.

Suitable cloning vectors can be constructed according to standard techniques, or selected from a large number of cloning vectors available in the art. While the cloning vector selected may vary according to the host cell intended to be used, useful cloning vectors will generally have the ability to self-replicate, may possess a single target for a particular restriction endonuclease, or may carry marker genes. Suitable examples include plasmids and bacterial viruses, e.g., pBR322, pMB9, ColE1, pCR1, RP4, pUC18, mp18, mp19, phage DNAs, and shuttle vectors such as pSA3 and pAT28. These and other cloning vectors are available from commercial vendors such as Clontech, BioRad, Stratagene, and Invitrogen.

Expression vectors containing these nucleic acids are useful to obtain host vector systems to produce proteins and polypeptides. It is implied that these expression vectors must be replicable in the host organisms either as episomes or as an integral part of the chromosomal DNA. Suitable expression vectors include plasmids, viral vectors, including adenoviruses, adeno-associated viruses, retroviruses, cosmids, etc. A number of expression vectors suitable for expression

in eukaryotic cells including yeast, avian, and mammalian cells are known in the art. One example of an expression vector is pcDNA3 (Invitrogen, San Diego, CA), in which transcription is driven by the cytomegalovirus (CMV) early promoter/enhancer. A particularly useful expression vector (system) is the baculovirus/insect system. Suitable vectors for expression in the baculovirus system include pBackPack9 (Clontech), pPbac and pMbac (Strategene). Adenoviral vectors are particularly useful for introducing genes into tissues *in vivo* because of their high levels of expression and efficient transformation of cells both *in vitro* and *in vivo*.

A vector of this invention can contain one or more polynucleotides encoding a polycystin transmembrane polypeptide. It can also contain polynucleotide sequences encoding other polypeptides that enhance, facilitate, or modulate the desired result, such as fusion components that facilitate protein purification, and sequences that increase immunogenicity of the resultant protein or polypeptide.

The vectors containing the polynucleotides of interest can be introduced into the host cell by any of a number of appropriate means, including electroporation, transfection employing calcium chloride, rubidium chloride, calcium phosphate, DEAE-dextran, or other substances; microprojectile bombardment; lipofection; and infection (where the vector is an infectious agent, such as vaccinia virus, which is discussed below). The choice of introducing vectors or polynucleotides will often depend on features of the host cell.

Once introduced into a suitable host cell, expression of a polycystin polypeptide can be determined using any assay known in the art. For example, presence of the polypeptide can be detected by RIA or ELISA of the culture supernatant (if the polypeptide is secreted) or cell lysates using antibodies reactive with the polycystin sequences or the fusion components (if also linked to the polypeptide).

Also embodied in the present invention are host cells transformed with polycystin polynucleotides as described above. Both prokaryotic and eukaryotic

host cells may be used. Prokaryotic hosts include bacterial cells, for example *E. coli* and Mycobacteria. Among eukaryotic hosts are yeast, insect, avian, plant and mammalian cells. Host systems are known in the art and need not be described in detail herein. Examples of mammalian host cells include but not limited to COS, HeLa, and CHO cells. Baculovirus systems are preferred.

The host cells of this invention can be used, inter alia, as repositories of polycystin polynucleotides, or as vehicles for production of polycystin polynucleotides and polypeptides.

The polynucleotides and gene delivery vehicles of this invention have several uses. They are useful, for example, in expression systems for the production of polycystin or polycystin-related polypeptides. They are also useful as hybridization probes to assay for the presence of polycystin polynucleotide or related sequences in a sample using methods well known to those in the art. Further, the polynucleotides are also useful as primers to effect amplification of desired polynucleotides. The polynucleotides of this invention are also useful in pharmaceutical compositions including vaccines and for gene therapy.

Uses of antibodies and polypeptides of the present invention

The antibodies and polypeptides embodied in this invention provide specific reagents that can be used in standard diagnostic procedures. Accordingly, the invention provides a method for detecting a polycystin-related polypeptide or tissue containing the polypeptide by contacting a sample suspected of containing the polypeptide with an antibody described herein. The presence of an antibody-antigen complex is indicative of the presence of the polycystin-related polypeptide.

Generally, to perform a diagnostic method of this invention, one of the compositions of this invention is provided as a reagent to detect a target in a sample with which it reacts. The target is supplied by obtaining a suitable biological sample from an individual for whom the diagnostic parameter is to be measured. Relevant biological samples are those obtained from individuals

suspected of having polycystin kidney disease. A number of tissues are prone to develop cysts during the progression of PKD. These tissues include but are not limited to kidney, liver, spleen, brain, as well as gastrointestinal, cardiovascular and musculoskeletal tissues. Cells or tissue sample used for a diagnostic analysis encompass body fluid, solid tissue samples, tissue cultures or cells derived therefrom and the progeny thereof, and sections of smears prepared from any of these sources. Typically, cells are obtained by resection, biopsy or endoscopic sampling; the cells may be used directly, stored frozen, maintained or expanded in culture. Non-limiting examples of cell types useful for detecting the presence of polycystin and/or polycystin-related protein include epithelial cells, endothelial cells, neuronal cells, and interstitial fibroblasts. If desired, the target may be partially purified from the sample before the assay is conducted.

The reaction is performed by contacting the antibody with the sample under conditions that will allow a complex to form between the antibody and the target. The reaction may be performed in solution, or on a solid tissue sample, for example, using histology sections. The formation of the complex is detected by a number of techniques known in the art. For example, the antibodies may be supplied with a label and unreacted antibodies may be removed from the complex; the amount of remaining label thereby indicating the amount of complex formed.

The amount of the polypeptides that are immunologically reactive with the antibodies of the present invention can be quantified by standard quantitative immunoassays. If the protein is secreted or shed from the cell in any appreciable amount, it may be detectable in plasma or serum samples. Alternatively, the target protein may be solubilized or extracted from a solid tissue sample. Before quantification, the protein may optionally be affixed to a solid phase, such as by a blot technique or using a capture antibody. A number of immunoassay methods are established in the art for performing the quantitation. For example, the protein may be mixed with a predetermined non-limiting amount of the reagent antibody specific for the protein. The reagent antibody may contain a directly attached label, such as an enzyme or a radioisotope, or a second labeled reagent may be

added, such as anti-immunoglobulin or protein A. For a solid-phase assay, unreacted reagents are removed by washing. For a liquid-phase assay, unreacted reagents are removed by some other separation technique, such as filtration or chromatography. The amount of label captured in the complex is positively related to the amount of target protein present in the test sample. Alternatively, a competitive assay in which the target protein is tested for its ability to compete with a labeled analog for binding sites on the specific antibody. In this case, the amount of label captured is negatively related to the amount of target protein present in a test sample. Results obtained using any such assay on a sample from a suspected polycyst-bearing source are compared with those from a non-polycystic source.

One important application of immunoassays employing the antibodies of the present invention is the determination of tissue and/or intracellular localization of the endogenous polycystin-related proteins. To discern the tissue distribution, frozen or fixed tissue sections and/or tissue homogenates can be stained using an above-described antibody at various concentrations. In testing each tissue for the expression of a polycystin-related proteins, it is also preferable to include a antibody known to react with a tissue-specific antigen that is differentially expressed in the tested tissue. Procedures for conducting immunohistological analysis are well established in the art and thus they are not detailed herein.

Also available in the art are a variety of techniques for examining the intracellular localization of a target polypeptide. Such techniques range from subcellular fractionation to cytoimmuno-staining and electron microscopy. Cell fractionation enables partial or complete separation of individual cellular organelles. An exemplary fractionation system is the hybrid Percoll/metrizamide discontinuous density gradient as described in (Storrie et al. (1990) Meth. Enzymol. **182**:203-225). This gradient system allows the isolation of cell organelles including lysosomes, mitochondria and partial separation of plasma membrane from cytosol and organelles such as Golgi apparatus and endoplasmic reticulum. Cells suitable for such fractionation analysis include but are not

limited to CHO cells and COS cells, that preferably overexpress the target polypeptides in order to enhance the detectable signal. After cell fractionation, various subcellular factions are typically assayed for the presence of the target polypeptide by immunoblotting with an appropriate antibody.

Cytoimmunostaining reveals the subcellular distribution of a target polypeptide by direct binding of an antibody specific for the target polypeptide present in a fixed cell. Typically, the cell to be stained is attached to a solid support to allow easy handling in the subsequent procedures. The second step for cell staining usually is to fix and permeabilize the cell to ensure free access of the antibody, although this step can be omitted when examining cell-surface antigens. After incubating cell preparations with the antibody, unbound antibody is removed by washing, and the bound antibody is detected either directly (if the primary antibody is labeled) or, more commonly, indirectly visualized using a labeled secondary antibody. In localizing a target polypeptide to a specific subcellular structure in a cell, co-staining with one or more marker antibodies specific for antigens differentially present in such structure is preferably performed. A battery of organelle specific antibodies is available in the art. Non-limiting examples include plasma membrane specific antibodies reactive with cell surface receptor HER2, ER specific antibodies directed to the ER resident protein Bip, and Golgi specific antibody α-adaptin. To detect and quantify the immunospecific binding, digital image analysis system coupled to conventional or confocal microscopy can be employed.

Applying the above described general techniques, a panel of approximately 8 domain-specific polyclonal antibodies as shown in Figures 1 and 2 detected in the crude membrane fractions of fetal kidney, liver as well as epithelial and astrocytoma cell lines, an endogenous polycystin-related protein of about 800 kD. The same antibodies recognized a smaller protein of approximately 600 kD in the membrane and cytosolic fractions of fetal brain. Expression of recombinant polycystin was characterized by immunoblotting and immunofluorescence analysis of COS cells, transiently expressing the full-length

polycystin and four different truncated variants. Truncated polycystin was localized to the Golgi apparatus, while the full-length polycystin exhibited a different pattern of expression.

Discerning the tissue distribution and subcellular localization of polycystin-related proteins is of prime importance in elucidating the biological functions of these proteins. It can also be used for pathology studies. To determine whether the amount of a polycystin-related proteins, particularly the ~600 kD or ~800 kD proteins is representative of polycyst-bearing tissue or cell, a comparative immunoassay involving tissues or cells suspected to be affected by the disease are compared with a suitable control sample. The selection of an appropriate control cell or tissue is dependent on the sample cell or tissue initially selected and its phenotype which is under investigation. Whereas the sample cell is derived from a polycystic tissue, one or more counterparts of non-polycystic precursors of the sample cell can be used as control cells. Counterparts would include, for example, cell lines established from the same or related cells to those found in the sample cell population. Preferably, a control matches the tissue, and/or cell type the tested sample is derived from. It is also preferable to analyze the control and the tested sample in parallel.

Kits comprising antibodies of the present invention

The present invention also encompasses kits containing the antibodies of this invention, preferably diagnostic kits. Kits embodied by this invention include those that allow someone to detect the presence or quantify the amount of a polycystin-related protein (particularly those having a molecular weight of ~600 kD or about ~800 kD) that are suspected to be present in a sample. The sample is optionally pre-treated for enrichment of the target being tested for. The user then applies a reagent contained in the kit in order to detect the changed level or alteration in the diagnostic component.

Each kit necessarily comprises the reagent which renders the procedure specific: a reagent antibody, used for detecting target protein; and optionally a

reagent polypeptide, used as a control for the antibody, or used for detecting target antibody that may be present in a sample to be analyzed. Optionally, the antibody contained in the kits may be conjugated with a label to permit detection of any complex formed with the target in the sample. Alternatively, a second reagent is provided that is capable of combining with the first reagent after it has bound to its target and thereby supplying the detectable label. For example, labeled anti-rabbit IgG may be provided as a secondary reagent for use with the exemplified polyclonal antibodies. Labeled avidin may be provided as a secondary reagent when the primary reagent has been conjugated with biotin.

Each reagent can be supplied in a solid form or dissolved/suspended in a liquid buffer suitable for inventory storage, and later for exchange or addition into the reaction medium when the test is performed. Suitable packaging is provided. The kit can optionally provide additional components that are useful in the procedure. These optional components include, but are not limited to, buffers, capture reagents, developing reagents, labels, reacting surfaces, means for detection, control samples, instructions, and interpretive information. The kits can be employed to test a variety of biological samples, including body fluid, solid tissue samples, tissue cultures or cells derived therefrom and the progeny thereof, and sections or smears prepared from any of these sources. Diagnostic procedures using the antibodies of this invention can be performed by diagnostic laboratories, experimental laboratories, practitioners, or private individuals.

Methods for modulating the biological activity of polycystin

Anti-fusion protein antibodies against three distant regions along the molecule were constructed. The production and characterization of antibodies against the N-terminal domain (anti-LRR) and C-terminal domain (anti-BD3) have previously been described (Ibraghimov-Beskrovnaya O. et al. (1997) Proc. Natl. Acad. Sci. USA 94:6397-6402). Anti-L2 antibody, which is positioned in the middle region of polycystin-1 in the REJ domain was constructed as described above.

The specificity of the anti-polycystin-1 antibodies was examined using recombinant polycystin-1.

Anti-L2 antibody specificity against the GST-L2 fusion protein expressed in bacterial cells was tested. Anti-L2 antibody specifically recognized the L2 domain when fused to GST. Additionally, these antibodies were able to precipitate *in vitro* translated polycystin-1 specifically. Thus, the antibodies used in this study were rigorously characterized for their ability to immunoprecipitate *in vitro* translated polycystin-1 as well as by Western and immunofluorescence analysis of recombinant polycystin-1.

To determine the subcellular localization of endogenous polycystin-1 in epithelial cells, immunostaining of polycystin-1 in MDCK cells was performed with antibodies. The antibodies used were to the N-terminal region (anti-LRR), C-terminal region (anti-BD3) and to the REJ domain in the middle portion of the protein (anti-L2). All antibodies showed clearly recognizable membrane staining at sites of cell-cell contact (Figure 11). No staining was observed with the secondary antibody alone as control. Isolated cells and free cell borders of contacting cells did not localize polycystin-1 at the membrane, although some intracellular staining can be seen. These data suggest that the compartmentalization of polycystin-1 is dynamic and that trafficking of polycystin-1 between the cytoplasm and plasma membrane compartments is a function of cell contact.

The polycystic kidney disease 1 (PKD1) gene encodes a novel protein with multiple cell recognition domains. Hughes J. et al. (1995) Cell **10**:151-159. The analysis of the three-dimensional structure of a single repeat showed that it is not a true member of Ig superfamily, although it has a characteristic ß-sandwich topology. Bycroft M. et al. (1999) EMBO J. **18**:297-305. Domains with this Ig-like fold are present in proteins as diverse as matrix proteins, receptors and enzymes, and in each case they have been shown to interact with extremely different ligands varying from small peptides (e.g., HLA) to giant proteins (e.g., titin oligomer). Bork A. et al. (1994) J. Mol. Biol. **242**:309-320.

Using antibodies against three different regions of polycystin-1: N-terminal (LRR), C-terminal, and the middle region (REJ), the experiments described herein clearly showed that polycystin-1 was predominantly expressed at sites of cell-cell contact in kidney epithelial cells, as was the case for endothelial cells. The homophilic binding potential of several Ig-like domains, i.e., Ig^a, Ig^b and Ig^c, containing 4, 5 and 6 domains, as clusters were analyzed as described below. Each region was translated *in vitro* and tested for the ability to bind to each region including itself in the form of immobilized fusion protein. The binding properties of all combinations were quantitatively analyzed as a percentage of binding of *in vitro* translated protein. In this type of assay the fusion proteins are present in a vast excess compared to the amount of the translated probe. Therefore, theoretically almost all of the translated probe should bind to immobilized fusion protein, even if binding is weak. Phizicky, E.M. & Fields, S. (1995) Microbiological Reviews **59**:94-123. In practice, deviations from quantitative binding occur if not all of the immobilized protein or/and *in vitro* translated probe is functionally active. Nevertheless, a functionally relevant interaction should result in significant retention of ligand. For example, estimates from affinity chromatography binding experiments on the N-NusA, NusA-RNA polymerase and RAP30/74-RNA polymerase II interactions indicate that at least 50% of these proteins are available for binding. Formoza, T. et al. (1991) Meth. Enzymol. **208**:24-45.

Strong homophilic interactions were detected between the Ig-like domains, which are calcium independent. The strongest interaction was detected for the combination Ig^c-Ig^c, where the bound fraction constituted up to 90%. The least efficient interaction, characterized by 20% binding was detected for the Ig^a-Ig^a and Ig^a-Ig^b combinations. Ig^b-Ig^b, Ig^b-Ig^c combinations demonstrated intermediate binding ranging from 25-45%. The observed difference in binding capacities could be due to the different number of Ig-domains in each construct, so that the higher number of repeats results in stronger binding because of higher avidity. It could also be due to the cooperative nature of this interaction. The homophilic

binding of polycystin-1 resembles that of chick NCAM where all of the five Ig-like domains are involved in homophilic interactions. Ranheim T.S. et al. (1996) Proc. Natl. Acad. Sci. USA **93**:4071-4075. It is possible that the homophilic interactions described in this study might mediate homodimerization in addition to homophilic adhesion at intercellular contacts. A similar mechanism was shown to be important in the functioning of the PECAM-1 protein and modulating its ligand binding state (homophilic or heterophilic). Sun J. et al. (1996) J. Biol. Chem. **271**:18561-18570. In addition, homotypic binding between the extracellular domains of cadherins mediates formation of complexes between parallel-oriented molecules on single cells and between cells, which is thought to cooperatively enhance adhesion. Brieher W.M. et al. (1996) J. Cell Biol. **135**:487-496. Similarly, the data shown herein suggest that cis interactions between polycystin-1 molecules, mediating homodimerization on the same membrane might coexist with trans-interactions between opposing molecules at the site of cell-cell contact.

To adequately assess the significance of the Ig-like domain homophilic interactions under consideration, they were compared them side by side with known interactions. One of those was the interaction between p53 and SV40 large T-antigen, which is known to be functionally significant. Lane D.P. et al. (1979) Nature **278**:261-262 and Iwabuchi K. et al. (1993) Oncogene **8**:1693-1696. The bound fraction of T-antigen comprised approximately 45% of the total probe in this system. The interaction between the PKD1 and PKD2 gene products also was used as a reference. Quian F. et al. (1997) Nature Genetics **16**:179-183 and Tsiokas L. et al. (1997) Proc. Natl. Acad. Sci. USA **94**:6965-6970. This interaction was initially identified by the two-hybrid assay and was further characterized using the *in vitro* binding assay. Approximately 1.5% of the input polycystin-1 probe bound to immobilized polycystin-2, while 6% of the labeled ligand was bound in the reverse combination. Quian F. et al. (1997) Nature Genetics **16**:179-183. Similarly, a weak PKD2-PKD1 gene product interaction was detected which never exceeded ~1% of binding in different buffer compositions. Thus, the strength of the homophilic interactions between the

various Ig-like regions of polycystin-1 as measured *in vitro* is more comparable to the known functionally significant p53-T antigen binding rather than to the weaker and likely transient interaction between polycystin-1 and -2.

The importance of this biochemical binding assay results was tested *in vivo* by assessing the effect of soluble Ig-like domains on cell adhesion using both cell monolayers and cells in suspension. It was shown that soluble Ig-like domains perturb *in vivo* intercellular adhesion in MDCK cell monolayers, suggesting that they are directly involved in intercellular adhesion. It was likewise shown that soluble Ig-like domains can interfere with cellular adhesion using a cell aggregation assay.

The formation and progression of ADPKD cysts is characterized by increased cell proliferation, resulting in expansion of the epithelium, which displays a relatively undifferentiated appearance. Grantham J. (1996) Amer. J. Kidney Diseases **28**:788-803 and Avner E.D. (1993) J. Cell Sci. **17**:217-222. The role of polycystin-1 in mediating cell-cell interactions, where such interactions are fundamental for cellular functions of proliferation, differentiation and maturation, is supported by a recent study of a targeted PKD1 mutation in mice. Lu W. et al. (1997) Nature Genetics **17**:179-181. This study demonstrates that polycystin-1 is critical in the establishment and maturation of normal tubular architecture. Lu W. et al. (1997), *supra*. It has been shown that the expression of polycystin-1 is continued into adult life at a lower level, where its functional activity might be required for cells to remain tightly associated in the epithelium. Peters D.J.M. et al. (1996) Laboratory Investigation **75**:221-230; Ibraghimov-Beskrovnaya O. et al. (1997) Proc. Natl. Acad. Sci. USA **94**:6397-640; Weston B.S. et al. (1997) Histochemical Journal **29**:847-856 (1997); and Ward C.J. et al. (1996) Proc. Natl. Acad. Sci. USA **93**:1524-1528. In addition, it is known that cell adhesion proteins play an important role in intercellular signaling. Gumbiner B.M. (1996) Cell **84**:345-357. The results presented herein show that the loss of intercellular interactions due to a mutated polycystin-1 can be an important step in molecular cystogenesis.

Thus, in view of the above, this invention provides a method for modulating cell-cell adhesion in a suitable tissue, comprising delivering to the tissue an effective amount of an agent that modulates the binding of polycystin in the tissue. In one aspect, the modulation of cell-cell or cell-matrix adhesion is a reduction of cell-cell or cell-matrix adhesion. In another aspect, the modulation of cell-cell or cell-matrix adhesion is an increase or to enhance cell-cell or cell-matrix adhesion mediated by polycystin in a suitable tissue. As used herein, a "suitable tissue" includes any tissue which polycystin, i.e., polycystin-1 or polycystin-2, is expressed as been described above.

In one aspect, the agent is any agent that inhibits polycystin-1 mediated cell-cell or cell-matrix adhesion. Such agents include, but are not limited to, agents such as the antibodies described herein that bind to the Ig-like domains of polycystin, polycystin fragments comprising the Ig-like domains and agents that inhibit the expression of polycystin, e.g., polycystin-1 or polycystin-2, in a cell. Such agents include, but are not limited to antisense polycystin DNA and ribozymes that specifically recognize or cleave polycystin RNA in a cell.

One of skill in the art is enabled to make and use the agents noted above using the methods and compositions described herein alone or in combination with the methods known to those of skill in the art.

Alternatively, this invention also provides methods to promote cell-cell or cell-matrix adhesion in a tissue by delivering to the cell or tissue an effective amount of polycystin-1 to the cell or a polypeptide comprising an Ig-like domain of polycystin to the cell or tissue. The polycystin is delivered in the form of a polynucleotide or polypeptide or protein. In addition, one can restore normal cell-cell or cell-matrix adhesion is a tissue containing soluble, mutated polycystin by removing or binding the mutated polycystin using the anti-polycystin antibodies described herein as well as those known in the art.

The methods of this invention can be practiced *in vitro*, *in vivo* or *ex vivo*. When practiced *in vitro*, the methods provides screens for therapeutic agents that augment or inhibit the biological activity of wild-type or mutated polycystin in a

cell or tissue. To practice the screen, suitable cell cultures or tissue cultures are first provided. The cell can be a cultured cell or a genetically modified cell in which wild-type or mutated polycystin transmembrane regions are expressed on the cell surface. Alternatively, the cells can be from a tissue biopsy. The cells are cultured under conditions (temperature, growth or culture medium and gas (CO_2)) and for an appropriate amount of time to attain exponential proliferation without density dependent constraints. It also is desirable to maintain an additional separate cell culture; one which does not receive the agent being tested as a control.

As is apparent to one of skill in the art, suitable cells may be cultured in microtiter plates and several agents may be assayed at the same time by noting genotypic changes or phenotypic changes.

When the agent is a composition other than a DNA or RNA nucleic acid molecule, the suitable conditions may be by directly added to the cell culture or added to culture medium for addition. As is apparent to those skilled in the art, an "effective" amount must be added which can be empirically determined.

For the purposes of this invention, an "agent" is intended to include, but not be limited to a biological or chemical compound such as a simple or complex organic or inorganic molecule, a peptide, a protein (e.g. antibody) or an oligonucleotide (e.g. anti-sense). A vast array of compounds can be synthesized, for example oligomers, such as oligopeptides and oligonucleotides, and synthetic organic compounds based on various core structures, and these are also included in the term "agent". In addition, various natural sources can provide compounds for screening, such as plant or animal extracts, and the like. It should be understood, although not always explicitly stated that the agent is used alone or in combination with another agent, having the same or different biological activity as the agents identified by the inventive screen. The agents and methods also are intended to be combined with other therapies.

When the agent is a nucleic acid, it can be added to the cell cultures by methods well known in the art, which includes, but is not limited to calcium

phosphate precipitation, microinjection or electroporation. Alternatively or additionally, the nucleic acid can be incorporated into an expression or insertion vector for incorporation into the cells. Vectors that contain both a promoter and a cloning site into which a polynucleotide can be operatively linked are well known in the art. Such vectors are capable of transcribing RNA *in vitro* or *in vivo*, and are commercially available from sources such as Stratagene (La Jolla, CA) and Promega Biotech (Madison, WI). In order to optimize expression and/or *in vitro* transcription, it may be necessary to remove, add or alter 5' and/or 3' untranslated portions of the clones to eliminate extra, potential inappropriate alternative translation initiation codons or other sequences that may interfere with or reduce expression, either at the level of transcription or translation. Alternatively, consensus ribosome binding sites can be inserted immediately 5' of the start codon to enhance expression. Examples of vectors are viruses, such as baculovirus and retrovirus, bacteriophage, adenovirus, adeno-associated virus, cosmid, plasmid, fungal vectors and other recombination vehicles typically used in the art which have been described for expression in a variety of eukaryotic and prokaryotic hosts, and may be used for gene therapy as well as for simple protein expression.

One can determine if the object of the method, *i.e.*, modulation of cell-cell or cell-matrix adhesion has been achieved by noting phenotypic change in the cell as described below or by alteration of transcript expression. Kits containing the agents and instructions necessary to perform the screen and *in vitro* method as described herein also are claimed.

When the subject is an animal such as a rat or mouse, the method provides a convenient animal model system which can be used prior to clinical testing of the therapeutic agent. It also can be useful to have a separate negative control group of cells or animals which are healthy and not treated, which provides a basis for comparison.

These agents of this invention and the above noted compounds and their derivatives may be used for the preparation of medicaments for use in the methods described herein.

In a preferred embodiment, an agent of the invention is administered to treat a pathology associated with abnormal polycystin expression such as PKD. Various delivery systems are known and can be used to administer a therapeutic agent of the invention, *e.g.*, encapsulation in liposomes, microparticles, microcapsules, expression by recombinant cells, receptor-mediated endocytosis (see, *e.g.*, Wu and Wu (1987) J. Biol. Chem. **262**:4429-4432), construction of a therapeutic nucleic acid as part of a retroviral or other vector, etc. Methods of delivery include but are not limited to intra-arterial, intra-muscular, intravenous, intranasal, and oral routes. In a specific embodiment, it may be desirable to administer the pharmaceutical compositions of the invention locally to the area in need of treatment; this may be achieved by, for example, and not by way of limitation, local infusion during surgery, by injection, or by means of a catheter.

The agents identified herein as effective for their intended purpose can be administered to subjects or individuals susceptible to or at risk of developing a disease associated with abnormal polycystin expression such as PKD. When the agent is administered to a subject such as a mouse, a rat or a human patient, the agent can be added to a pharmaceutically acceptable carrier and systemically or topically administered to the subject. Therapeutic amounts can be empirically determined and will vary with the pathology being treated, the subject being treated and the efficacy and toxicity of the agent.

Administration *in vivo* can be effected in one dose, continuously or intermittently throughout the course of treatment. Methods of determining the most effective means and dosage of administration are well known to those of skill in the art and will vary with the composition used for therapy, the purpose of the therapy, the target cell being treated, and the subject being treated. Single or multiple administrations can be carried out with the dose level and pattern being

selected by the treating physician. Suitable dosage formulations and methods of administering the agents can be found below.

An agent of the present invention also referred to herein as the active ingredient, may be administered for therapy by any suitable route including oral, rectal, nasal, topical (including transdermal, aerosol, buccal and sublingual), vaginal, parental (including subcutaneous, intramuscular, intravenous and intradermal) and pulmonary. It will also be appreciated that the preferred route will vary with the condition and age of the recipient, and the disease being treated.

Ideally, the agent should be administered to achieve peak concentrations of the active compound at sites of disease. This may be achieved, for example, by the intravenous injection of the agent, optionally in saline, or orally administered, for example, as a tablet, capsule or syrup containing the active ingredient. Desirable blood levels of the agent may be maintained by a continuous infusion to provide a therapeutic amount of the active ingredient within disease tissue. The use of operative combinations is contemplated to provide therapeutic combinations requiring a lower total dosage of each component antiviral agent than may be required when each individual therapeutic compound or drug is used alone, thereby reducing adverse effects.

It should be understood that in addition to the ingredients particularly mentioned above, the formulations of this invention may include other agents conventional in the art having regard to the type of formulation in question, for example, those suitable for oral administration may include such further agents as sweeteners, thickeners and flavoring agents. It also is intended that the agents, compositions and methods of this invention be combined with other suitable compositions and therapies.

The following examples are intended to illustrate, but not limit this invention.

EXAMPLES

EXPERIMENT NO 1 - PRODUCTION OF ANTI-POLYCYSTIN ANTIBODIES

Example 1: Production and characterization of polyclonal antibodies raised against the transmembrane domain of polycystin

A panel of seven GST-fusion proteins containing sequences corresponding to a specific loop region (see Figure 2) and one MBP-fusion protein comprising sequences outside the loop region of the polycystin transmembrane domain were expressed in *E. coli* and used to immunize rabbits. The production and characterization of the anti-loop 4 antibodies were detailed below.

A fragment of polycystin cDNA corresponding to amino acids 3364-3578 was cloned into pGEX vector (Pharmacia) for production of FP-L4 fusion protein *E. coli* (Figure 2). *E. coli* DH5 alpha cells carrying this construct were grown overnight, diluted 1:10 and induced with 0.1 mM IPTG for 3 hours. Fusion protein was isolated as suggested by the manufacturer (Pharmacia) and injected into two rabbits for production of polyclonal antisera. Antibodies were shown to specifically recognize corresponding immunogen (FP-L4) on western blot. In addition, produced anti-FP-L4 antibodies specifically recognized truncated polycystin, expressed in baculovirus/insect system.

Example 2: Fractionation of tissue homogenates

To separate the particulate fractions (or crude membranes) from the cytosolic fractions, tissues were homogenized in 7 volume of homogenization buffer containing 10 mM HEPES, pH 7.4, 0.25 M sucrose, 0.5 mM $MgCl_2$, 0.1 mM PMSF, 0.75 mM benzamidine, 1 μg/ml aprotinin, 1 μg/ml leupeptin, and 1 μg/ml pepstatin. The homogenates were then centrifuged at 1,100 x g for 15 min at 4°C, and the supernatant was filtered through cheesecloth. Total tissue membranes were pelleted by centrifugation at 140,000 x g for 1 hour at 4°C and the supernatants were collected as the cytosolic fractions.

The fractionation of subcellular structures was carried out by differential centrifugation. Homogenates prepared as described above were first centrifuged

at 600 x g for 10 min at 4°C. The resulting supernatant $S600_I$ was collected, and the pellet $P600_I$ was resuspended in homogenization buffer and then centrifuged under the same condition to yield the supernatant $S600_{II}$ and the pellet $P600_{II}$ fractions. Fraction $S600_I$ containing the cytosolic contents as well as fraction $S600_{II}$ containing the membrane structures of the cells were then combined and subjected to high speed centrifugation at 150,000 x g for 10 min at 4°C. The resulting pellet, P15K, containing large organelles including mitochondria and lysosomes were collected, and the supernatant S15K was further fractionated at 150,000 x g for 60 min at 4°C to yield fraction S150K and P150K. Whereas S150K contains cytosolic components, P150K contains low density membrane structures such as plasma membrane, Endoplasmic reticulum and Golgi apparatus. The presence of a polycystin-related protein in various cell fractions was then determined by immunoassays employing one or more of the antibodies described herein. A polycystin-related protein having a molecular weight higher then 200 kD was predominantly detected in the membrane fractions P15K and P150K and not in the cytosolic fraction S150K of both the kidney and liver homogenates. This suggests that the polycystin-related protein expressed in these two tissues is associated with one or more cellular membrane structures, including plasma membrane, mitochondria, lysosomes, Endoplasmic reticulum and Golgi apparatus. Fractionation of fetal brain tissues, however, revealed that a polycystin-related protein having a lower molecular weight than the one expressed in the kidney and liver was associated with both the cytosolic fraction (S150K) and the microsomal fraction (P150K).

To further investigate the possibility that the polycystin-related protein expressed in the kidney is an integral membrane protein, membrane fractions was subjected to a "high salt" wash using, e.g., 0.3 M potassium chloride. The membrane bound polycystin-related protein was resistant to "high salt" washing. No polycystin-related protein expressed in the kidneys was dislodged from the membrane and released to the supernatant fraction (S150K KCl) after high speed centrifugation. This result suggests that the polycystin-related protein expressed

in the kidneys is tightly bound to the cellular membranes, and likely to be an integral membrane protein.

Example 3: Gel electrophoresis and immunoblotting

Proteins of each tissue fraction were separated on 3-12% gradient SDS polyacrylamide gels. Transfer of proteins to nitrocellulose was performed by electroblotting. For immunoblotting membranes were pre-blocked in Blotto (5% nonfat dry milk in PBS, pH 7.4) for 1 hour, then incubated overnight with 1:1 00 diluted anti-FP-L4 antibodies. After washing membranes three times for 10 min in Blotto, immunoblots were incubated with 1:1000 diluted peroxidase-conjugated goat anti-rabbit IgG for 1 hour, washed and developed by ECL. A protein band of ~ 800 kD was detected in the membrane fractions of kidney and liver tissues. Similar ~ 800 kD band was also detected in a number of cell lines (see Figure 10D). Another protein band of ~ 600 kD was detected in the membrane and cytosolic fractions of the fetal brain homogenates.

Example 4: Polycystin expression in baculovirus/insect system and in COS cells.

Nhe-delta mutant deleted with amino acids 290-2960 (Figure 3) was generated for expression in baculovirus/insect system. Polycystin cDNA was cloned into pBacPAK9 transfer vector (Clontech). Insect cells Sf21 were cotransfected with transfer-polycystin plasmid and viral DNA and incubated for 72 hr. Several individual recombinant virus plaques were analyzed for recombinant protein production. Total cell lysates infected with individual plaques were separated by SDS-PAGE and analyzed by immunoblotting with anti-Loop4 antibodies. Expected immunoreactive band of ~ 170 kD, corresponding to the truncated polycystin was detected (see Figure 7).

Another deletion mutant (HTM3) containing the C-terminal portion of polycystin that encompasses most of the transmembrane domain and the entire intracellular domain was cloned into an expression vector. Transient expression

of the truncated polycystin was detected by immunoblotting cell lysates obtained from the COS1 cells transfected with the vector (Figures 8-9). No expression of the recombinant protein was found in the COS1 cells transfected with a control vector.

EXPERIMENT NO 2: CELL-CELL/CELL-MATRIX ADHESION

Example 5: Anti-polycystin-1 antibodies preparation

All antibodies were raised in rabbits against fusion proteins representing different domains of polycystin-1. Anti-LRR (Res. 27-360) and anti-BD3 (Res. 4097-4302) were affinity purified as described. Anti-L2 antibody was produced against GST fusion protein containing part of REJ domain of polycystin-1 (Res. 2714-3074).

Example 6: Expression of recombinant polycystin-1 in baculovirus/insect cell systems

Truncated polycystin-1 was expressed by using BacPAK TM Baculovirus Expression System (Clontech) according to the manufacturer's instructions. Briefly, PKD1 cDNA inserts HTM3 and Nhe delta were subcloned into pBacPAK9 transfer vector and co-transfected with BacPAK6 viral DNA into Sf21 insect cells. Individual plaques from the supernatant co-transfection medium were analyzed and selected for the high level of polycystin-1 protein production as assayed by Western blotting.

Example 7: Immunofluoresence

MDCK cells (source) or baculovirus infected Sf21 cells were grown on glass coverslips and immunostained as described in Ibraghimov-Beskrovnaya, O. et al. (1997) Proc. Natl. Acad. Sci. 94:6397-6402. The primary antibodies were used at a dilution of 1:100 followed by incubation with FITC labeled goat anti-rabbit secondary antibody at a dilution 1:200. Cells were examined using a Zeiss Axioplan microscope.

Example 8: Production of fusion proteins for *in vitro* binding assay

The cluster of Ig-like domains of polycystin-1 was subdivided into three constructs: Ig^a (domains II-V (amino acids 843-1200)), Ig^b (domains VI-X (amino acids 1205-1625)) and Ig^c (domains XI-XVI (amino acids 1626-2136)) and subcloned into pGEX-1 vector (Pharmacia) for production of GST fusion proteins designated GST-Ig^a, GST-Ig^b and GST-Ig^c, respectively. The cDNA fragments for each construct were synthesized by PCR using as template the full-length human PKD1 cDNA described previously in Ibraghimov-Beskrovnaya O. et al. (1997) Proc. Natl. Acad. Sci. 94:6397-6402. The C-terminal region of polycystin-1 (MBP-PKD1) (Res. 4077-4302) was constructed as an MBP fusion protein by cloning in the expression vector pMALc2 (NEB). The GST-p53 construct (Res.73-390) was produced as GST-fusion protein. The GST fusion proteins were purified from supernatants by affinity chromatography on Glutathione-Sepharose (Pharmacia) as recommended by the manufacturer.

Experiment 9: *In vitro* translation probes

Translation of the PKD1 constructs *in vitro* was performed using the TNT Coupled Reticulocyte Lysate System (Promega) as recommended by the manufacturer. The Ig-like domains of polycystin-1: Ig^a (domains II-V), Ig^b (domains VI-X) and Ig^c (domains XI-XVI) were subcloned downstream of the oligo **GTAATACGACTCACTATAGGG**CGAGCCACCATGG (SEQ ID NO:3), containing the T7 RNA polymerase promoter (bold) followed by an AUG initiation codon in a Kozak consensus context (underlined). This oligo was inserted between the BamHI and EcoRI sites of the pGEX-4T-1 vector (Pharmacia) downstream of GST coding region, such that the same construct can be used for either GST fusion protein production or for the *in vitro* translation of the insert without the GST portion. ^{35}S-PKD2 probe (Res. 657-968) and ^{35}S-T-antigen probe (res. 87-708) were generated in the same manner.

GST-fusion proteins or GST alone were immobilized individually onto Glutathione Sepharose (Pharmacia). MBP-PKD1 fusion protein or MBP-lacZ as

control were immobilized onto amylose resin (NEB). Twenty (20) μL of beads with ~10 μg of immobilized fusion proteins were used for each binding reaction. Approximately 10 μl of *in vitro* translated ^{35}S-labeled probe were incubated for 3hours at room temperature with immobilized fusion proteins in 0.1 ml of binding buffer (10 mM HEPES, pH 7.4,100 mM NaCl, 1 mM $CaCl_2$, 2 mM $MgCl_2$, 0.75 mM benzamidine, 0.1 mM PMSF) and washed with 20 column volumes of the same buffer. The polycystin-2 and polycystin-1 interaction assay was also performed in another buffer (10 mM Tris, pH 7.4, 200 mM NaCl,1 mM EDTA). The ^{35}S-translated material bound to the beads was resolved by SDS-PAGE with input ^{35}S probe run in parallel. The gels were exposed to film (X-Omat AR, Kodak) as well as quantified using a PhosphorImager with ImageQuant (v. 3.2) software (Molecular Dynamics). Only bands representing the full-length product of *in vitro* translation were used for quantification in each binding reaction and bound fractions were estimated as percentage of input of ^{35}S translated probe.

SDS-PAGE was carried out on 3-12% or 5-15% gradient gels in the presence of 1% 2-mercaptoethanol and transferred to nitrocellulose for immunoblot analysis as described 43 Primary anti-polycystin-1 antibodies were used at a dilution 1:100 and secondary goat anti-rabbit-HRP antibodies (Boehringer Mannheim) were used at a dilution 1:1000.

Experiment 10: Disruption of cell-cell adhesion in cell monolayers and aggregation assay

The disruption of intercellular adhesion was performed by the method of Wheelock et al. (1987) J. Cell Biochem. **34**:187-202. MDCK cells were grown 24 hours to 70% confluency in media with 10% fetal bovine serum. The complete media was replaced with control serum-free media alone or with media containing either GST carrier protein or GST-Ig^a, GST-Ig^b and GST-Ig^c fusion proteins (1 nM each) as described above. Cells were incubated for 30 hours and live cell images were collected using a Nikon Eclipse 200 microscope equipped with a

Sony CCD/RGB camera DXC-151 and Scionimage 1.62a software (Scion Corporation).

The aggregation assay was performed as described in DeLisser et al. (1993) J. Biol. Chem. **268**:16037-16046, with minor modifications. Briefly, MDCK cells were plated at $5x10^6$ cells/10cm plate and grown for 24 hours. Cells were harvested by incubation in PBS with 10 mM EDTA for 15 min followed by incubation with 0.01% trypsin for 2 min. After washing the cells were resuspended at ~$1x10^6$/ml in serum free media alone or media with GST protein or with GST-Ig^a, GST-Ig^b and GST-Ig^c at a concentration of 7 nM each. Cells were transferred to a 24-well plastic tray, previously blocked with 3% BSA in PBS and rotated at 100 rpm at 37°C for 1.5 hour and images of live cells were collected as described above.

While the invention has been described in detail herein and with reference to specific embodiments thereof, it will be apparent to one skilled in the art that various changes and modifications can be made to the invention as described above without departing from the spirit and scope thereof.

CLAIMS

1. An isolated antibody or a fragment thereof that binds to an epitope present in the transmembrane domain of polycystin and specifically recognizes a polycystin-related polypeptide having an apparent molecular weight in the range of about 600 to about 800 kD.

2. An isolated antibody of claim 1, wherein the polypeptide has an apparent molecular weight of about 600 kD.

3. An isolated antibody of claim 1, wherein the polypeptide has an apparent molecular weight of about 800 kD.

4. An isolated antibody comprising an epitope, wherein the epitope comprises a peptide having amino acids as shown in Figure 1 (SEQ ID NO:2) selected from the group consisting of amino acid residues 2621 to 2710, amino acid residues 2734 to 3094, amino acid residues 3116 to 3300, amino acid residues 3364 to 3578, amino acid residues 3623 to 3688, amino acid residues 3710 to 3914, amino acid residues 3931 to 4046, amino acid residues 2166 to 2599, amino acid residues 4097 to 4302, amino acid residues 4148 to 4219, amino acid residues 4220 to 4302,amino acid residues 27 to 360, amino acid residues 843 to 1200, amino acid residues 1205 to 1625, and amino acid residues 1626 to 2136.

5. An isolated antibody or a fragment thereof that specifically binds to the transmembrane domain of an integral membrane protein that is associated with polycystic kidney disease, wherein the integral membrane protein also binds to a reference antibody selected from the group consisting of anti-FP-L1, anti-FP-L2, anti-FP-L3, anti-FP-L4, anti-FP-L5, anti-FP-L6, anti-FP-L7, anti-MAL-REJ antibody, anti-MAL-BD3 antibody, anti-FP-46-2 antibody, anti-FP-46-1c antibody, or anti-FP-LRR antibody.

6. An isolated antibody of any of claims 1 to 5, wherein the antibody is a polyclonal antibody.

7. An isolated antibody of any of claims 1 to 5, wherein the antibody is a monoclonal antibody.

8. An isolated antibody of any of claims 1 to 5 labeled with a detectable label.

9. A composition comprising a carrier and an antibody of any of claims 1 to 5.

10. A hybridoma cell line that produces the monoclonal antibody of claim 7.

11. An isolated antibody of any of claims 1 to 5, wherein the polypeptide or protein is expressed in a tissue selected from the group consisting of kidney, brain, liver and neuronal tissues.

12. A recombinant polypeptide comprising a polypeptide fragment of polycystin, wherein the fragment is a membrane-spanning segment of polycystin selected from the group consisting of loop 1, loop 2, loop 3, loop 4 and loop 7.

13. A recombinant polypeptide comprising a polypeptide fragment of polycystin, wherein the fragment comprises a peptide having amino acids as shown in Figure 1 (SEQ ID NO:2) selected from the group consisting of amino acid residues 2621 to 2710, amino acid residues 2734 to 3094, amino acid residues 3116 to 3300, amino acid residues 3364 to 3578, amino acid residues 3623 to 3688, amino acid residues 3710 to 3914, amino acid residues 3931 to

4046, amino acid residues 2166 to 2599, amino acid residues 4097 to 4302, amino acid residues 4148 to 4219, amino acid residues 4220 to 4302, amino acid residues 27 to 360, amino acid residues 843 to 1200, amino acid residues 1205 to 1625, and amino acid residues 1626 to 2136.

15. A composition comprising a carrier and a polypeptide of claim 13.

16. An isolated polynucleotide encoding the recombinant polypeptide of claim 13.

17. A gene delivery vehicle comprising the polynucleotide of claim 16.

18. A host cell transformed with the isolated polynucleotide of claim 16.

19. An isolated polypeptide having an apparent molecular weight in the range of about 600 to about 800 kD that specifically binds to an antibody or fragment thereof of claim 1.

20. An isolated polypeptide of claim 19, wherein the polypeptide has an apparent molecular weight of about 600 kD.

21. The isolated polypeptide of claim 19, wherein the polypeptide has an apparent molecular weight of about 800 kD.

22. A diagnostic kit for detecting a polycystin-related polypeptide present in a sample, comprising an antibody of any of claims 1 to 5, and instructions for the use of the antibody to detect the polypeptide.

23. A method for modulating cell-cell adhesion in a suitable tissue, comprising delivering to the tissue an effective amount of an agent that modulates the binding of polycystin in the tissue.

24. The method of claim 22, wherein the modulation of cell-cell or cell-matrix adhesion is a reduction of cell-cell or cell-matrix adhesion.

25. The method of claim 24, wherein the agent prevents or inhibits transcription and/or translation of a polycystin polypeptide in a cell.

26. The method of claim 24, wherein the agent is an antisense polynucleotide to an isolated polynucleotide of claim 16.

27. The method of claim 24, wherein the agent is a ribozyme that inhibits translation of an isolated polynucleotide of claim 16.

28. The method of claim 22, wherein the modulation of cell-cell or cell-matrix adhesion is promotion or enhancement of cell-cell or cell-matrix adhesion in a suitable cell or tissue.

29. The method of claim 28, wherein an effective amount of a polycystin Ig-like domain is delivered to the cell or tissue.

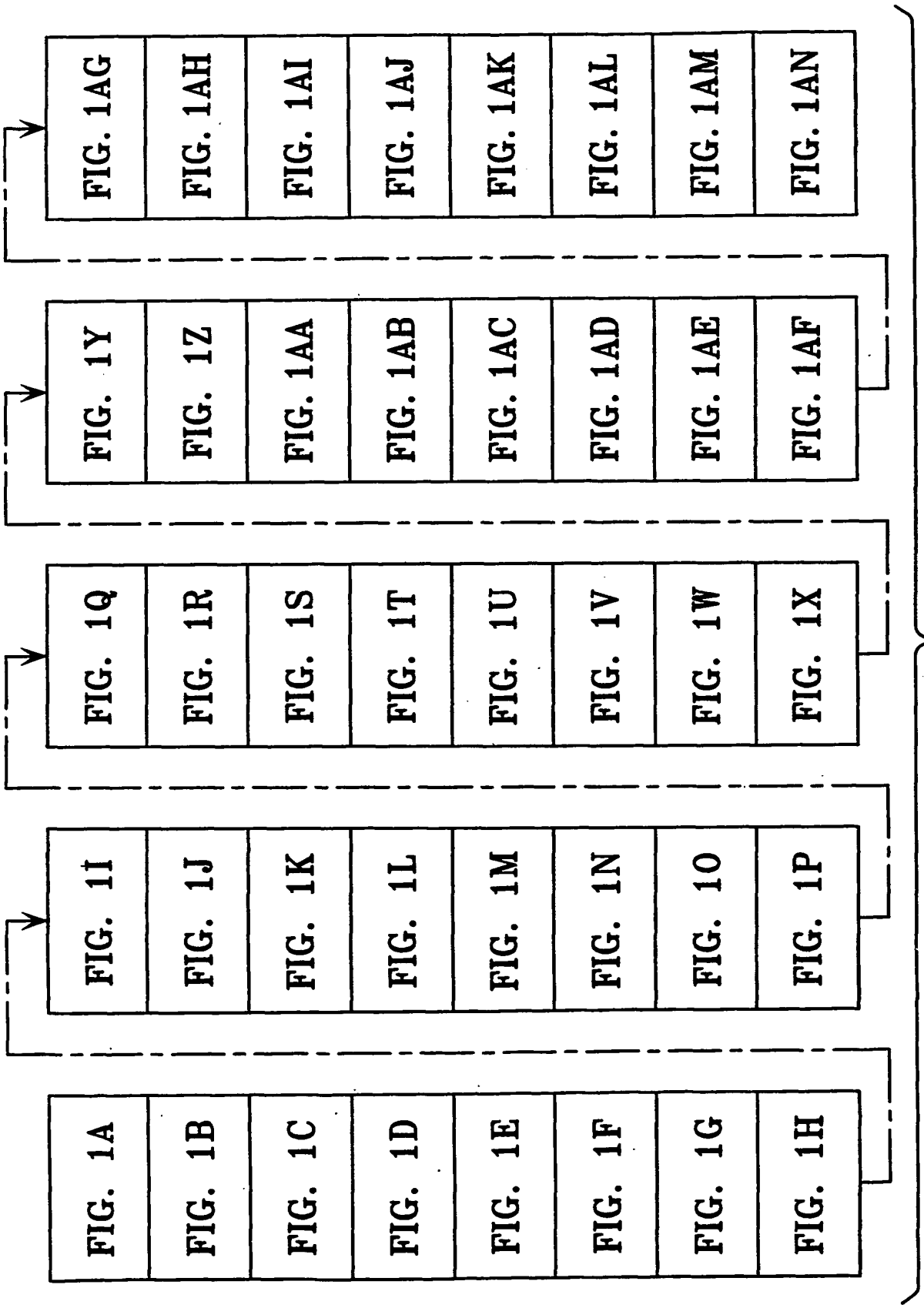


FIG. 1A
FIG. 1B
FIG. 1C
FIG. 1D
FIG. 1E
FIG. 1F
FIG. 1G
FIG. 1H
FIG. 1I
FIG. 1J
FIG. 1K
FIG. 1L
FIG. 1M
FIG. 1N
FIG. 1O
FIG. 1P
FIG. 1Q
FIG. 1R
FIG. 1S
FIG. 1T
FIG. 1U
FIG. 1V
FIG. 1W
FIG. 1X
FIG. 1Y
FIG. 1Z
FIG. 1AA
FIG. 1AB
FIG. 1AC
FIG. 1AD
FIG. 1AE
FIG. 1AF
FIG. 1AG
FIG. 1AH
FIG. 1AI
FIG. 1AJ
FIG. 1AK
FIG. 1AL
FIG. 1AM
FIG. 1AN

FIG. 1

```
GCTCAGCAGC AGGTCGCGGC CGCAGCCCCA TCCAGCCCGC GCCCGCCATG CCGTCCGCGG    60

GCCCCGCCTG AGCTGCGGCC TCCGCGCGCG GGCGGGCCTG GGGACGGCGG GGCCATGCGC   120

GCGCTGCCCT AACG ATG CCG CCC GCC GCG CCC GCC CGC CTG GCG CTG GCC      170
                Met Pro Pro Ala Ala Pro Ala Arg Leu Ala Leu Ala
                  1               5                  10

CTG GGC CTG GGC CTG TGG CTC GGG GCG CTG GCG GGG GGC CCC GGG CGC      218
Leu Gly Leu Gly Leu Trp Leu Gly Ala Leu Ala Gly Gly Pro Gly Arg
          15                  20                  25

GGC TGC GGG CCC TGC GAG CCC CCC TGC CTC TGC GGC CCA GCG CCC GGC      266
Gly Cys Gly Pro Cys Glu Pro Pro Cys Leu Cys Gly Pro Ala Pro Gly
      30                  35                  40

GCC GCC TGC CGC GTC AAC TGC TCG GGC CGC GGG CTG CGG ACG CTC GGT      314
Ala Ala Cys Arg Val Asn Cys Ser Gly Arg Gly Leu Arg Thr Leu Gly
  45                  50                  55                  60

CCC GCG CTG CGC ATC CCC GCG GAC GCC ACA GCG CTA GAC GTC TCC CAC      362
Pro Ala Leu Arg Ile Pro Ala Asp Ala Thr Ala Leu Asp Val Ser His
                  65                  70                  75

AAC CTG CTC CGG GCG CTG GAC GTT GGG CTC CTG GCG AAC CTC TCG GCG      410
Asn Leu Leu Arg Ala Leu Asp Val Gly Leu Leu Ala Asn Leu Ser Ala
              80                  85                  90
```

FIG. 1A

```
CTG GCA GAG CTG GAT ATA AGC AAC AAC AAG ATT TCT ACG TTA GAA GAA      458
Leu Ala Glu Leu Asp Ile Ser Asn Asn Lys Ile Ser Thr Leu Glu Glu
         95                 100                 105

GGA ATA TTT GCT AAT TTA TTT AAT TTA AGT GAA ATA AAC CTG AGT GGG      506
Gly Ile Phe Ala Asn Leu Phe Asn Leu Ser Glu Ile Asn Leu Ser Gly
    110                 115                 120

AAC CCG TTT GAG TGT GAC TGT GGC CTG GCG TGG CTG CCG CGA TGG GCG      554
Asn Pro Phe Glu Cys Asp Cys Gly Leu Ala Trp Leu Pro Arg Trp Ala
125                 130                 135                 140

GAG GAG CAG CAG GTG CGG GTG GTG CAG CCC GAG GCA GCC ACG TGT GCT      602
Glu Glu Gln Gln Val Arg Val Val Gln Pro Glu Ala Ala Thr Cys Ala
                145                 150                 155

GGG CCT GGC TCC CTG GCT GGC CAG CCT CTG CTT GGC ATC CCC TTG CTG      650
Gly Pro Gly Ser Leu Ala Gly Gln Pro Leu Leu Gly Ile Pro Leu Leu
            160                 165                 170

GAC AGT GGC TGT GGT GAG GAG TAT GTC GCC TGC CTC CCT GAC AAC AGC      698
Asp Ser Gly Cys Gly Glu Glu Tyr Val Ala Cys Leu Pro Asp Asn Ser
        175                 180                 185

TCA GGC ACC GTG GCA GCA GTG TCC TTT TCA GCT GCC CAC GAA GGC CTG      746
Ser Gly Thr Val Ala Ala Val Ser Phe Ser Ala Ala His Glu Gly Leu
    190                 195                 200
```

FIG. 1B

```
CTT CAG CCA GAG GCC TGC AGC GCC TTC TGC TTC TCC ACC GGC CAG GGC      794
Leu Gln Pro Glu Ala Cys Ser Ala Phe Cys Phe Ser Thr Gly Gln Gly
205                 210                 215                 220

CTC GCA GCC CTC TCG GAG CAG GGC TGG TGC CTG TGT GGG GCG GCC CAG      842
Leu Ala Ala Leu Ser Glu Gln Gly Trp Cys Leu Cys Gly Ala Ala Gln
                225                 230                 235

CCC TCC AGT GCC TCC TTT GCC TGC CTG TCC CTC TGC TCC GGC CCC CCG      890
Pro Ser Ser Ala Ser Phe Ala Cys Leu Ser Leu Cys Ser Gly Pro Pro
            240                 245                 250

CCA CCT CCT GCC CCC ACC TGT AGG GGC CCC ACC CTC CTC CAG CAC GTC      938
Pro Pro Pro Ala Pro Thr Cys Arg Gly Pro Thr Leu Leu Gln His Val
        255                 260                 265

TTC CCT GCC TCC CCA GGG GCC ACC CTG GTG GGG CCC CAC GGA CCT CTG      986
Phe Pro Ala Ser Pro Gly Ala Thr Leu Val Gly Pro His Gly Pro Leu
    270                 275                 280

GCC TCT GGC CAG CTA GCA GCC TTC CAC ATC GCT GCC CCG CTC CCT GTC     1034
Ala Ser Gly Gln Leu Ala Ala Phe His Ile Ala Ala Pro Leu Pro Val
285                 290                 295                 300

ACT GCC ACA CGC TGG GAC TTC GGA GAC GGC TCC GCC GAG GTG GAT GCC     1082
Thr Ala Thr Arg Trp Asp Phe Gly Asp Gly Ser Ala Glu Val Asp Ala
                305                 310                 315
```

FIG. 1C

```
GCT GGG CCG GCT GCC TCG CAT CGC TAT GTG CTG CCT GGG CGC TAT CAC    1130
Ala Gly Pro Ala Ala Ser His Arg Tyr Val Leu Pro Gly Arg Tyr His
            320                 325                 330

GTG ACG GCC GTG CTG GCC CTG GGG GCC GGC TCA GCC CTG CTG GGG ACA    1178
Val Thr Ala Val Leu Ala Leu Gly Ala Gly Ser Ala Leu Leu Gly Thr
        335                 340                 345

GAC GTG CAG GTG GAA GCG GCA CCT GCC GCC CTG GAG CTC GTG TGC CCG    1226
Asp Val Gln Val Glu Ala Ala Pro Ala Ala Leu Glu Leu Val Cys Pro
    350                 355                 360

TCC TCG GTG CAG AGT GAC GAG AGC CTC GAC CTC AGC ATC CAG AAC CGC    1274
Ser Ser Val Gln Ser Asp Glu Ser Leu Asp Leu Ser Ile Gln Asn Arg
365                 370                 375                 380

GGT GGT TCA GGC CTG GAG GCC GCC TAC AGC ATC GTG GCC CTG GGC GAG    1322
Gly Gly Ser Gly Leu Glu Ala Ala Tyr Ser Ile Val Ala Leu Gly Glu
                385                 390                 395

GAG CCG GCC CGA GCG GTG CAC CCG CTC TGC CCC TCG GAC ACG GAG ATC    1370
Glu Pro Ala Arg Ala Val His Pro Leu Cys Pro Ser Asp Thr Glu Ile
            400                 405                 410

TTC CCT GGC AAC GGG CAC TGC TAC CGC CTG GTG GTG GAG AAG GCG GCC    1418
Phe Pro Gly Asn Gly His Cys Tyr Arg Leu Val Val Glu Lys Ala Ala
        415                 420                 425
```

FIG. 1D

```
TGG CTG CAG GCG CAG GAG CAG TGT CAG GCC TGG GCC GGG GCC GCC CTG    1466
Trp Leu Gln Ala Gln Glu Gln Cys Gln Ala Trp Ala Gly Ala Ala Leu
    430                 435                 440

GCA ATG GTG GAC AGT CCC GCC GTG CAG CGC TTC CTG GTC TCC CGG GTC    1514
Ala Met Val Asp Ser Pro Ala Val Gln Arg Phe Leu Val Ser Arg Val
445                 450                 455                 460

ACC AGG TGC CTA GAC GTG TGG ATC GGC TTC TCG ACT GTG CAG GGG GTG    1562
Thr Arg Cys Leu Asp Val Trp Ile Gly Phe Ser Thr Val Gln Gly Val
                465                 470                 475

GAG GTG GGC CCA GCG CCG CAG GGC GAG GCC TTC AGC CTG GAG AGC TGC    1610
Glu Val Gly Pro Ala Pro Gln Gly Glu Ala Phe Ser Leu Glu Ser Cys
            480                 485                 490

CAG AAC TGG CTG CCC GGG GAG CCA CAC CCA GCC ACA GCC GAG CAC TGC    1658
Gln Asn Trp Leu Pro Gly Glu Pro His Pro Ala Thr Ala Glu His Cys
        495                 500                 505

GTC CGG CTC GGG CCC ACC GGG TGG TGT AAC ACC GAC CTG TGC TCA GCG    1706
Val Arg Leu Gly Pro Thr Gly Trp Cys Asn Thr Asp Leu Cys Ser Ala
    510                 515                 520

CCG CAC AGC TAC GTC TGC GAG CTG CAG CCC GGA GGC CCA GTG CAG GAT    1754
Pro His Ser Tyr Val Cys Glu Leu Gln Pro Gly Gly Pro Val Gln Asp
525                 530                 535                 540
```

FIG. 1E

```
GCC GAG AAC CTC CTC GTG GGA GCG CCC AGT GGG GAC CTG CAG GGA CCC     1802
Ala Glu Asn Leu Leu Val Gly Ala Pro Ser Gly Asp Leu Gln Gly Pro
                545                 550                 555

CTG ACG CCT CTG GCA CAG CAG GAC GGC CTC TCA GCC CCG CAC GAG CCC     1850
Leu Thr Pro Leu Ala Gln Gln Asp Gly Leu Ser Ala Pro His Glu Pro
            560                 565                 570

GTG GAG GTC ATG GTA TTC CCG GGC CTG CGT CTG AGC CGT GAA GCC TTC     1898
Val Glu Val Met Val Phe Pro Gly Leu Arg Leu Ser Arg Glu Ala Phe
        575                 580                 585

CTC ACC ACG GCC GAA TTT GGG ACC CAG GAG CTC CGG CGG CCC GCC CAG     1946
Leu Thr Thr Ala Glu Phe Gly Thr Gln Glu Leu Arg Arg Pro Ala Gln
    590                 595                 600

CTG CGG CTG CAG GTG TAC CGG CTC CTC AGC ACA GCA GGG ACC CCG GAG     1994
Leu Arg Leu Gln Val Tyr Arg Leu Leu Ser Thr Ala Gly Thr Pro Glu
605                 610                 615                 620

AAC GGC AGC GAG CCT GAG AGC AGG TCC CCG GAC AAC AGG ACC CAG CTG     2042
Asn Gly Ser Glu Pro Glu Ser Arg Ser Pro Asp Asn Arg Thr Gln Leu
                625                 630                 635

GCC CCC GCG TGC ATG CCA GGG GGA CGC TGG TGC CCT GGA GCC AAC ATC     2090
Ala Pro Ala Cys Met Pro Gly Gly Arg Trp Cys Pro Gly Ala Asn Ile
            640                 645                 650
```

FIG. 1F

```
TGC TTG CCG CTG GAC GCC TCC TGC CAC CCC CAG GCC TGC GCC AAT GGC     2138
Cys Leu Pro Leu Asp Ala Ser Cys His Pro Gln Ala Cys Ala Asn Gly
        655                 660                 665

TGC ACG TCA GGG CCA GGG CTA CCC GGG GCC CCC TAT GCG CTA TGG ACA     2186
Cys Thr Ser Gly Pro Gly Leu Pro Gly Ala Pro Tyr Ala Leu Trp Arg
    670                 675                 680

GAG TTC CTC TTC TCC GTT CCC GCG GGG CCC CCC GCG CAG TAC TCG GTC     2234
Glu Phe Leu Phe Ser Val Pro Ala Gly Pro Pro Ala Gln Tyr Ser Val
685                 690                 695                 700

ACC CTC CAC GGC CAG GAT GTC CTC ATG CTC CCT GGT GAC CTC GTT GGC     2282
Thr Leu His Gly Gln Asp Val Leu Met Leu Pro Gly Asp Leu Val Gly
                705                 710                 715

TTG CAG CAC GAC GCT GGC CCT GGC GCC CTC CTG CAC TGC TCG CCG GCT     2330
Leu Gln His Asp Ala Gly Pro Gly Ala Leu Leu His Cys Ser Pro Ala
            720                 725                 730

CCC GGC CAC CCT GGT CCC CGG GCC CCG TAC CTC TCC GCC AAC GCC TCG     2378
Pro Gly His Pro Gly Pro Arg Ala Pro Tyr Leu Ser Ala Asn Ala Ser
        735                 740                 745

TCA TGG CTG CCC CAC TTG CCA GCC CAG CTG GAG GGC ACT TGG GCC TGC     2426
Ser Trp Leu Pro His Leu Pro Ala Gln Leu Glu Gly Thr Trp Ala Cys
    750                 755                 760
```

FIG. 1G

```
CCT GCC TGT GCC CTG CGG CTG CTT GCA GCC ACG GAA CAG CTC ACC GTG     2474
Pro Ala Cys Ala Leu Arg Leu Leu Ala Ala Thr Glu Gln Leu Thr Val
765                 770                 775                 780

CTG CTG GGC TTG AGG CCC AAC CCT GGA CTG CGG CTG CCT GGG CGC TAT     2522
Leu Leu Gly Leu Arg Pro Asn Pro Gly Leu Arg Leu Pro Gly Arg Tyr
                785                 790                 795

GAG GTC CGG GCA GAG GTG GGC AAT GGC GTG TCC AGG CAC AAC CTC TCC     2570
Glu Val Arg Ala Glu Val Gly Asn Gly Val Ser Arg His Asn Leu Ser
            800                 805                 810

TGC AGC TTT GAC GTG GTC TCC CCA GTG GCT GGG CTG CGG GTC ATC TAC     2618
Cys Ser Phe Asp Val Val Ser Pro Val Ala Gly Leu Arg Val Ile Tyr
        815                 820                 825

CCT GCC CCC CGC GAC GGC CGC CTC TAC GTG CCC ACC AAC GGC TCA GCC     2666
Pro Ala Pro Arg Asp Gly Arg Leu Tyr Val Pro Thr Asn Gly Ser Ala
    830                 835                 840

TTG GTG CTC CAG GTG GAC TCT GGT GCC AAC GCC ACG GCC ACG GCT CGC     2714
Leu Val Leu Gln Val Asp Ser Gly Ala Asn Ala Thr Ala Thr Ala Arg
845                 850                 855                 860

TGG CCT GGG GGC AGT GTC AGC GCC CGC TTT GAG AAT GTC TGC CCT GCC     2762
Trp Pro Gly Gly Ser Val Ser Ala Arg Phe Glu Asn Val Cys Pro Ala
                865                 870                 875
```

FIG. 1H

```
CTG GTG GCC ACC TTC GTG CCC GGC TGC CCC TGG GAG ACC AAC GAT ACC    2810
Leu Val Ala Thr Phe Val Pro Gly Cys Pro Trp Glu Thr Asn Asp Thr
            880                 885                 890

CTG TTC TCA GTG GTA GCA CTG CCG TGG CTC AGT GAG GGG GAG CAC GTG    2858
Leu Phe Ser Val Val Ala Leu Pro Trp Leu Ser Glu Gly Glu His Val
        895                 900                 905

GTG GAC GTG GTG GTG GAA AAC AGC GCC AGC CGG GCC AAC CTC AGC CTG    2906
Val Asp Val Val Val Glu Asn Ser Ala Ser Arg Ala Asn Leu Ser Leu
    910                 915                 920

CGG GTG ACG GCG GAG GAG CCC ATC TGT GGC CTC CGC GCC ACG CCC AGC    2954
Arg Val Thr Ala Glu Glu Pro Ile Cys Gly Leu Arg Ala Thr Pro Ser
925                 930                 935                 940

CCC GAG GCC CGT GTA CTG CAG GGA GTC CTA GTG AGG TAC AGC CCC GTG    3002
Pro Glu Ala Arg Val Leu Gln Gly Val Leu Val Arg Tyr Ser Pro Val
                945                 950                 955

GTG GAG GCC GGC TCG GAC ATG GTC TTC CGG TGG ACC ATC AAC GAC AAG    3050
Val Glu Ala Gly Ser Asp Met Val Phe Arg Trp Thr Ile Asn Asp Lys
            960                 965                 970

CAG TCC CTG ACC TTC CAG AAC GTG GTC TTC AAT GTC ATT TAT CAG AGC    3098
Gln Ser Leu Thr Phe Gln Asn Val Val Phe Asn Val Ile Tyr Gln Ser
        975                 980                 985
```

FIG. 1I

```
GCG GCG GTC TTC AAG CTC TCA CTG ACG GCC TCC AAC CAC GTG AGC AAC    3146
Ala Ala Val Phe Lys Leu Ser Leu Thr Ala Ser Asn His Val Ser Asn
    990                 995                1000

GTC ACC GTG AAC TAC AAC GTA ACC GTG GAG CGG ATG AAC AGG ATG CAG    3194
Val Thr Val Asn Tyr Asn Val Thr Val Glu Arg Met Asn Arg Met Gln
1005                1010                1015                1020

GGT CTG CAG GTC TCC ACA GTG CCG GCC GTG CTG TCC CCC AAT GCC ACG    3242
Gly Leu Gln Val Ser Thr Val Pro Ala Val Leu Ser Pro Asn Ala Thr
                1025                1030                1035

CTA GCA CTG ACG GCG GGC GTG CTG GTG GAC TCG GCC GTG GAG GTG GCC    3290
Leu Ala Leu Thr Ala Gly Val Leu Val Asp Ser Ala Val Glu Val Ala
            1040                1045                1050

TTC CTG TGG ACC TTT GGG GAT GGG GAG CAG GCC CTC CAC CAG TTC CAG    3338
Phe Leu Trp Thr Phe Gly Asp Gly Glu Gln Ala Leu His Gln Phe Gln
        1055                1060                1065

CCT CCG TAC AAC GAG TCC TTC CCG GTT CCA GAC CCC TCG GTG GCC CAG    3386
Pro Pro Tyr Asn Glu Ser Phe Pro Val Pro Asp Pro Ser Val Ala Gln
    1070                1075                1080

GTG CTG GTG GAG CAC AAT GTC ATG CAC ACC TAC GCT GCC CCA GGT GAG    3434
Val Leu Val Glu His Asn Val Met His Thr Tyr Ala Ala Pro Gly Glu
1085                1090                1095                1100
```

FIG. 1J

```
TAC CTC CTG ACC GTG CTG GCA TCT AAT GCC TTC GAG AAC CTG ACG CAG     3482
Tyr Leu Leu Thr Val Leu Ala Ser Asn Ala Phe Glu Asn Leu Thr Gln
                1105                1110                1115

CAG GTG CCT GTG AGC GTG CGC GCC TCC CTG CCC TCC GTG GCT GTG GGT     3530
Gln Val Pro Val Ser Val Arg Ala Ser Leu Pro Ser Val Ala Val Gly
            1120                1125                1130

GTG AGT GAC GGC GTC CTG GTG GCC GGC CGG CCC GTC ACC TTC TAC CCG     3578
Val Ser Asp Gly Val Leu Val Ala Gly Arg Pro Val Thr Phe Tyr Pro
        1135                1140                1145

CAC CCG CTG CCC TCG CCT GGG GGT GTT CTT TAC ACG TGG GAC TTC GGG     3626
His Pro Leu Pro Ser Pro Gly Gly Val Leu Tyr Thr Trp Asp Phe Gly
    1150                1155                1160

GAC GGC TCC CCT GTC CTG ACC CAG AGC CAG CCG GCT GCC AAC CAC ACC     3674
Asp Gly Ser Pro Val Leu Thr Gln Ser Gln Pro Ala Ala Asn His Thr
1165                1170                1175                1180

TAT GCC TCG AGG GGC ACC TAC CAC GTG CGC CTG GAG GTC AAC AAC ACG     3722
Tyr Ala Ser Arg Gly Thr Tyr His Val Arg Leu Glu Val Asn Asn Thr
                1185                1190                1195

GTG AGC GGT GCG GCG GCC CAG GCG GAT GTG CGC GTC TTT GAG GAG CTC     3770
Val Ser Gly Ala Ala Ala Gln Ala Asp Val Arg Val Phe Glu Glu Leu
            1200                1205                1210
```

FIG. 1K

```
CGC GGA CTC AGC GTG GAC ATG AGC CTG GCC GTG GAG CAG GGC GCC CCC    3818
Arg Gly Leu Ser Val Asp Met Ser Leu Ala Val Glu Gln Gly Ala Pro
        1215                1220                1225

GTG GTG GTC AGC GCC GCG GTG CAG ACG GGC GAC AAC ATC ACG TGG ACC    3866
Val Val Val Ser Ala Ala Val Gln Thr Gly Asp Asn Ile Thr Trp Thr
    1230                1235                1240

TTC GAC ATG GGG GAC GGC ACC GTG CTG TCG GGC CCG GAG GCA ACA GTG    3914
Phe Asp Met Gly Asp Gly Thr Val Leu Ser Gly Pro Glu Ala Thr Val
1245                1250                1255                1260

GAG CAT GTG TAC CTG CGG GCA CAG AAC TGC ACA GTG ACC GTG GGT GCG    3962
Glu His Val Tyr Leu Arg Ala Gln Asn Cys Thr Val Thr Val Gly Ala
                1265                1270                1275

GCC AGC CCC GCC GGC CAC CTG GCC CGG AGC CTG CAC GTG CTG GTC TTC    4010
Ala Ser Pro Ala Gly His Leu Ala Arg Ser Leu His Val Leu Val Phe
            1280                1285                1290

GTC CTG GAG GTG CTG CGC GTT GAA CCC GCC GCC TGC ATC CCC ACG CAG    4058
Val Leu Glu Val Leu Arg Val Glu Pro Ala Ala Cys Ile Pro Thr Gln
        1295                1300                1305

CCT GAC GCG CGG CTC ACG GCC TAC GTC ACC GGG AAC CCG GCC CAC TAC    4106
Pro Asp Ala Arg Leu Thr Ala Tyr Val Thr Gly Asn Pro Ala His Tyr
    1310                1315                1320
```

FIG. 1L

```
CTC TTC GAC TGG ACC TTC GGG GAT GGC TCC TCC AAC ACG ACC GTG CGG    4154
Leu Phe Asp Trp Thr Phe Gly Asp Gly Ser Ser Asn Thr Thr Val Arg
1325                1330                1335                1340

GGG TGC CCG ACG GTG ACA CAC AAC TTC ACG CGG AGC GGC ACG TTC CCC    4202
Gly Cys Pro Thr Val Thr His Asn Phe Thr Arg Ser Gly Thr Phe Pro
                1345                1350                1355

CTG GCG CTG GTG CTG TCC AGC CGC GTG AAC AGG GCG CAT TAC TTC ACC    4250
Leu Ala Leu Val Leu Ser Ser Arg Val Asn Arg Ala His Tyr Phe Thr
            1360                1365                1370

AGC ATC TGC GTG GAG CCA GAG GTG GGC AAC GTC ACC CTG CAG CCA GAG    4298
Ser Ile Cys Val Glu Pro Glu Val Gly Asn Val Thr Leu Gln Pro Glu
        1375                1380                1385

AGG CAG TTT GTG CAG CTC GGG GAC GAG GCC TGG CTG GTG GCA TGT GCC    4346
Arg Gln Phe Val Gln Leu Gly Asp Glu Ala Trp Leu Val Ala Cys Ala
    1390                1395                1400

TGG CCC CCG TTC CCC TAC CGC TAC ACC TGG GAC TTT GGC ACC GAG GAA    4394
Trp Pro Pro Phe Pro Tyr Arg Tyr Thr Trp Asp Phe Gly Thr Glu Glu
1405                1410                1415                1420

GCC GCC CCC TCC CGT GCC AGG GGC CCT GAG GTG ACG TTC ATC TAC CGA    4442
Ala Ala Pro Thr Arg Ala Arg Gly Pro Glu Val Thr Phe Ile Tyr Arg
                1425                1430                1435
```

FIG. 1M

```
GAC CCA GGC TCC TAT CTT GTG ACA GTC ACC GCG TCC AAC AAC ATC TCT    4490
Asp Pro Gly Ser Tyr Leu Val Thr Val Thr Ala Ser Asn Asn Ile Ser
            1440                1445                1450

GCT GCC AAT GAC TCA GCC CTG GTG GAG GTG CAG GAG CCC GTG CTG GTC    4538
Ala Ala Asn Asp Ser Ala Leu Val Glu Val Gln Glu Pro Val Leu Val
        1455                1460                1465

ACC AGC ATC AAG GTC AAT GGC TCC CTT GGG CTG GAG CTG CAG CAG CCG    4586
Thr Ser Ile Lys Val Asn Gly Ser Leu Gly Leu Glu Leu Gln Gln Pro
    1470                1475                1480

TAC CTG TTC TCT GCT GTG GGC CGT GGG CGC CCC GCC AGC TAC CTG TGG    4634
Tyr Leu Phe Ser Ala Val Gly Arg Gly Arg Pro Ala Ser Tyr Leu Trp
1485                1490                1495                1500

GAT CTG GGG GAC GGT GGG TGG CTC GAG GGT CCG GAG GTC ACC CAC GCT    4682
Asp Leu Gly Asp Gly Gly Trp Leu Glu Gly Pro Glu Val Thr His Ala
                1505                1510                1515

TAC AAC AGC ACA GGT GAC TTC ACC GTT AGG GTG GCC GGC TGG AAT GAG    4730
Tyr Asn Ser Thr Gly Asp Phe Thr Val Arg Val Ala Gly Trp Asn Glu
            1520                1525                1530

GTG AGC CGC AGC GAG GCC TGG CTC AAT GTG ACG GTG AAG CGG CGC GTG    4778
Val Ser Arg Ser Glu Ala Trp Leu Asn Val Thr Val Lys Arg Arg Val
        1535                1540                1545
```

FIG. 1N

```
CGG GGG CTC GTC GTC AAT GCA AGC CGC ACG GTG GTG CCC CTG AAT GGG    4826
Arg Gly Leu Val Val Asn Ala Ser Arg Thr Val Val Pro Leu Asn Gly
    1550                1555                1560

AGC GTG AGC TTC AGC ACG TCG CTG GAG GCC GGC AGT GAT GTG CGC TAT    4874
Ser Val Ser Phe Ser Thr Ser Leu Glu Ala Gly Ser Asp Val Arg Tyr
1565                1570                1575                1580

TCC TGG GTG CTC TGT GAC CGC TGC ACG CCC ATC CCT GGG GGT CCT ACC    4922
Ser Trp Val Leu Cys Asp Arg Cys Thr Pro Ile Pro Gly Gly Pro Thr
                1585                1590                1595

ATC TCT TAC ACC TTC CGC TCC GTG GGC ACC TTC AAT ATC ATC GTC ACG    4970
Ile Ser Tyr Thr Phe Arg Ser Val Gly Thr Phe Asn Ile Ile Val Thr
            1600                1605                1610

GCT GAG AAC GAG GTG GGC TCC GCC CAG GAC AGC ATC TTC GTC TAT GTC    5018
Ala Glu Asn Glu Val Gly Ser Ala Gln Asp Ser Ile Phe Val Tyr Val
        1615                1620                1625

CTG CAG CTC ATA GAG GGG CTG CAG GTG GTG GGC GGT GGC CGC TAC TTC    5066
Leu Gln Leu Ile Glu Gly Leu Gln Val Val Gly Gly Gly Arg Tyr Phe
    1630                1635                1640

CCC ACC AAC CAC ACG GTA CAG CTG CAG GCC GTG GTT AGG GAT GGC ACC    5114
Pro Thr Asn His Thr Val Gln Leu Gln Ala Val Val Arg Asp Gly Thr
1645                1650                1655                1660
```

FIG. 10

```
AAC GTC TCC TAC AGC TGG ACT GCC TGG AGG GAC AGG GGC CCG GCC CTG     5162
Asn Val Ser Tyr Ser Trp Thr Ala Trp Arg Asp Arg Gly Pro Ala Leu
                1665                1670                1675

GCC GGC AGC GGC AAA GGC TTC TCG CTC ACC GTG CTC GAG GCC GGC ACC     5210
Ala Gly Ser Gly Lys Gly Phe Ser Leu Thr Val Leu Glu Ala Gly Thr
            1680                1685                1690

TAC CAT GTG CAG CTG CGG GCC ACC AAC ATG CTG GGC AGC GCC TGG GCC     5258
Tyr His Val Gln Leu Arg Ala Thr Asn Met Leu Gly Ser Ala Trp Ala
        1695                1700                1705

GAC TGC ACC ATG GAC TTC GTG GAG CCT GTG GGG TGG CTG ATG GTG GCC     5306
Asp Cys Thr Met Asp Phe Val Glu Pro Val Gly Trp Leu Met Val Ala
    1710                1715                1720

GCC TCC CCG AAC CCA GCT GCC GTC AAC ACA AGC GTC ACC CTC AGT GCC     5354
Ala Ser Pro Asn Pro Ala Ala Val Asn Thr Ser Val Thr Leu Ser Ala
1725                1730                1735                1740

GAG CTG GCT GGT GGC AGT GGT GTC GTA TAC ACT TGG TCC TTG GAG GAG     5402
Glu Leu Ala Gly Gly Ser Gly Val Val Tyr Thr Trp Ser Leu Glu Glu
                1745                1750                1755

GGG CTG AGC TGG GAG ACC TCC GAG CCA TTT ACC ACC CAT AGC TTC CCC     5450
Gly Leu Ser Trp Glu Thr Ser Glu Pro Phe Thr Thr His Ser Phe Pro
            1760                1765                1770
```

FIG. 1P

```
ACA CCC GGC CTG CAC TTG GTC ACC ATG ACG GCA GGG AAC CCG CTG GGC    5498
Thr Pro Gly Leu His Leu Val Thr Met Thr Ala Gly Asn Pro Leu Gly
        1775                1780                1785

TCA GCC AAC GCC ACC GTG GAA GTG GAT GTG CAG GTG CCT GTG AGT GGC    5546
Ser Ala Asn Ala Thr Val Glu Val Asp Val Gln Val Pro Val Ser Gly
    1790                1795                1800

CTC AGC ATC AGG GCC AGC GAG CCC GGA GGC AGC TTC GTG GCG GCC GGG    5594
Leu Ser Ile Arg Ala Ser Glu Pro Gly Gly Ser Phe Val Ala Ala Gly
1805                1810                1815                1820

TCC TCT GTG CCC TTT TGG GGG CAG CTG GCC ACG GGC ACC AAT GTG AGC    5642
Ser Ser Val Pro Phe Trp Gly Gln Leu Ala Thr Gly Thr Asn Val Ser
                1825                1830                1835

TGG TGC TGG GCT GTG CCC GGC GGC AGC AGC AAG CGT GGC CCT CAT GTC    5690
Trp Cys Trp Ala Val Pro Gly Gly Ser Ser Lys Arg Gly Pro His Val
            1840                1845                1850

ACC ATG GTC TTC CCG GAT GCT GGC ACC TTC TCC ATC CGG CTC AAT GCC    5738
Thr Met Val Phe Pro Asp Ala Gly Thr Phe Ser Ile Arg Leu Asn Ala
        1855                1860                1865

TCC AAC GCA GTC AGC TGG GTC TCA GCC ACG TAC AAC CTC ACG GCG GAG    5786
Ser Asn Ala Val Ser Trp Val Ser Ala Thr Tyr Asn Leu Thr Ala Glu
    1870                1875                1880
```

FIG. 1Q

```
GAG CCC ATC GTG GGC CTG GTG CTG TGG GCC AGC AGC AAG GTG GTG GCG    5834
Glu Pro Ile Val Gly Leu Val Leu Trp Ala Ser Ser Lys Val Val Ala
1885                1890                1895                1900

CCC GGG CAG CTG GTC CAT TTT CAG ATC CTG CTG GCT GCC GGC TCA GCT    5882
Pro Gly Gln Leu Val His Phe Gln Ile Leu Leu Ala Ala Gly Ser Ala
                1905                1910                1915

GTC ACC TTC CGC CTG CAG GTC GGC GGG GCC AAC CCC GAG GTG CTC CCC    5930
Val Thr Phe Arg Leu Gln Val Gly Gly Ala Asn Pro Glu Val Leu Pro
            1920                1925                1930

GGG CCC CGT TTC TCC CAC AGC TTC CCC CGC GTC GGA GAC CAC GTG GTG    5978
Gly Pro Arg Phe Ser His Ser Phe Pro Arg Val Gly Asp His Val Val
        1935                1940                1945

AGC GTG CGG GGC AAA AAC CAC GTG AGC TGG GCC CAG GCG CAG GTG CGC    6026
Ser Val Arg Gly Lys Asn His Val Ser Trp Ala Gln Ala Gln Val Arg
    1950                1955                1960

ATC GTG GTG CTG GAG GCC GTG AGT GGG CTG CAG GTG CCC AAC TGC TGC    6074
Ile Val Val Leu Glu Ala Val Ser Gly Leu Gln Val Pro Asn Cys Cys
1965                1970                1975                1980

GAG CCT GGC ATC GCC ACG GGC ACT GAG AGG AAC TTC ACA GCC CGC GTG    6122
Glu Pro Gly Ile Ala Thr Gly Thr Glu Arg Asn Phe Thr Ala Arg Val
                1985                1990                1995
```

FIG. 1R

```
CAG CGC GGC TCT CGG GTC GCC TAC GCC TGG TAC TTC TCG CTG CAG AAG    6170
Gln Arg Gly Ser Arg Val Ala Tyr Ala Trp Tyr Phe Ser Leu Gln Lys
            2000                2005                2010

GTC CAG GGC GAC TCG CTG GTC ATC CTG TCG GGC CGC GAC GTC ACC TAC    6218
Val Gln Gly Asp Ser Leu Val Ile Leu Ser Gly Arg Asp Val Thr Tyr
        2015                2020                2025

ACG CCC GTG GCC GCG GGG CTG TTG GAG ATC CAG GTG CGC GCC TTC AAC    6266
Thr Pro Val Ala Ala Gly Leu Leu Glu Ile Gln Val Arg Ala Phe Asn
    2030                2035                2040

GCC CTG GGC AGT GAG AAC CGC ACG CTG GTG CTG GAG GTT CAG GAC GCC    6314
Ala Leu Gly Ser Glu Asn Arg Thr Leu Val Leu Glu Val Gln Asp Ala
2045                2050                2055                2060

GTC CAG TAT GTG GCC CTG CAG AGC GGC CCC TGC TTC ACC AAC CGC TCG    6362
Val Gln Tyr Val Ala Leu Gln Ser Gly Pro Cys Phe Thr Asn Arg Ser
                2065                2070                2075

GCG CAG TTT GAG GCC GCC ACC AGC CCC AGC CCC CGG CGT GTG GCC TAC    6410
Ala Gln Phe Glu Ala Ala Thr Ser Pro Ser Pro Arg Arg Val Ala Tyr
            2080                2085                2090

CAC TGG GAC TTT GGG GAT GGG TCG CCA GGG CAG GAC ACA GAT GAG CCC    6458
His Trp Asp Phe Gly Asp Gly Ser Pro Gly Gln Asp Thr Asp Glu Pro
        2095                2100                2105
```

FIG. 1S

```
AGG GCC GAG CAC TCC TAC CTG AGG CCT GGG GAC TAC CGC GTG CAG GTG    6506
Arg Ala Glu His Ser Tyr Leu Arg Pro Gly Asp Tyr Arg Val Gln Val
    2110                2115                2120

AAC GCC TCC AAC CTG GTG AGC TTC TTC GTG GCG CAG GCC ACG GTG ACC    6554
Asn Ala Ser Asn Leu Val Ser Phe Phe Val Ala Gln Ala Thr Val Thr
2125                2130                2135                2140

GTC CAG GTG CTG GCC TGC CGG GAG CCG GAG GTG GAC GTG GTC CTG CCC    6602
Val Gln Val Leu Ala Cys Arg Glu Pro Glu Val Asp Val Val Leu Pro
                2145                2150                2155

CTG CAG GTG CTG ATG CGG CGA TCA CAG CGC AAC TAC TTG GAG GCC CAC    6650
Leu Gln Val Leu Met Arg Arg Ser Gln Arg Asn Tyr Leu Glu Ala His
            2160                2165                2170

GTT GAC CTG CGC GAC TGC GTC ACC TAC CAG ACT GAG TAC CGC TGG GAG    6698
Val Asp Leu Arg Asp Cys Val Thr Tyr Gln Thr Glu Tyr Arg Trp Glu
        2175                2180                2185

GTG TAT CGC ACC GCC AGC TGC CAG CGG CCG GGG CGC CCA GCG CGT GTG    6746
Val Tyr Arg Thr Ala Ser Cys Gln Arg Pro Gly Arg Pro Ala Arg Val
    2190                2195                2200

GCC CTG CCC GGC GTG GAC GTG AGC CGG CCT CGG CTG GTG CTG CCG CGG    6794
Ala Leu Pro Gly Val Asp Val Ser Arg Pro Arg Leu Val Leu Pro Arg
2205                2210                2215                2220
```

FIG. 1T

```
CTG GCG CTG CCT GTG GGG CAC TAC TGC TTT GTG TTT GTC GTG TCA TTT    6842
Leu Ala Leu Pro Val Gly His Tyr Cys Phe Val Phe Val Val Ser Phe
                2225                2230                2235

GGG GAC ACG CCA CTG ACA CAG AGC ATC CAG GCC AAT GTG ACG GTG GCC    6890
Gly Asp Thr Pro Leu Thr Gln Ser Ile Gln Ala Asn Val Thr Val Ala
            2240                2245                2250

CCC GAG CGC CTG GTG CCC ATC ATT GAG GGT GGC TCA TAC CGC GTG TGG    6938
Pro Glu Arg Leu Val Pro Ile Ile Glu Gly Gly Ser Tyr Arg Val Trp
        2255                2260                2265

TCA GAC ACA CGG GAC CTG GTG CTG GAT GGG AGC GAG TCC TAC GAC CCC    6986
Ser Asp Thr Arg Asp Leu Val Leu Asp Gly Ser Glu Ser Tyr Asp Pro
    2270                2275                2280

AAC CTG GAG GAC GGC GAC CAG ACG CCG CTC AGT TTC CAC TGG GCC TGT    7034
Asn Leu Glu Asp Gly Asp Gln Thr Pro Leu Ser Phe His Trp Ala Cys
2285                2290                2295                2300

GTG GCT TCG ACA CAG AGG GAG GCT GGC GGG TGT GCG CTG AAC TTT GGG    7082
Val Ala Ser Thr Gln Arg Glu Ala Gly Gly Cys Ala Leu Asn Phe Gly
                2305                2310                2315

CCC CGC GGG AGC AGC ACG GTC ACC ATT CCA CGG GAG CGG CTG GCG GCT    7130
Pro Arg Gly Ser Ser Thr Val Thr Ile Pro Arg Glu Arg Leu Ala Ala
            2320                2325                2330
```

FIG. 1U

```
GGC GTG GAG TAC ACC TTC AGC CTG ACC GTG TGG AAG GCC GGC CGC AAG    7178
Gly Val Glu Tyr Thr Phe Ser Leu Thr Val Trp Lys Ala Gly Arg Lys
       2335                2340                2345

GAG GAG GCC ACC AAC CAG ACG GTG CTG ATC CGG AGT GGC CGG GTG CCC    7226
Glu Glu Ala Thr Asn Gln Thr Val Leu Ile Arg Ser Gly Arg Val Pro
    2350                2355                2360

ATT GTG TCC TTG GAG TGT GTG TCC TGC AAG GCA CAG GCC GTG TAC GAA    7274
Ile Val Ser Leu Glu Cys Val Ser Cys Lys Ala Gln Ala Val Tyr Glu
2365                2370                2375                2380

GTG AGC CGC AGC TCC TAC GTG TAC TTG GAG GGC CGC TGC CTC AAT TGC    7322
Val Ser Arg Ser Ser Tyr Val Tyr Leu Glu Gly Arg Cys Leu Asn Cys
                2385                2390                2395

AGC AGC GGC TCC AAG CGA GGG CGG TGG GCT GCA CGT ACG TTC AGC AAC    7370
Ser Ser Gly Ser Lys Arg Gly Arg Trp Ala Ala Arg Thr Phe Ser Asn
            2400                2405                2410

AAG ACG CTG GTG CTG GAT GAG ACC ACC ACA TCC ACG GGC AGT GCA GGC    7418
Lys Thr Leu Val Leu Asp Glu Thr Thr Thr Ser Thr Gly Ser Ala Gly
       2415                2420                2425

ATG CGA CTG GTG CTG CGG CGG GGC GTG CTG CGG GAC GGC GAG GGA TAC    7466
Met Arg Leu Val Leu Arg Arg Gly Val Leu Arg Asp Gly Glu Gly Tyr
    2430                2435                2440
```

FIG. 1V

```
ACC TTC ACG CTC ACG GTG CTG GGC CGC TCT GGC GAG GAG GAG GGC TGC    7514
Thr Phe Thr Leu Thr Val Leu Gly Arg Ser Gly Glu Glu Glu Gly Cys
2445                2450                2455                2460

GCC TCC ATC CGC CTG TCC CCC AAC CGC CCG CCG CTG GGG GGC TCT TGC    7562
Ala Ser Ile Arg Leu Ser Pro Asn Arg Pro Pro Leu Gly Gly Ser Cys
                    2465                2470                2475

CGC CTC TTC CCA CTG GGC GCT GTG CAC GCC CTC ACC ACC AAG GTG CAC    7610
Arg Leu Phe Pro Leu Gly Ala Val His Ala Leu Thr Thr Lys Val His
            2480                2485                2490

TTC GAA TGC ACG GGC TGG CAT GAC GCG GAG GAT GCT GGC GCC CCG CTG    7658
Phe Glu Cys Thr Gly Trp His Asp Ala Glu Asp Ala Gly Ala Pro Leu
        2495                2500                2505

GTG TAC GCC CTG CTG CTG CGG CGC TGT CGC CAG GGC CAC TGC GAG GAG    7706
Val Tyr Ala Leu Leu Leu Arg Arg Cys Arg Gln Gly His Cys Glu Glu
    2510                2515                2520

TTC TGT GTC TAC AAG GGC AGC CTC TCC AGC TAC GGA GCC GTG CTG CCC    7754
Phe Cys Val Tyr Lys Gly Ser Leu Ser Ser Tyr Gly Ala Val Leu Pro
2525                2530                2535                2540

CCG GGT TTC AGG CCA CAC TTC GAG GTG GGC CTG GCC GTG GTG GTG CAG    7802
Pro Gly Phe Arg Pro His Phe Glu Val Gly Leu Ala Val Val Val Gln
                    2545                2550                2555
```

FIG. 1W

```
GAC CAG CTG GGA GCC GCT GTG GTC GCC CTC AAC AGG TCT TTG GCC ATC     7850
Asp Gln Leu Gly Ala Ala Val Val Ala Leu Asn Arg Ser Leu Ala Ile
            2560                2565                2570

ACC CTC CCA GAG CCC AAC GGC AGC GCA ACG GGG CTC ACA GTC TGG CTG     7898
Thr Leu Pro Glu Pro Asn Gly Ser Ala Thr Gly Leu Thr Val Trp Leu
        2575                2580                2585

CAC GGG CTC ACC GCT AGT GTG CTC CCA GGG CTG CTG CGG CAG GCC GAT     7946
His Gly Leu Thr Ala Ser Val Leu Pro Gly Leu Leu Arg Gln Ala Asp
    2590                2595                2600

CCC CAG CAC GTC ATC GAG TAC TCG TTG GCC CTG GTC ACC GTG CTG AAC     7994
Pro Gln His Val Ile Glu Tyr Ser Leu Ala Leu Val Thr Val Leu Asn
2605                2610                2615                2620

GAG TAC GAG CGG GCC CTG GAC GTG GCG GCA GAG CCC AAG CAC GAG CGG     8042
Glu Tyr Glu Arg Ala Leu Asp Val Ala Ala Glu Pro Lys His Glu Arg
                2625                2630                2635

CAG CAC CGA GCC CAG ATA CGC AAG AAC ATC ACG GAG ACT CTG GTG TCC     8090
Gln His Arg Ala Gln Ile Arg Lys Asn Ile Thr Glu Thr Leu Val Ser
            2640                2645                2650

CTG AGG GTC CAC ACT GTG GAT GAC ATC CAG CAG ATC GCT GCT GCG CTG     8138
Leu Arg Val His Thr Val Asp Asp Ile Gln Gln Ile Ala Ala Ala Leu
        2655                2660                2665
```

FIG. 1X

```
GCC CAG TGC ATG GGG CCC AGC AGG GAG CTC GTA TGC CGC TCG TGC CTG    8186
Ala Gln Cys Met Gly Pro Ser Arg Glu Leu Val Cys Arg Ser Cys Leu
    2670                2675                2680

AAG CAG ACG CTG CAC AAG CTG GAG GCC ATG ATG CTC ATC CTG CAG GCA    8234
Lys Gln Thr Leu His Lys Leu Glu Ala Met Met Leu Ile Leu Gln Ala
2685                2690                2695                2700

GAG ACC ACC GCG GGC ACC GTG ACG CCC ACC GCC ATC GGA GAC AGC ATC    8282
Glu Thr Thr Ala Gly Thr Val Thr Pro Thr Ala Ile Gly Asp Ser Ile
                2705                2710                2715

CTC AAC ATC ACA GGA GAC CTC ATC CAC CTG GCC AGC TCG GAC GTG CGG    8330
Leu Asn Ile Thr Gly Asp Leu Ile His Leu Ala Ser Ser Asp Val Arg
            2720                2725                2730

GCA CCA CAG CCC TCA GAG CTG GGA GCC GAG TCA CCA TCT CGG ATG GTG    8378
Ala Pro Gln Pro Ser Glu Leu Gly Ala Glu Ser Pro Ser Arg Met Val
        2735                2740                2745

GCG TCC CAG GCC TAC AAC CTG ACC TCT GCC CTC ATG CGC ATC CTC ATG    8426
Ala Ser Gln Ala Tyr Asn Leu Thr Ser Ala Leu Met Arg Ile Leu Met
    2750                2755                2760

CGC TCC CGC GTG CTC AAC GAG GAG CCC CTG ACG CTG GCG GGC GAG GAG    8474
Arg Ser Arg Val Leu Asn Glu Glu Pro Leu Thr Leu Ala Gly Glu Glu
2765                2770                2775                2780
```

FIG. 1Y

```
ATC GTG GCC CAG GGC AAG CGC TCG GAC CCG CGG AGC CTG CTG TGC TAT    8522
Ile Val Ala Gln Gly Lys Arg Ser Asp Pro Arg Ser Leu Leu Cys Tyr
                2785                2790                2795

GGC GGC GCC CCA GGG CCT GGC TGC CAC TTC TCC ATC CCC GAG GCT TTC    8570
Gly Gly Ala Pro Gly Pro Gly Cys His Phe Ser Ile Pro Glu Ala Phe
            2800                2805                2810

AGC GGG GCC CTG GCC AAC CTC AGT GAC GTG GTG CAG CTC ATC TTT CTG    8618
Ser Gly Ala Leu Ala Asn Leu Ser Asp Val Val Gln Leu Ile Phe Leu
        2815                2820                2825

GTG GAC TCC AAT CCC TTT CCC TTT GGC TAT ATC AGC AAC TAC ACC GTC    8666
Val Asp Ser Asn Pro Phe Pro Phe Gly Tyr Ile Ser Asn Tyr Thr Val
    2830                2835                2840

TCC ACC AAG GTG GCC TCG ATG GCA TTC CAG ACA CAG GCC GGC GCC CAG    8714
Ser Thr Lys Val Ala Ser Met Ala Phe Gln Thr Gln Ala Gly Ala Gln
2845                2850                2855                2860

ATC CCC ATC GAG CGG CTG GCC TCA GAG CGC GCC ATC ACC GTG AAG GTG    8762
Ile Pro Ile Glu Arg Leu Ala Ser Glu Arg Ala Ile Thr Val Lys Val
                2865                2870                2875

CCC AAC AAC TCG GAC TGG GCT GCC CGG GGC CAC CGC AGC TCC GCC AAC    8810
Pro Asn Asn Ser Asp Trp Ala Ala Arg Gly His Arg Ser Ser Ala Asn
            2880                2885                2890
```

FIG. 1Z

```
TCC GCC AAC TCC GTT GTG GTC CAG CCC CAG GCC TCC GTC GGT GCT GTG    8858
Ser Ala Asn Ser Val Val Val Gln Pro Gln Ala Ser Val Gly Ala Val
        2895                2900                2905

GTC ACC CTG GAC AGC AGC AAC CCT GCG GCC GGG CTG CAT CTG CAG CTC    8906
Val Thr Leu Asp Ser Ser Asn Pro Ala Ala Gly Leu His Leu Gln Leu
    2910                2915                2920

AAC TAT ACG CTG CTG GAC GGC CAC TAC CTG TCT GAG GAA CCT GAG CCC    8954
Asn Tyr Thr Leu Leu Asp Gly His Tyr Leu Ser Glu Glu Pro Glu Pro
2925                2930                2935                2940

TAC CTG GCA GTC TAC CTA CAC TCG GAG CCC CGG CCC AAT GAG CAC AAC    9002
Tyr Leu Ala Val Tyr Leu His Ser Glu Pro Arg Pro Asn Glu His Asn
                2945                2950                2955

TGC TCG GCT AGC AGG AGG ATC CGC CCA GAG TCA CTC CAG GGT GCT GAC    9050
Cys Ser Ala Ser Arg Arg Ile Arg Pro Glu Ser Leu Gln Gly Ala Asp
            2960                2965                2970

CAC CGG CCC TAC ACC TTC TTC ATT TCC CCG GGG AGC AGA GAC CCA GCG    9098
His Arg Pro Tyr Thr Phe Phe Ile Ser Pro Gly Ser Arg Asp Pro Ala
        2975                2980                2985

GGG AGT TAC CAT CTG AAC CTC TCC AGC CAC TTC CGC TGG TCG GCG CTG    9146
Gly Ser Tyr His Leu Asn Leu Ser Ser His Phe Arg Trp Ser Ala Leu
    2990                2995                3000
```

FIG. 1AA

```
CAG GTG TCC GTG GGC CTG TAC ACG TCC CTG TGC CAG TAC TTC AGC GAG    9194
Gln Val Ser Val Gly Leu Tyr Thr Ser Leu Cys Gln Tyr Phe Ser Glu
3005                3010                3015                3020

GAG GAC ATG GTG TGG CGG ACA GAG GGG CTG CTG CCC CTG GAG GAG ACC    9242
Glu Asp Met Val Trp Arg Thr Glu Gly Leu Leu Pro Leu Glu Glu Thr
                3025                3030                3035

TCG CCC CGC CAG GCC GTC TGC CTC ACC CGC CAC CTC ACC GCC TTC GGC    9290
Ser Pro Arg Gln Ala Val Cys Leu Thr Arg His Leu Thr Ala Phe Gly
            3040                3045                3050

GCC AGC CTC TTC GTG CCC CCA AGC CAT GTC CGC TTT GTG TTT CCT GAG    9338
Ala Ser Leu Phe Val Pro Pro Ser His Val Arg Phe Val Phe Pro Glu
        3055                3060                3065

CCG ACA GCG GAT GTA AAC TAC ATC GTC ATG CTG ACA TGT GCT GTG TGC    9386
Pro Thr Ala Asp Val Asn Tyr Ile Val Met Leu Thr Cys Ala Val Cys
    3070                3075                3080

CTG GTG ACC TAC ATG GTC ATG GCC GCC ATC CTG CAC AAG CTG GAC CAG    9434
Leu Val Thr Tyr Met Val Met Ala Ala Ile Leu His Lys Leu Asp Gln
3085                3090                3095                3100

TTG GAT GCC AGC CGG GGC CGC GCC ATC CCT TTC TGT GGG CAG CGG GGC    9482
Leu Asp Ala Ser Arg Gly Arg Ala Ile Pro Phe Cys Gly Gln Arg Gly
                3105                3110                3115
```

FIG. 1AB

```
CGC TTC AAG TAC GAG ATC CTC GTC AAG ACA GGC TGG GGC CGG GGC TCA    9530
Arg Phe Lys Tyr Glu Ile Leu Val Lys Thr Gly Trp Gly Arg Gly Ser
            3120                3125                3130

GGT ACC ACG GCC CAC GTG GGC ATC ATG CTG TAT GGG GTG GAC AGC CGG    9578
Gly Thr Thr Ala His Val Gly Ile Met Leu Tyr Gly Val Asp Ser Arg
        3135                3140                3145

AGC GGC CAC CGG CAC CTG GAC GGC GAC AGA GCC TTC CAC CGC AAC AGC    9626
Ser Gly His Arg His Leu Asp Gly Asp Arg Ala Phe His Arg Asn Ser
    3150                3155                3160

CTG GAC ATC TTC CGG ATC GCC ACC CCG CAC AGC CTG GGT AGC GTG TGG    9674
Leu Asp Ile Phe Arg Ile Ala Thr Pro His Ser Leu Gly Ser Val Trp
3165                3170                3175                3180

AAG ATC CGA GTG TGG CAC GAC AAC AAA GGG CTC AGC CCT GCC TGG TTC    9722
Lys Ile Arg Val Trp His Asp Asn Lys Gly Leu Ser Pro Ala Trp Phe
                3185                3190                3195

CTG CAG CAC GTC ATC GTC AGG GAC CTG CAG ACG GCA CGC AGC GCC TTC    9770
Leu Gln His Val Ile Val Arg Asp Leu Gln Thr Ala Arg Ser Ala Phe
            3200                3205                3210

TTC CTG GTC AAT GAC TGG CTT TCG GTG GAG ACG GAG GCC AAC GGG GGC    9818
Phe Leu Val Asn Asp Trp Leu Ser Val Glu Thr Glu Ala Asn Gly Gly
        3215                3220                3225
```

FIG. 1AC

```
CTG GTG GAG AAG GAG GTG CTG GCC GCG AGC GAC GCA GCC CTT TTG CGC    9866
Leu Val Glu Lys Glu Val Leu Ala Ala Ser Asp Ala Ala Leu Leu Arg
    3230                3235                3240

TTC CGG CGC CTG CTG GTG GCT GAG CTG CAG CGT GGC TTC TTT GAC AAG    9914
Phe Arg Arg Leu Leu Val Ala Glu Leu Gln Arg Gly Phe Phe Asp Lys
3245                3250                3255                3260

CAC ATC TGG CTC TCC ATA TGG GAC CGG CCG CCT CGT AGC CGT TTC ACT    9962
His Ile Trp Leu Ser Ile Trp Asp Arg Pro Pro Arg Ser Arg Phe Thr
                3265                3270                3275

CGC ATC CAG AGG GCC ACC TGC TGC GTT CTC CTC ATC TGC CTC TTC CTG   10010
Arg Ile Gln Arg Ala Thr Cys Cys Val Leu Leu Ile Cys Leu Phe Leu
            3280                3285                3290

GGC GCC AAC GCC GTG TGG TAC GGG GCT GTT GGC GAC TCT GCC TAC AGC   10058
Gly Ala Asn Ala Val Trp Tyr Gly Ala Val Gly Asp Ser Ala Tyr Ser
        3295                3300                3305

ACG GGG CAT GTG TCC AGG CTG AGC CCG CTG AGC GTC GAC ACA GTC GCT   10106
Thr Gly His Val Ser Arg Leu Ser Pro Leu Ser Val Asp Thr Val Ala
    3310                3315                3320

GTT GGC CTG GTG TCC AGC GTG GTT GTC TAT CCC GTC TAC CTG GCC ATC   10154
Val Gly Leu Val Ser Ser Val Val Val Tyr Pro Val Tyr Leu Ala Ile
3325                3330                3335                3340
```

FIG. 1AD

```
CTT TTT CTC TTC CGG ATG TCC CGG AGC AAG GTG GCT GGG AGC CCG AGC   10202
Leu Phe Leu Phe Arg Met Ser Arg Ser Lys Val Ala Gly Ser Pro Ser
                3345                3350                3355

CCC ACA CCT GCC GGG CAG CAG GTG CTG GAC ATC GAC AGC TGC CTG GAC   10250
Pro Thr Pro Ala Gly Gln Gln Val Leu Asp Ile Asp Ser Cys Leu Asp
            3360                3365                3370

TCG TCC GTG CTG GAC AGC TCC TTC CTC ACG TTC TCA GGC CTC CAC GCT   10298
Ser Ser Val Leu Asp Ser Ser Phe Leu Thr Phe Ser Gly Leu His Ala
        3375                3380                3385

GAG GCC TTT GTT GGA CAG ATG AAG AGT GAC TTG TTT CTG GAT GAT TCT   10346
Glu Ala Phe Val Gly Gln Met Lys Ser Asp Leu Phe Leu Asp Asp Ser
    3390                3395                3400

AAG AGT CTG GTG TGC TGG CCC TCC GGC GAG GGA ACG CTC AGT TGG CCG   10394
Lys Ser Leu Val Cys Trp Pro Ser Gly Glu Gly Thr Leu Ser Trp Pro
3405                3410                3415                3420

GAC CTG CTC AGT GAC CCG TCC ATT GTG GGT AGC AAT CTG CGG CAG CTG   10442
Asp Leu Leu Ser Asp Pro Ser Ile Val Gly Ser Asn Leu Arg Gln Leu
                3425                3430                3435

GCA CGG GGC CAG GCG GGC CAT GGG CTG GGC CCA GAG GAG GAC GGC TTC   10490
Ala Arg Gly Gln Ala Gly His Gly Leu Gly Pro Glu Glu Asp Gly Phe
            3440                3445                3450
```

FIG. 1AE

```
TCC CTG GCC AGC CCC TAC TCG CCT GCC AAA TCC TTC TCA GCA TCA GAT   10538
Ser Leu Ala Ser Pro Tyr Ser Pro Ala Lys Ser Phe Ser Ala Ser Asp
        3455                3460                3465

GAA GAC CTG ATC CAG CAG GTC CTT GCC GAG GGG GTC AGC AGC CCA GCC   10586
Glu Asp Leu Ile Gln Gln Val Leu Ala Glu Gly Val Ser Ser Pro Ala
    3470                3475                3480

CCT ACC CAA GAC ACC CAC ATG GAA ACG GAC CTG CTC AGC AGC CTG TCC   10634
Pro Thr Gln Asp Thr His Met Glu Thr Asp Leu Leu Ser Ser Leu Ser
3485                3490                3495                3500

AGC ACT CCT GGG GAG AAG ACA GAG ACG CTG GCG CTG CAG AGG CTG GGG   10682
Ser Thr Pro Gly Glu Lys Thr Glu Thr Leu Ala Leu Gln Arg Leu Gly
                3505                3510                3515

GAG CTG GGG CCA CCC AGC CCA GGC CTG AAC TGG GAA CAG CCC CAG GCA   10730
Glu Leu Gly Pro Pro Ser Pro Gly Leu Asn Trp Glu Gln Pro Gln Ala
            3520                3525                3530

GCG AGG CTG TCC AGG ACA GGA CTG GTG GAG GGT CTG CGG AAG CGC CTG   10778
Ala Arg Leu Ser Arg Thr Gly Leu Val Glu Gly Leu Arg Lys Arg Leu
        3535                3540                3545

CTG CCG GCC TGG TGT GCC TCC CTG GCC CAC GGG CTC AGC CTG CTC CTG   10826
Leu Pro Ala Trp Cys Ala Ser Leu Ala His Gly Leu Ser Leu Leu Leu
    3550                3555                3560
```

FIG. 1AF

```
GTG GCT GTG GCT GTG GCT GTC TCA GGG TGG GTG GGT GCG AGC TTC CCC   10874
Val Ala Val Ala Val Ala Val Ser Gly Trp Val Gly Ala Ser Phe Pro
3565                3570                3575                3580

CCG GGC GTG AGT GTT GCG TGG CTC CTG TCC AGC AGC GCC AGC TTC CTG   10922
Pro Gly Val Ser Val Ala Trp Leu Leu Ser Ser Ser Ala Ser Phe Leu
                    3585                3590                3595

GCC TCA TTC CTC GGC TGG GAG CCA CTG AAG GTC TTG CTG GAA GCC CTG   10970
Ala Ser Phe Leu Gly Trp Glu Pro Leu Lys Val Leu Leu Glu Ala Leu
            3600                3605                3610

TAC TTC TCA CTG GTG GCC AAG CGG CTG CAC CCG GAT GAA GAT GAC ACC   11018
Tyr Phe Ser Leu Val Ala Lys Arg Leu His Pro Asp Glu Asp Asp Thr
        3615                3620                3625

CTG GTA GAG AGC CCG GCT GTG ACG CCT GTG AGC GCA CGT GTG CCC CGC   11066
Leu Val Glu Ser Pro Ala Val Thr Pro Val Ser Ala Arg Val Pro Arg
    3630                3635                3640

CTA CGG CCA CCC CAC GGC TTT GCA CTC TTC CTG GCC AAG GAA GAA GCC   11114
Val Arg Pro Pro His Gly Phe Ala Leu Phe Leu Ala Lys Glu Glu Ala
3645                3650                3655                3660

CGC AAG GTC AAG AGG CTA CAT GGC ATG CTG CGG AGC CTC CTG GTG TAC   11162
Arg Lys Val Lys Arg Leu His Gly Met Leu Arg Ser Leu Leu Val Tyr
                    3665                3670                3675
```

FIG. 1AG

```
ATG CTT TTT CTG CTG GTG ACC CTG CTG GCC AGC TAT GGG GAT GCC TCA   11210
Met Leu Phe Leu Leu Val Thr Leu Leu Ala Ser Tyr Gly Asp Ala Ser
            3680                3685                3690

TGC CAT GGG CAC GCC TAC CGT CTG CAA AGC GCC ATC AAG CAG GAG CTG   11258
Cys His Gly His Ala Tyr Arg Leu Gln Ser Ala Ile Lys Gln Glu Leu
        3695                3700                3705

CAC AGC CGG GCC TTC CTG GCC ATC ACG CGG TCT GAG GAG CTC TGG CCA   11306
His Ser Arg Ala Phe Leu Ala Ile Thr Arg Ser Glu Glu Leu Trp Pro
    3710                3715                3720

TGG ATG GCC CAC GTG CTG CTG CCC TAC GTC CAC GGG AAC CAG TCC AGC   11354
Trp Met Ala His Val Leu Leu Pro Tyr Val His Gly Asn Gln Ser Ser
3725                3730                3735                3740

CCA GAG CTG GGG CCC CCA CGG CTG CGG CAG GTG CGG CTG CAG GAA GCA   11402
Pro Glu Leu Gly Pro Pro Arg Leu Arg Gln Val Arg Leu Gln Glu Ala
                3745                3750                3755

CTC TAC CCA GAC CCT CCC GGC CCC AGG GTC CAC ACG TGC TCG GCC GCA   11450
Leu Tyr Pro Asp Pro Pro Gly Pro Arg Val His Thr Cys Ser Ala Ala
            3760                3765                3770

GGA GGC TTC AGC ACC AGC GAT TAC GAC GTT GGC TGG GAG AGT CCT CAC   11498
Gly Gly Phe Ser Thr Ser Asp Tyr Asp Val Gly Trp Glu Ser Pro His
        3775                3780                3785
```

FIG. 1AH

```
AAT GGC TCG GGG ACG TGG GCC TAT TCA GCG CCG GAT CTG CTG GGG GCA   11546
Asn Gly Ser Gly Thr Trp Ala Tyr Ser Ala Pro Asp Leu Leu Gly Ala
    3790                3795                3800

TGG TCC TGG GGC TCC TGT GCC GTG TAT GAC AGC GGG GGC TAC GTG CAG   11594
Trp Ser Trp Gly Ser Cys Ala Val Tyr Asp Ser Gly Gly Tyr Val Gln
3805                3810                3815                3820

GAG CTG GGC CTG AGC CTG GAG GAG AGC CGC GAC CGG CTG CGC TTC CTG   11642
Glu Leu Gly Leu Ser Leu Glu Glu Ser Arg Asp Arg Leu Arg Phe Leu
                3825                3830                3835

CAG CTG CAC AAC TGG CTG GAC AAC AGG AGC CGC GCT GTG TTC CTG GAG   11690
Gln Leu His Asn Trp Leu Asp Asn Arg Ser Arg Ala Val Phe Leu Glu
            3840                3845                3850

CTC ACG CGC TAC AGC CCG GCC GTG GGG CTG CAC GCC GCC GTC ACG CTG   11738
Leu Thr Arg Tyr Ser Pro Ala Val Gly Leu His Ala Ala Val Thr Leu
        3855                3860                3865

CGC CTC GAG TTC CCG GCG GCC GGC CGC GCC CTG GCC GCC CTC AGC GTC   11786
Arg Leu Glu Phe Pro Ala Ala Gly Arg Ala Leu Ala Ala Leu Ser Val
    3870                3875                3880

CGC CCC TTT GCG CTG CGC CGC CTC AGC GCG GGC CTC TCG CTG CCT CTG   11834
Arg Pro Phe Ala Leu Arg Arg Leu Ser Ala Gly Leu Ser Leu Pro Leu
3885                3890                3895                3900
```

FIG. 1AI

```
CTC ACC TCG GTG TGC CTG CTG CTG TTC GCC GTG CAC TTC GCC GTG GCC   11882
Leu Thr Ser Val Cys Leu Leu Leu Phe Ala Val His Phe Ala Val Ala
                3905                3910                3915

GAG GCC CGT ACT TGG CAC AGG GAA GGG CGC TGG CGC GTG CTG CGG CTC   11930
Glu Ala Arg Thr Trp His Arg Glu Gly Arg Val Arg Val Leu Arg Leu
            3920                3925                3930

GGA GCC TGG GCG CGG TGG CTG CTG GTG GCG CTG ACG GCG GCC ACG GCA   11978
Gly Ala Trp Ala Arg Trp Leu Leu Val Ala Leu Thr Ala Ala Thr Ala
        3935                3940                3945

CTG GTA CGC CTC GCC CAG CTG GGT GCC GCT GAC CGC CAG TGG ACC CGT   12026
Leu Val Arg Leu Ala Gln Leu Gly Ala Ala Asp Arg Gln Trp Thr Arg
    3950                3955                3960

TTC GTG CGC GGC CGC CCG CGC CGC TTC ACT AGC TTC GAC CAG GTG GCG   12074
Phe Val Arg Gly Arg Pro Arg Arg Phe Thr Ser Phe Asp Gln Val Ala
3965                3970                3975                3980

GAG CTG AGC TCC GCA GCC CGT GGC CTG GCG GCC TCG CTG CTC TTC CTG   12122
Gln Leu Ser Ser Ala Ala Arg Gly Leu Ala Ala Ser Leu Leu Phe Leu
                3985                3990                3995

CTT TTG GTC AAG GCT GCC CAG CAG CTA CGC TTC GTG CGC CAG TGG TCC   12170
Leu Leu Val Lys Ala Ala Gln Gln Leu Arg Phe Val Arg Gln Trp Ser
            4000                4005                4010
```

FIG. 1AJ

GTC TTT GGC AAG ACA TTA TGC CGA GCT CTG CCA GAG CTC CTG GGG GTC 12218
Val Phe Gly Lys Thr Leu Cys Arg Ala Leu Pro Glu Leu Leu Gly Val
4015 4020 4025

ACC TTG GGC CTG GTG GTG CTC GGG GTA GCC TAC GCC CAG CTG GCC ATC 12266
Thr Leu Gly Leu Val Val Leu Gly Val Ala Tyr Ala Gln Leu Ala Ile
4030 4035 4040

CTG CTC GTG TCT TCC TGT GTG GAC TCC CTC TGG AGC GTG GCC CAG GCC 12314
Leu Leu Val Ser Ser Cys Val Asp Ser Leu Trp Ser Val Ala Gln Ala
4045 4050 4055 4060

CTG TTG GTG CTG TGC CCT GGG ACT GGG CTC TCT ACC CTG TGT CCT GCC 12362
Leu Leu Val Leu Cys Pro Gly Thr Gly Leu Ser Thr Leu Cys Pro Ala
4065 4070 4075

GAG TCC TGG CAC CTG TCA CCC CTG CTG TGT GTG GGG CTC TGG GCA CTG 12410
Glu Ser Trp His Leu Ser Pro Leu Leu Cys Val Gly Leu Trp Ala Leu
4080 4085 4090

CGG CTG TGG GGC GCC CTA CGG CTG GGG GCT GTT ATT CTC CGC TGG CGC 12458
Arg Leu Trp Gly Ala Leu Arg Leu Gly Ala Val Ile Leu Arg Trp Arg
4095 4100 4105

TAC CAC GCC TTG CGT GGA GAG CTG TAC CGG CCG GCC TGG GAG CCC CAG 12506
Tyr His Ala Leu Arg Gly Glu Leu Tyr Arg Pro Ala Trp Glu Pro Gln
4110 4115 4120

FIG. 1AK

```
GAC TAC GAG ATG GTG GAG TTG TTC CTG CGC AGG CTG CGC CTC TGG ATG   12554
Asp Tyr Glu Met Val Glu Leu Phe Leu Arg Arg Leu Arg Leu Trp Met
4125                4130                4135                4140

GGC CTC AGC AAG GTC AAG GAG TTC CGC CAC AAA GTC CGC TTT GAA GGG   12602
Gly Leu Ser Lys Val Lys Glu Phe Arg His Lys Val Arg Phe Glu Gly
                4145                4150                4155

ATG GAG CCG CTG CCC TCT CGC TCC TCC AGG GGC TCC AAG GTA TCC CCG   12650
Met Glu Pro Leu Pro Ser Arg Ser Ser Arg Gly Ser Lys Val Ser Pro
            4160                4165                4170

GAT GTG CCC CCA CCC AGC GCT GGC TCC GAT GCC TCG CAC CCC TCC ACC   12698
Asp Val Pro Pro Pro Ser Ala Gly Ser Asp Ala Ser His Pro Ser Thr
        4175                4180                4185

TCC TCC AGC CAG CTG GAT GGG CTG AGC GTG AGC CTG GGC CGG CTG GGG   12746
Ser Ser Ser Gln Leu Asp Gly Leu Ser Val Ser Leu Gly Arg Leu Gly
    4190                4195                4200

ACA AGG TGT GAG CCT GAG CCC TCC CGC CTC CAA GCC GTG TTC GAG GCC   12794
Thr Arg Cys Glu Pro Glu Pro Ser Arg Leu Gln Ala Val Phe Glu Ala
4205                4210                4215                4220

CTG CTC ACC CAG TTT GAC CGA CTC AAC CAG GCC ACA GAG GAC GTC TAC   12842
Leu Leu Thr Gln Phe Asp Arg Leu Asn Gln Ala Thr Glu Asp Val Tyr
                4225                4230                4235
```

FIG. 1AL

```
CAG CTG GAG CAG CAG CTG CAC AGC CTG CAA GGC CGC AGG AGC AGC CGG   12890
Gln Leu Glu Gln Gln Leu His Ser Leu Gln Gly Arg Arg Ser Ser Arg
            4240                4245                4250

GCG CCC GCC GGA TCT TCC CGT GGC CCA TCC CCG GGC CTG CGG CCA GCA   12938
Ala Pro Ala Gly Ser Ser Arg Gly Pro Ser Pro Gly Leu Arg Pro Ala
        4255                4260                4265

CTG CCC AGC CGC CTT GCC CGG GCC AGT CGG GGT GTG GAC CTG GCC ACT   12986
Leu Pro Ser Arg Leu Ala Arg Ala Ser Arg Gly Val Asp Leu Ala Thr
    4270                4275                4280

GGC CCC AGC AGG ACA CCC CTT CGG GCC AAG AAC AAG GTC CAC CCC AGC   13034
Gly Pro Ser Arg Thr Pro Leu Arg Ala Lys Asn Lys Val His Pro Ser
4285                4290                4295                4300

AGC ACT TAGTCCTCCT TCCTGGCGGG GGTGGGCCGT GGAGTCGGAG TGGACACCGC   13090
Ser Thr

TCAGTATTAC TTTCTGCCGC TGTCAAGGCC GAGGGCCAGG CAGAATGGCT GCACGTAGGT 13150

TCCCCAGAGA GCAGGCAGGG GCATCTGTCT GTCTGTGGGC TTCAGCACTT TAAAGAGGCT 13210

GTGTGGCCAA CCAGGACCCA GGGTCCCCTC CCCAGCTCCC TTGGGAAGGA CACAGCAGTA 13270

TTGGACGGTT TCTAGCCTCT GAGATGCTAA TTTATTTCCC CGAGTCCTCA GGTACAGCGG 13330
```

FIG. 1AM

GCTGTGCCCG GCCCCACCCC CTGGGCAGAT GTCCCCCACT GCTAAGGCTG CTGGCTTCAG 13390

GGAGGGTTAG CCTGCACCGC CGCCACCCTG CCCCTAAGTT ATTACCTCTC CAGTTCCTAC 13450

CGTACTCCCT GCACCGTCTC ACTGTGTGTC TCGTGTCAGT AATTTATATG GTGTTAAAAT 13510

GTGTATATTT TTGTATGTCA CTATTTTCAC TAGGGCTGAG GGGCCTGCGC CCAGAGCTGG 13570

CCTCCCCCAA CACCTGCTGC GCTTGGTAGG TGTGGTGGCG TTATGGCAGC CCGGCTGCTG 13630

CTTGGATGCG AGCTTGGCCT TGGGCCGGTG CTGGGGGCAC AGCTGTCTGC CAGGCACTCT 13690

CATCACCCCA GAGGCCTTGT CATCCTCCCT TGCCCCAGGC CAGGTAGCAA GAGAGCAGCG 13750

CCCAGGCCTG CTGGCATCAG GTCTGGGCAA GTAGCAGGAC TAGGCATGTC AGAGGACCCC 13810

AGGGTGGTTA GAGGAAAAGA CTCCTCCTGG GGGCTGGCTC CCAGGGTGGA GGAAGGTGAC 13870

TGTGTGTGTG TGTGTGTGCG CGCGCGCACG CGCGAGTGTG CTGTATGGCC CAGGCAGCCT 13930

CAAGGCCCTC GGAGCTGGCT GTGCCTGCTT CTGTGTACCA CTTCTGTGGG CATGGCCGCT 13990

TCTAGAGCCT CGACACCCCC CCAACCCCCG CACCAAGCAG ACAAAGTCAA TAAAAGAGCT 14050

GTCTGACTGC 14060

FIG. 1AN

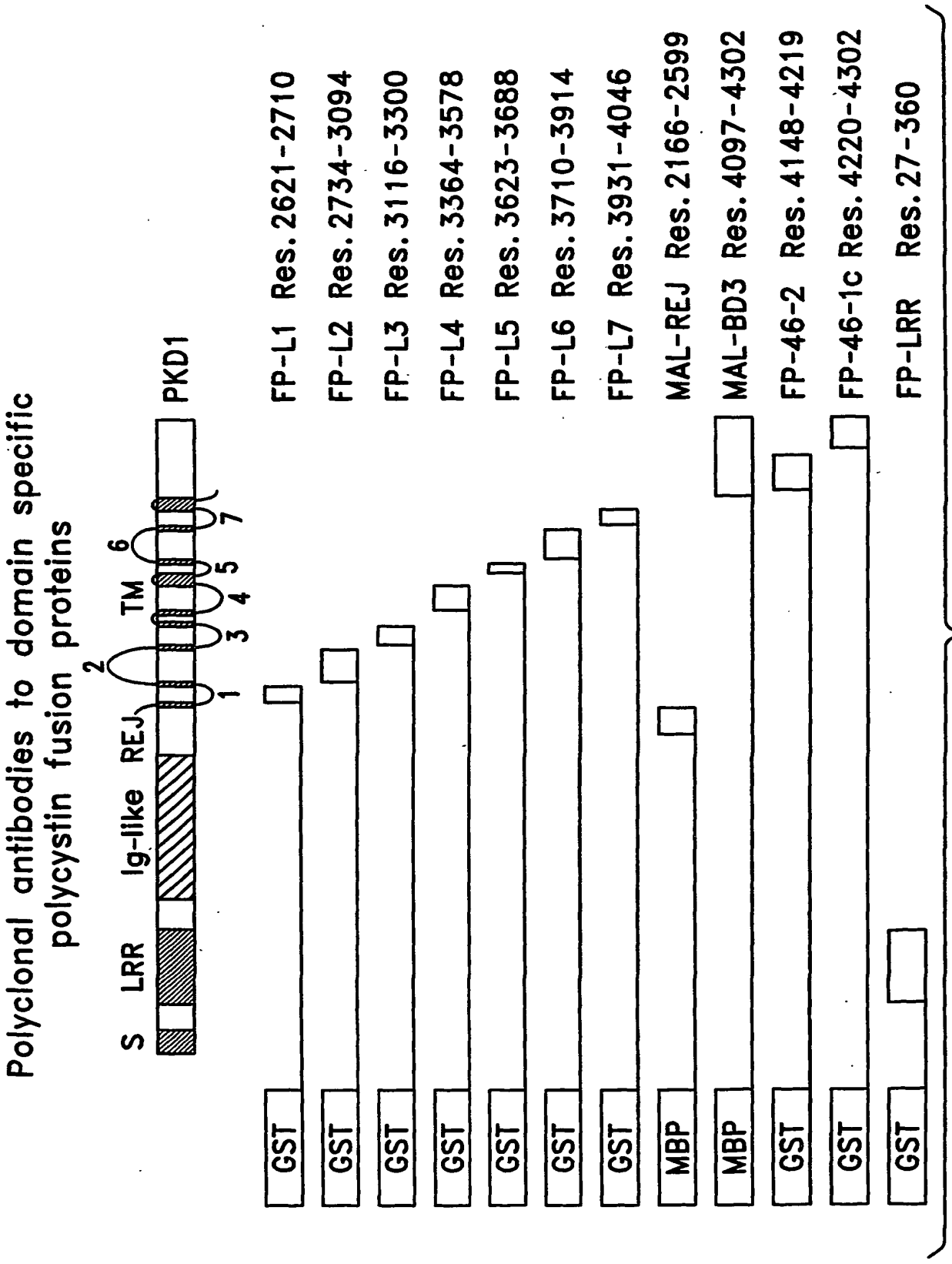

Polyclonal antibodies to domain specific polycystin fusion proteins
S LRR Ig-like REJ TM PKD1
1 2 3 4 5 6 7
FP-L1 Res. 2621-2710
FP-L2 Res. 2734-3094
FP-L3 Res. 3116-3300
FP-L4 Res. 3364-3578
FP-L5 Res. 3623-3688
FP-L6 Res. 3710-3914
FP-L7 Res. 3931-4046
MAL-REJ Res. 2166-2599
MAL-BD3 Res. 4097-4302
FP-46-2 Res. 4148-4219
FP-46-1c Res. 4220-4302
FP-LRR Res. 27-360
GST
MBP

FIG. 2

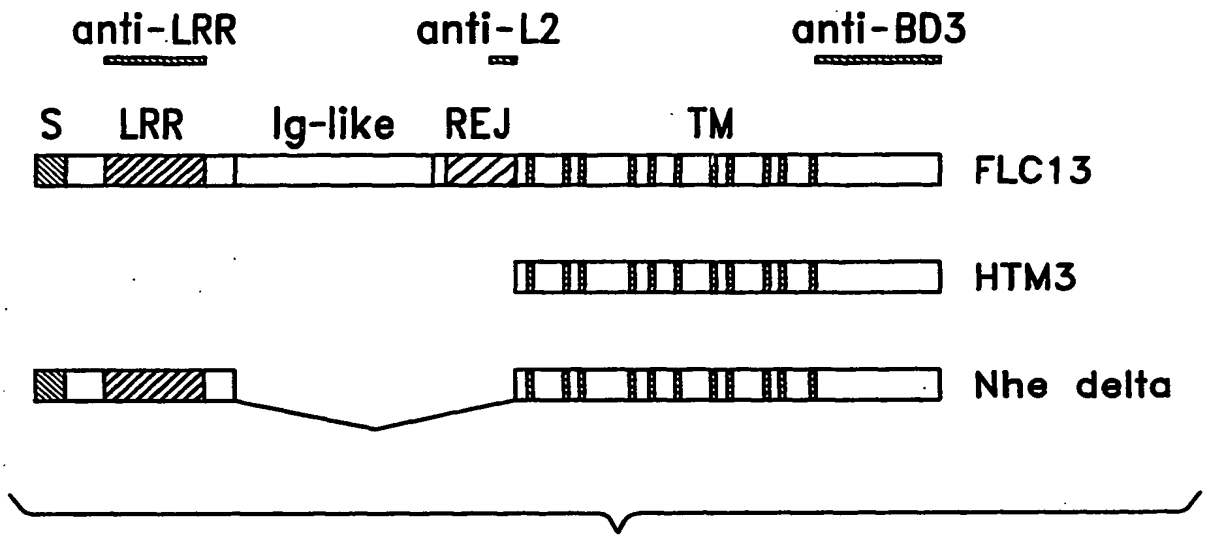
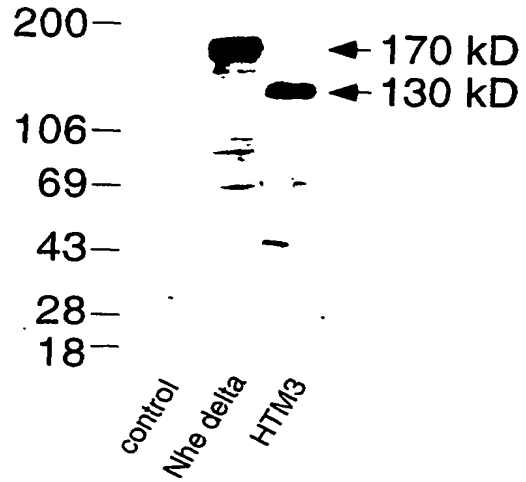

anti-LRR
anti-L2
anti-BD3
S
LRR
Ig-like
REJ
TM
FLC13
HTM3
Nhe delta

FIG. 3A

FIG. 3B


200
106
69
43
28
18
170 kD
130 kD
control
Nhe delta
HTM3

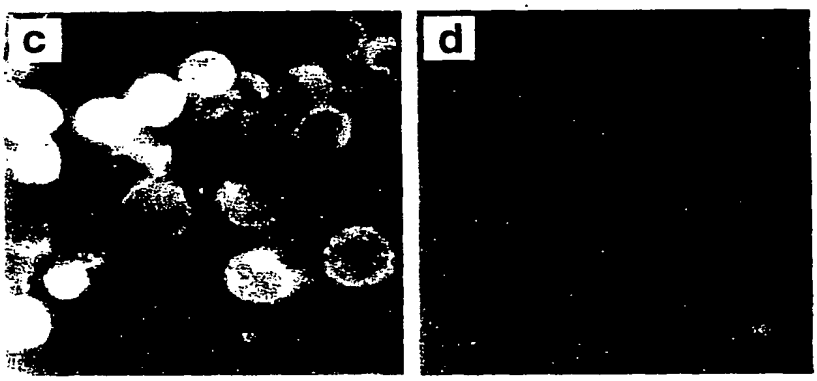


FIG. 3C FIG. 3D

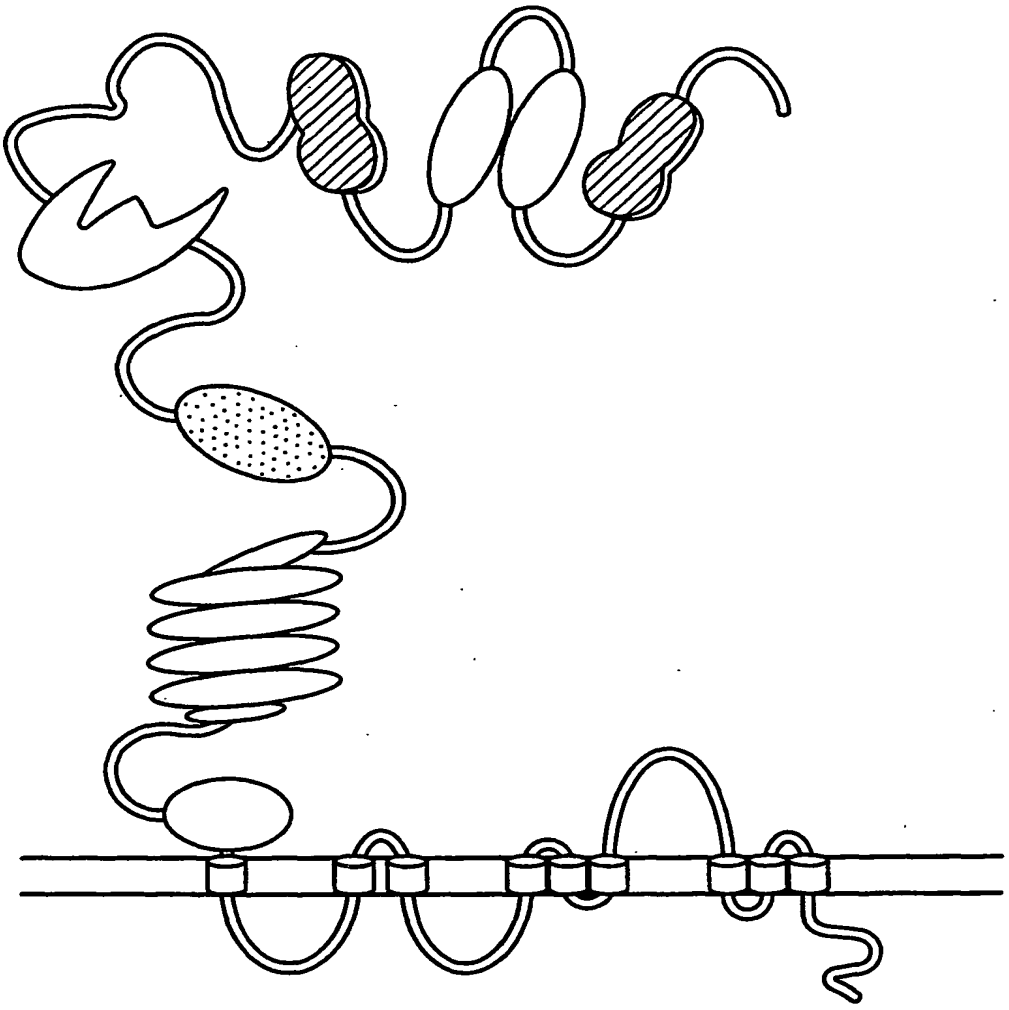

N - amino flanking region
C - carboxy flanking region
LRR - leucine-rich repeats
Ig-like domains
C-type lectin domain
REJ - domain with homology to the receptor for egg jelly
LDL-like domain
TM - putative transmembrane region

FIG. 4

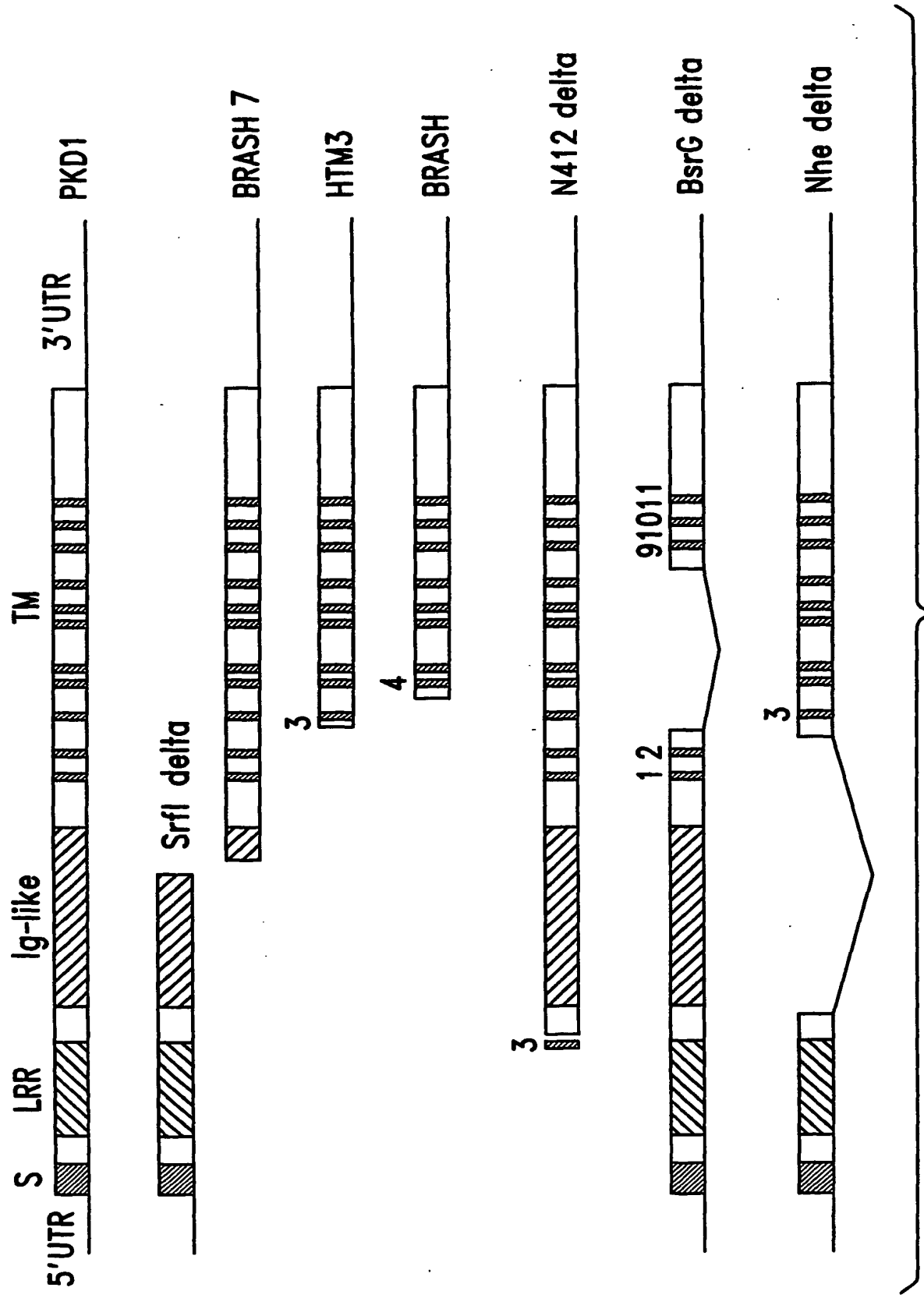

5'UTR
S
LRR
Ig-like
TM
3'UTR
PKD1
Srf1 delta
BRASH 7
3
HTM3
4
BRASH
3
N412 delta
12
91011
BsrG delta
3
Nhe delta

FIG. 5

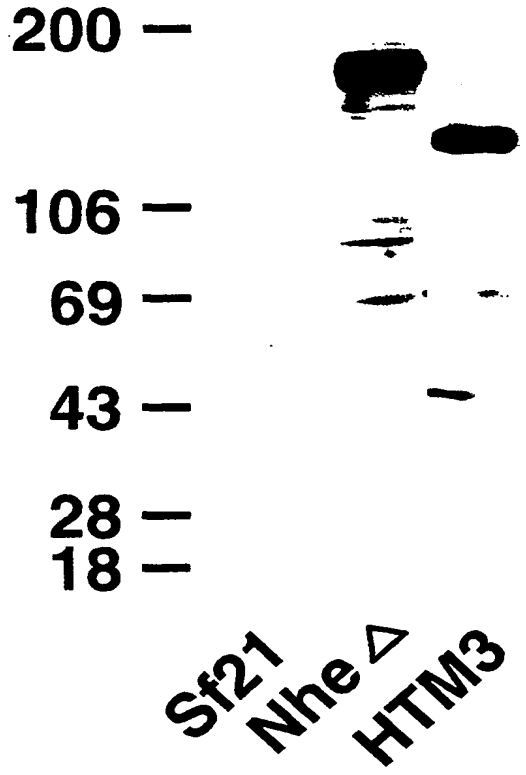
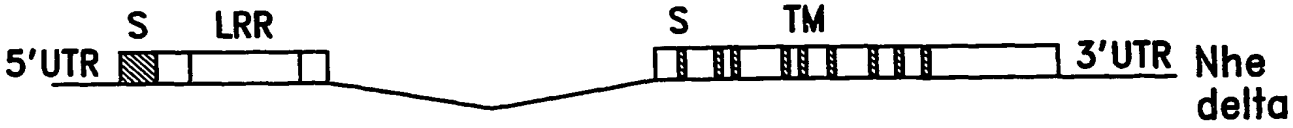


FIG. 6


S
LRR
S
TM
5'UTR
3'UTR
Nhe
delta

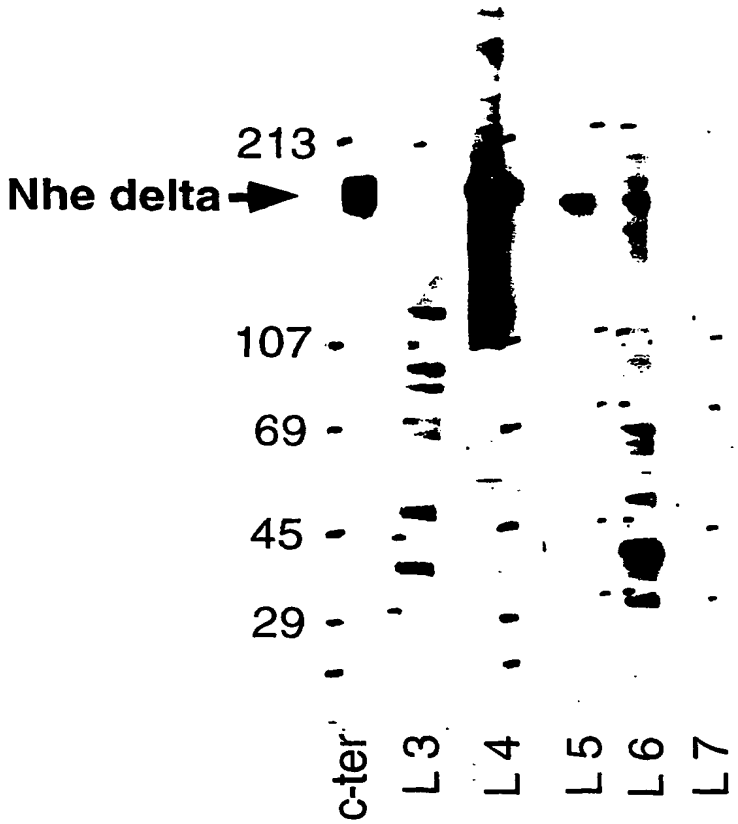

213
Nhe delta
107
69
45
29
c-ter
L3
L4
L5
L6
L7

FIG. 7

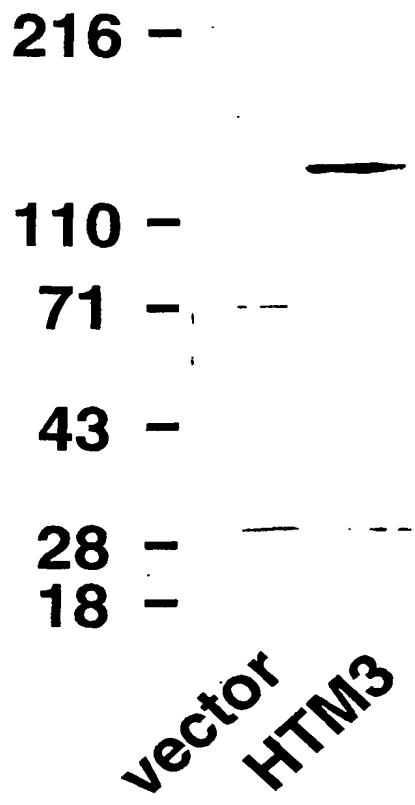


FIG. 8

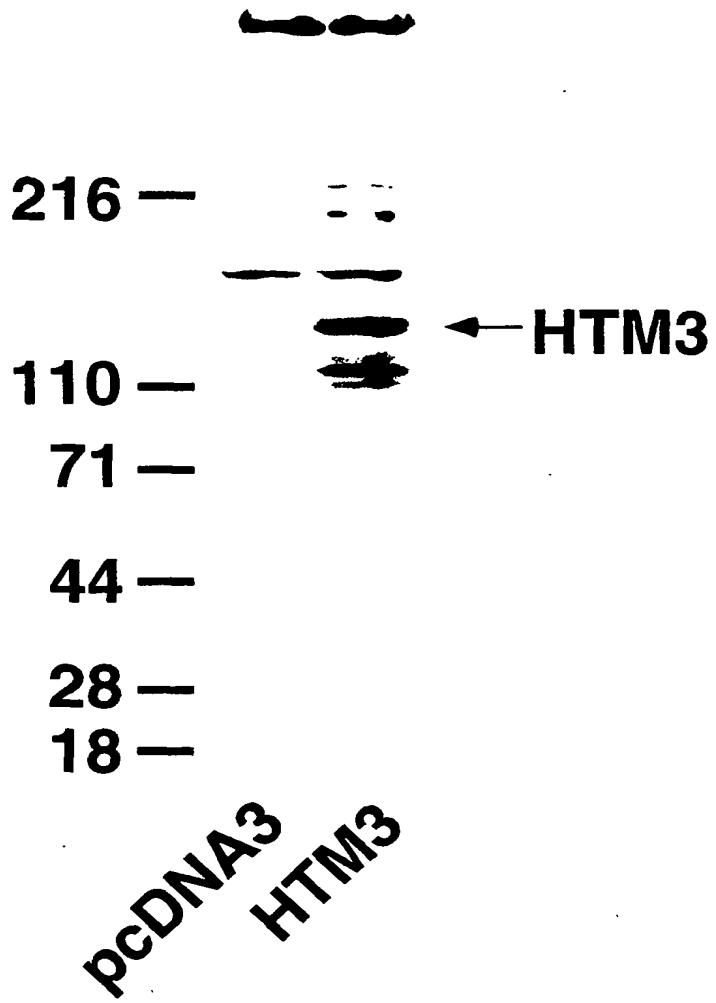


FIG. 9

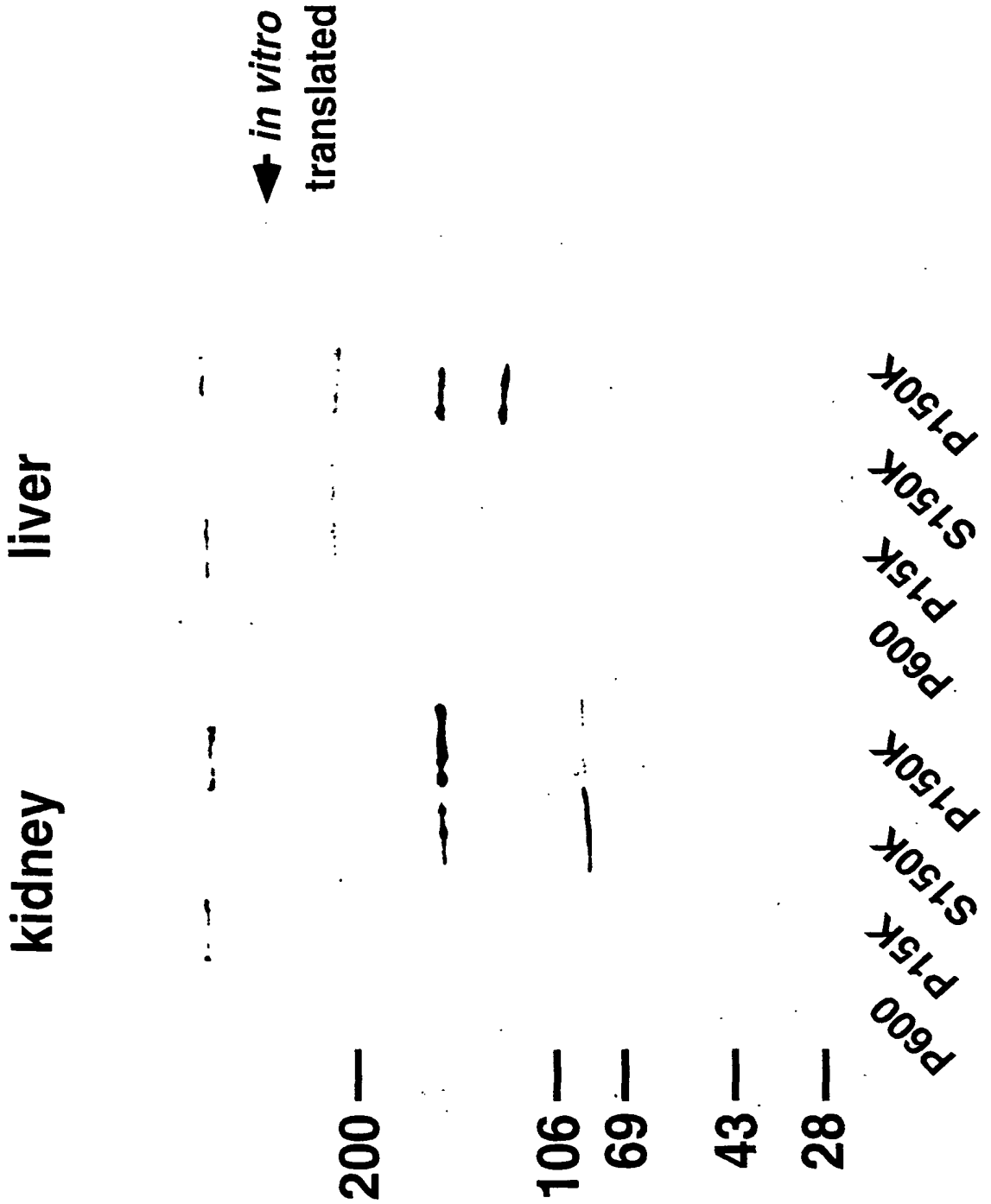

in vitro translated
kidney
liver
200
106
69
43
28
P600
P150K
S150K
P150K
P600
P150K
S150K
P150K

FIG. 10A

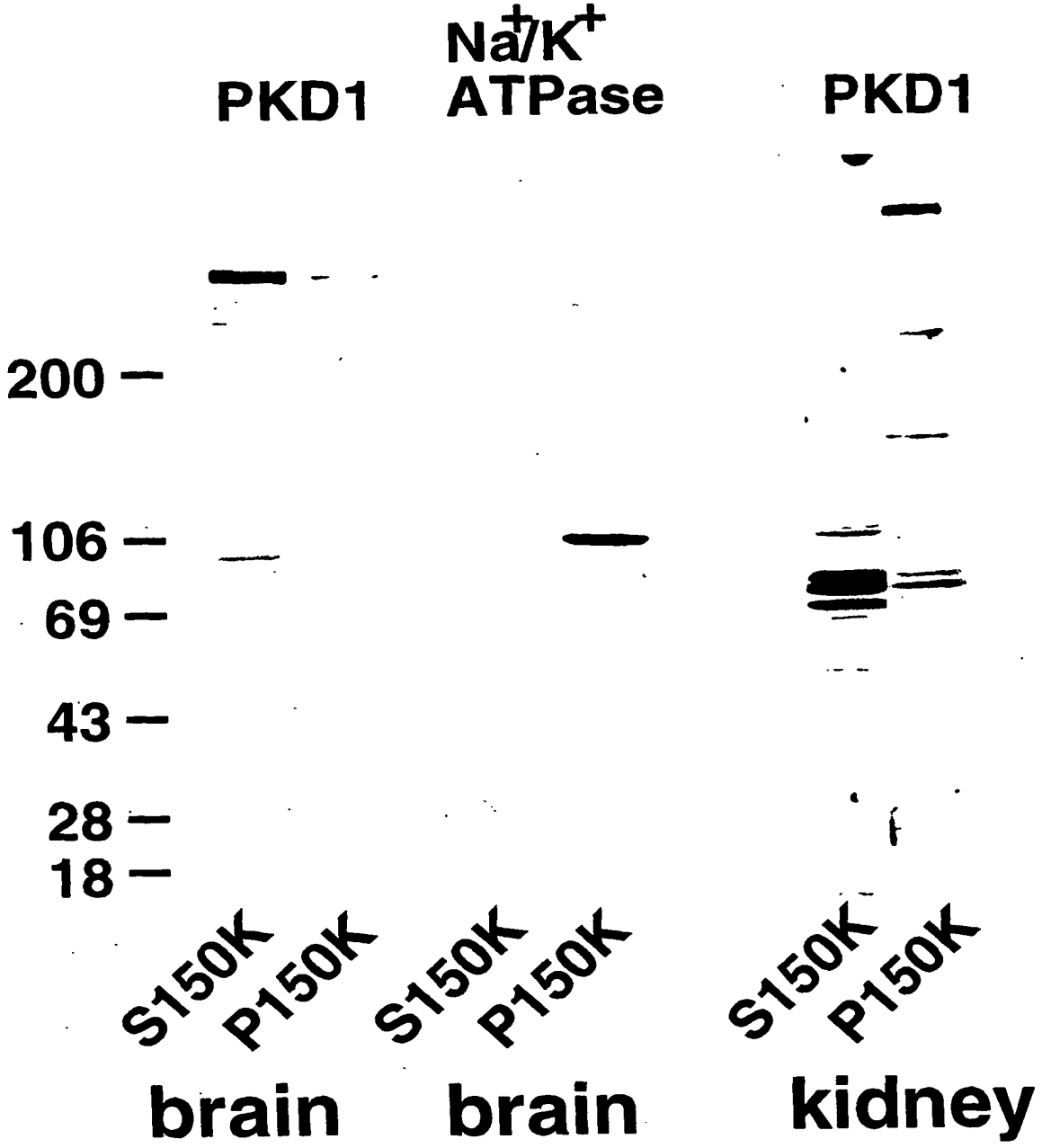

PKD1
Na+/K+ ATPase
PKD1
200
106
69
43
28
18
S150K
P150K
S150K
P150K
S150K
P150K
brain
brain
kidney

FIG. 10B

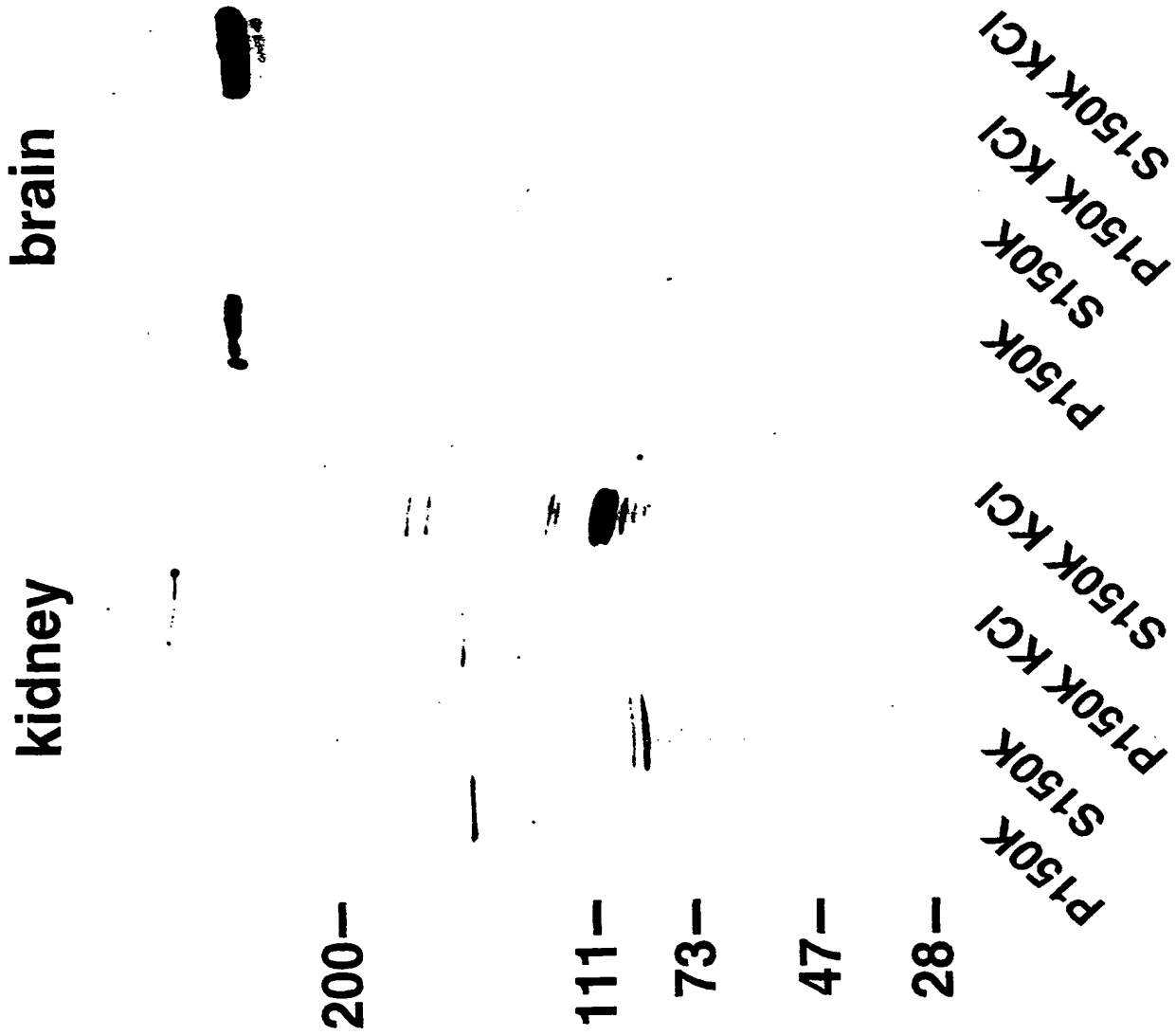


FIG. 10C

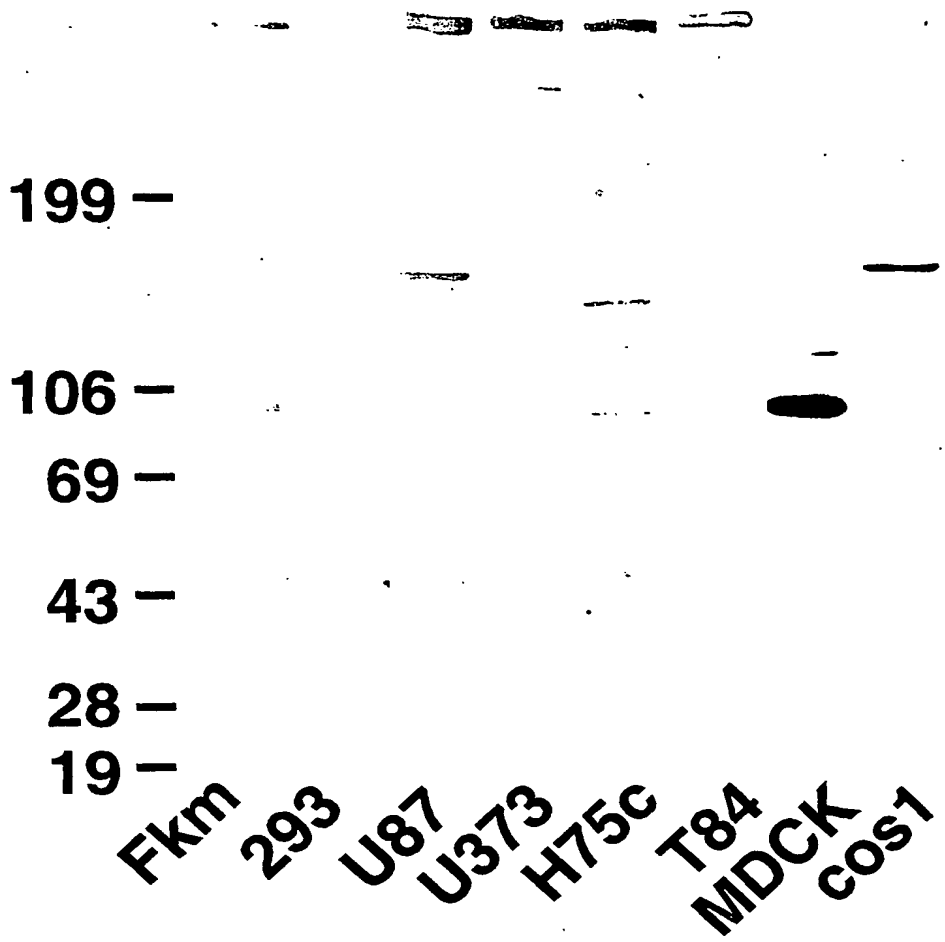


FIG. 10D

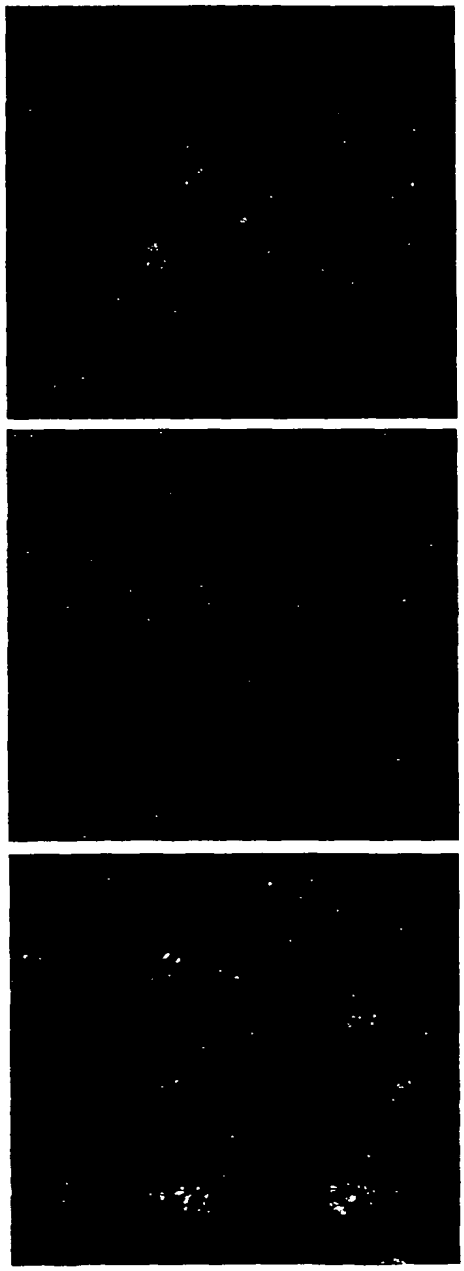


FIG. 11

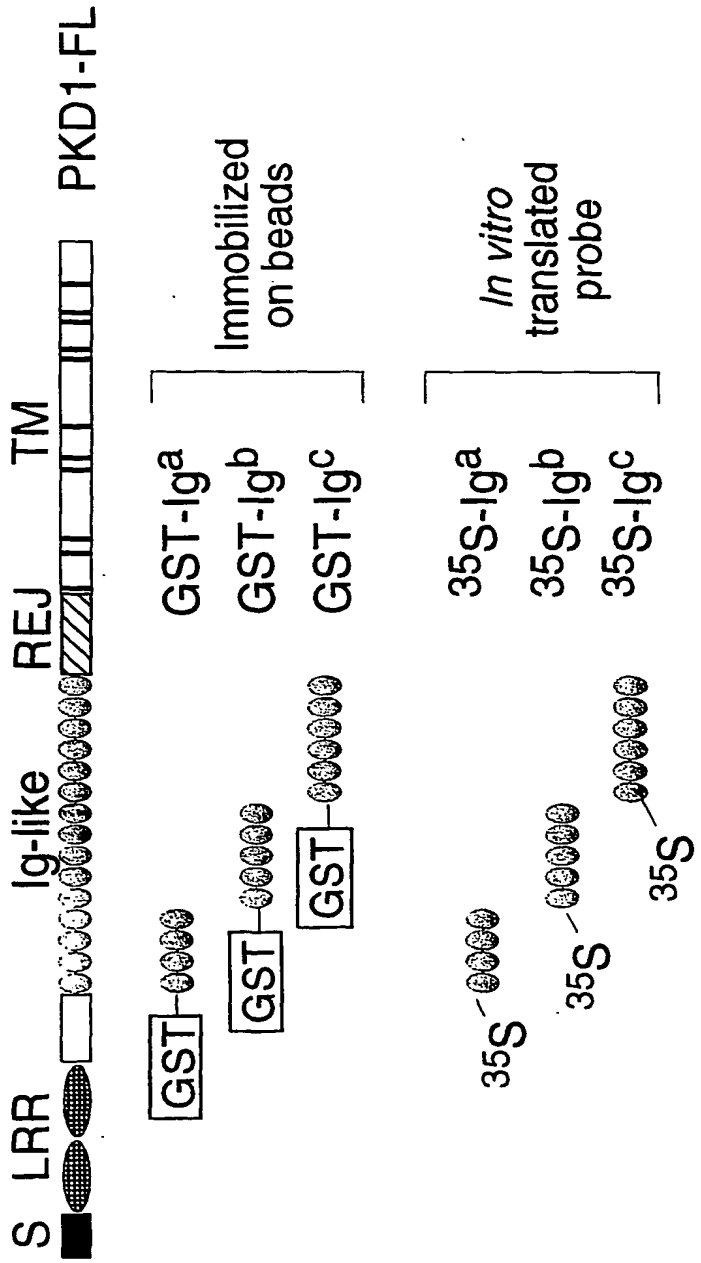

S
LRR
Ig-like
REJ
TM
PKD1-FL
GST
GST
GST
GST-Ig^a
GST-Ig^b
GST-Ig^c
Immobilized on beads
35S
35S
35S
35S-Ig^a
35S-Ig^b
35S-Ig^c
In vitro translated probe

FIG. 12A

FIG. 12B

FIG. 12C

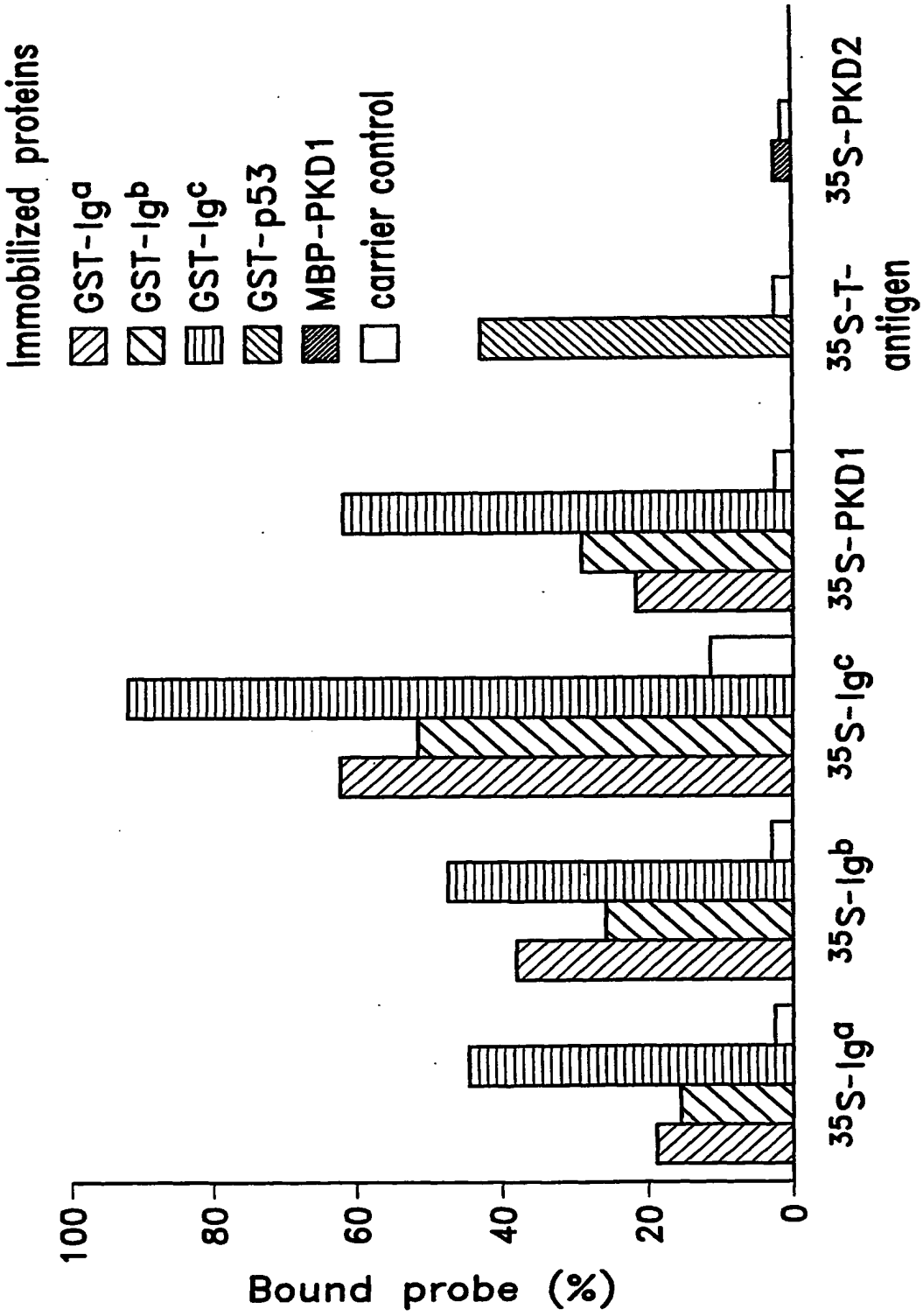

Immobilized proteins
GST-Ig^a
GST-Ig^b
GST-Ig^c
GST-p53
MBP-PKD1
carrier control
35S-Ig^a
35S-Ig^b
35S-Ig^c
35S-PKD1
35S-T-antigen
35S-PKD2
Bound probe (%)
100
80
60
40
20
0

FIG. 13

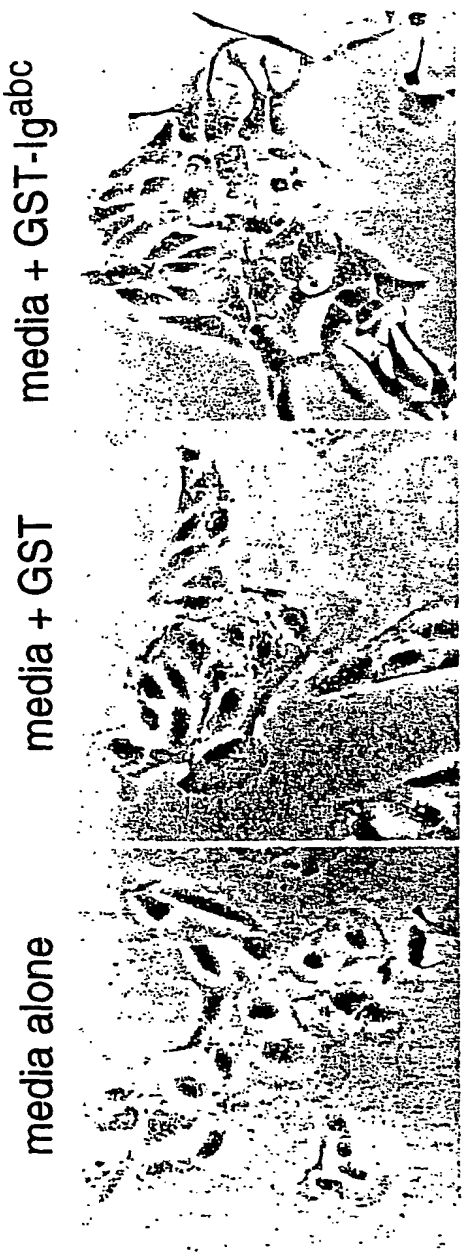


FIG. 14A

FIG. 14B

INTERNATIONAL SEARCH REPORT

Int. ional Application No
PCT/US 99/25091

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C07K16/28 C07K14/47 C12N5/12 A61K39/395 A61K38/17
C12N15/866 C12N5/10 G01N33/577 G01N33/68 C12N15/11
A61P13/12

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C07K C12N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 95 34573 A (BRIGHAM AND WOMEN'S HOSPITAL) 21 December 1995 (1995-12-21) claims 1-35	1-29
X	VAN ADELSBERG J ET AL: "Polycystin expression is temporally and spatially regulated during renal development." AMERICAN JOURNAL OF PHYSIOLOGY, (1997) 272 F602-F609, XP000892142 page F602, line 37 - line 48	1-3,6
A	PALSSON R ET AL: "Characterization and cell distribution of polycystin, the product of autosomal dominant polycystc kidney disease gene 1." MOLECULAR MEDICINE, (1996) 2 702-11, XP000892141 abstract	1-29

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
20 April 2000	11/05/2000

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Le Flao, K

1

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 99/25091

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. [X] Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

Remark: Although claims 23-29 partially (as far as an in vivo method is concernerd) are directed to a method of treatment of the human/ animal body, the search has been carried out and based on the alleged effects of the compound/composition.

2. [ ] Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. [ ] As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int ional Application No
PCT/US 99/25091

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9534573 A	21-12-1995	AU 2766195 A	05-01-1996
		US 5891628 A	06-04-1999

PATENT COOPERATION TREATY

PCT

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference	FOR FURTHER ACTION	See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)
GA0154PCT		
International application No.	International filing date *(day/month/year)*	Priority date *(day/month/year)*
PCT/US99/25091	25/10/1999	26/10/1998
International Patent Classification (IPC) or national classification and IPC C07K16/28		
Applicant GENZYME CORPORATION et al.		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.

2. This REPORT consists of a total of 11 sheets, including this cover sheet.

   ☐ This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

   These annexes consist of a total of sheets.

3. This report contains indications relating to the following items:

   - I ☒ Basis of the report
   - II ☐ Priority
   - III ☒ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
   - IV ☐ Lack of unity of invention
   - V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations suporting such statement
   - VI ☐ Certain documents cited
   - VII ☒ Certain defects in the international application
   - VIII ☒ Certain observations on the international application

Date of submission of the demand	Date of completion of this report
26/05/2000	12.12.2000
Name and mailing address of the international preliminary examining authority: European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Authorized officer Maucher, C Telephone No. +49 89 2399 7415

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/US99/25091

I. Basis of the report

1. This report has been drawn on the basis of *(substitute sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments (Rules 70.16 and 70.17).)*:
Description, pages:

1-58 as originally filed

Claims, No.:

1-13,15-29 as originally filed

Drawings, sheets:

1/35-35/35 as originally filed

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☐ furnished subsequently to this Authority in written form.
- ☐ furnished subsequently to this Authority in computer readable form.
- ☐ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☐ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

- ☐ the description, pages:
- ☐ the claims, Nos.:

☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

   (Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)

6. Additional observations, if necessary:

III. Non-establishment of opinion with regard to novelty, inventive step and industrial applicability

The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

☐ the entire international application.

☒ claims Nos. 23-29 with respect to industrial applicability.

because:

☒ the said international application, or the said claims Nos. claims 23-29 relate to the following subject matter which does not require an international preliminary examination (*specify*):
see separate sheet

☐ the description, claims or drawings (*indicate particular elements below*) or said claims Nos. are so unclear that no meaningful opinion could be formed (*specify*):

☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed.

☐ no international search report has been established for the said claims Nos. .

2. A meaningful international preliminary examination report cannot be carried out due to the failure of the nucleotide and/or amino acid sequence listing to comply with the standard provided for in Annex C of the Administrative Instructions:

☐ the written form has not been furnished or does not comply with the standard.

☐ the computer readable form has not been furnished or does not comply with the standard.

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N) Yes: Claims

	No:	Claims	1-13, 15-29
Inventive step (IS)	Yes:	Claims	
	No:	Claims	1-13, 15-29
Industrial applicability (IA)	Yes:	Claims	1-22
	No:	Claims	

2. Citations and explanations
see separate sheet

VII. Certain defects in the international application

The following defects in the form or contents of the international application have been noted:
see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:
see separate sheet

Point III:

Claims 23-29 relate to subject-matter considered by this Authority to be covered by the provisions of Rule 67.1(iv) PCT. Consequently, no opinion has been formulated with respect to the industrial applicability of the subject-matter of these claims (Article 34(4)(a)(i) PCT).

Said claims do not explicitly exclude treatment of the human or animal body.

Point V:

Reference is made to the following documents:

D1: WO-A-95 34573
D2: AMERICAN JOURNAL OF PHYSIOLOGY, (1997) 272, F602-F609
D3: MOLECULAR MEDICINE, (1996) 2 702-11

The document D4 was not cited in the international search report. A copy of the document is appended hereto:

D4: WO-A-95 34649

1. Articles 33(2) and (3) PCT

1.1. Claim 1 does not meet the requirements of Article 33(2) or (3) PCT in the light of any one of the documents D1, D2 or D4:

The subject-matter of **claim 1** is anticipated by D3, since D3 also describes an antibody recognizing a polycystin-related polypeptide having a molecular weight of 642 kD (page 705, left to right column, bridging sentence and page 703, right column, full paragraph). Polycystin is synonymous for PKD1 protein (see page 7, lines 21-22 of the present application).

Furthermore, D2 seems to anticipate claim 1, since it discloses the use of antibodies against peptides derived from the membrane-spanning portion of the PKD1 gene (abstract). On figure 3B, second lane, the antibody does not only bind to polypeptides of 485, 245 and 55kD, but also to a polypeptide which seems to have a molecular weight between 600 and 800kD.

Moreover, D4 discloses an antibody specific for the PKD1 protein or a fragment thereof with the amino acid sequence disclosed in figures 10 or 15 (page 82, lines 2-3; claim 20 in connection with page 27, lines 17-18 (entire sequence apart from its extreme 5' end) and page 29, lines 21-22 (entire transcript)).

The same applies to dependent **claims 6** (D1, page 26, lines 8-9; D3, rabbits were immunized and the anti-peptide antibodies were purified: page 703, right column, full paragraph; D2, F602, right column, last full paragraph, 1. line), **claim 7** (D1, page 26, lines 8-9, D4, page 82, line 20), **claim 8** (D1, page 36, lines 23-26; D4, page 82, lines 23-24) and **claim 11** (D1, page 19, lines 2-7; D2, D3: abstract: kidney; D4: page 76, line 26).

The subject-matter of dependent **claims 2-3** is not considered novel, in the light of D4, figures 7, 10 and 15. The same applies to **claims 19-21**, which disclose said polypeptide.

1.2. The subject-matter of **claims 4-5** does not appear to be novel in the light of D1-D4 for the following reasons:
Remark: Claim 4 is interpreted as disclosing an isolated antibody specific for an (antigen comprising an) epitope instead of an antibody comprising an epitope (see also VIII, 3).

Claims 4 and 5 are considered to embrace an antibody against the full length PKD-1 gene product. Thus, the subject-matter of both claims is anticipated by an antibody directed against the full length PKD-1 gene product of D1, claim 15 and D4, claim 20 in connection with page 27, lines 17-18 (see also page 82, lines 2-3).

Furthermore, an antibody which was raised against a fusion protein comprising GST and PKD1 coding region (D1, page 36, lines 22-30) does also anticipate the subject-matter of **claim 5**.

Moreover, **claim 4** lacks novelty in view of D2 and D3. These documents disclose antibodies raised against linear peptides of epitopes that are located within the regions referred to in claim 4: D2 discloses the peptide B145 for antibody production (page F602, right column, last full paragraph), which is located between the animo acids 3623 and 3688 and the peptide B146 (page F602, right column, last full paragraph), which is located between the amino acids 3710 and 3914 of polycystin. D3 mentions the peptide P2 for antibody production (page 703, right column, lines 20-21), which is located between amino acids 4097 and 4302 of polycystin (see the full length amino acid sequence in D4, fig. 15). Consequently, these antibodies are expected to bind also in the regions of interest of claim 4.

1.3. Due to its broad wording, **claim 9** does not appear novel in the light of D1. D1 also discloses preparations (page 54, lines 25-31) containing antibodies against PKD1 (page 50, line 27 to page 51, line 3) and carriers (pages 55-56, bridging sentence).

1.4. The subject-matter of **claim 10** is not considered novel in the light of D1 (page 27, line 26) or D4 (page 82, line 18), since both documents disclose hybridomas for producing antibodies.
Even if novelty could be established, a claim directed to a hybridoma would not be inventive, since the skilled person knows how to produce hybridomas for particular antibodies.

1.5. The subject-matter of **claims 12-13** does not meet the requirements of Article 33(2) PCT in the light fo D1, D3 or D4 for the following reasons:

- It is not apparent in which structural features recombinant polypeptide as defined in claim 12 or 13 differs from wild type polycystin. Therefore, the scope of said claims is considered to embrace also wildtype polycystin as disclosed in D1 (native full length polypeptide polycystin in figure 6; see also

page 19, lines 36-37).
D1 furthermore discloses a fusion protein of the PKD1 coding region and GST (page 36, lines 22-30), which is a recombinant polypeptide and therefore also anticipates the subject-matter of claims 12-13.

Moreover, the PKD1 protein comprises many domains also present in other proteins, for instance the LDL module (D1, page 90, lines 5-10, see also page 88, lines 5-8; see also the present application, page 2, line 27 to page 3, line 24).
Thus, proteins comprising the same domains like, in this case the LDL receptor, also anticipate the subject-matter of claim 13.

- Claim 12 is furthermore anticipated by D3 (page 704, column 1, 1. paragraph) and claim 13 is anticipated by D3 (page 703, right column, lines 12 and 16).

- Claims 12-13 are moreover anticipated by D4, figure 10, which discloses the entire PKD1 gene sequence apart from its extreme 5' end (page 27, lines 17-18) and figure 15, which discloses the sequence of the PKD1 transcript (page 29, lines 21-22).

1.6. The subject-matter of **claim 15** is anticipated by D1 (claim 33) disclosing a composition comprising the PKD1 gene product and a carrier.
Furthermore, the peptides coupled to BSA in D3 (page 703, lines 20-23) also fall within the scope of claim 15.
Said subject-matter is also anticipated by D4, page 19, lines 9-12.

1.7. **Claim 16** does not meet the requirements of Article 33(2) PCT, since D1 discloses the full length polynucleotide PKD1 in figure 6 and the PKD1 coding region fused to GST on page 36, lines 22-26.
D4 also anticipates the subject-matter of claim 16 disclosing the nucleic acid sequence according to figures 10 and 15 (see claims 2 and 4).

1.8. **Claim 17** is not novel in the light of D1 or D4.
D1 discloses pGEX vectors comprising PKD1 protein coding sequence or

PKD1 fused to the GST coding sequence (page 21, lines 12-34).
D4 also describes a vector comprising a nucleic acid sequence (page 17, lines 21-22 and claim 12).

1.9. **Claim 18** is anticipated by the disclosed engineered cell lines expressing the PKD1 protein of D1, page 23, last line and D4, claim 13 or page 17, lines 23-25.

1.10. The kit disclosed in **claim 22** is not novel, since D1 also discloses such a kit comprising an anti PKD1 protein (polycystin) antibody (page 58, lines 19-20). Said subject-matter is furthermore disclosed in D4, page 20, line 7.

1.11. The subject-matter of **claims 23-29** does not appear to be novel in the light of D4, page 8, line 16 ff.

2. Industrial Applicability

For the assessment of the present **claims 23-29** on the question whether they are industrially applicable, no unified criteria exist in the PCT contracting States. The patentability can also be dependent upon the formulation of the claims. The EPO, for example, does not recognize as industrially applicable the subject-matter of claims to the use of a compound in medical treatment, but may allow, however, claims to a known compound for first use in medical treatment and the use of such a compound for the manufacture of a medicament for a new medical treatment.

Point VII:

1. Contrary to the requirements of Rule 5.1(a)(ii) PCT, the documents D1-D4 have not been identified in the description and the relevant background art disclosed therein hs not been briefly discussed.

2. The expression "incorporated herein by reference" used throughout the document (e.g. page 1, line 8) has not been deleted, although the description should be

supported by itself.

3. Claim 14 is missing. The claims have not been renumbered for clarity reasons (Rule 6.1b) PCT).

4. The statement concerning related applications on page 1, first paragraph, is not relevant for international files. It has however not been deleted.

Point VIII:

1. The subject-matter of claims 1-3, and therefore also claim 10, as well as claims 19-21, is unclear (Article 6 PCT). The definition of the protein in terms of its roughly estimated molecular weight without the disclosure of the molecular weight determination method that has been used does not enable the skilled person to identify the said protein.

2. Claims 1 and 22 lack clarity (Article 6 PCT) in the following respects:

   - the term "polycystin-related polypeptide" is open to interpretation and leaves the reader in doubt as to the exact nature of the protein (see also VIII, 1).

   - the wording "apparent molecular weight...of about" is open to interpretation and does not define the specificity of the claimed antibody. The combined use of obscure definitions concerning the antigen to be recognized does not permit the unambiguous identification of antibodies falling within the scope of these claims.

3. The subject-matter of claim 4 is unclear (Article 6 PCT), since an antibody comprising an epitope does not exist (see also V, 1.2).

4. The subject-matter of claim 5 is unclear (Article 6 PCT), since the disclosed antibody is identified by internal designations which are meaningless to the skilled person. The applicant is invited to remove this defect.
   Reference could have been made to depositions, where available.

5. The subject-matter of claim 19 is unclear (Article 6 PCT), since the polypeptide of claim 19 is defined by the unclear antibody disclosed in claim 1 (see VIII, 1, 2).

6. The subject-matter of claim 22 is unclear (Article 6 PCT), since it contains the feature "instructions for use" which is not a technical feature for a kit claim (see Rule 6.3(a) PCT and the Guidelines, IV, 2.4(e)).

7. The vague and imprecise statement "spirit and scope thereof" (page 58, line 17) implies that the subject-matter for which protection is sought may be different to that defined in the claims, thereby resulting in lack of clarity of the claims (Article 6 PCT) when used to interpret them (see the Guidelines, C-III, 4.3a). This statement has not been amended to remove inconsistency.

PATENT COOPERATION TREATY

PCT

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference	**FOR FURTHER ACTION** see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.	
126881206140		
International application No.	International filing date *(day/month/year)*	(Earliest) Priority Date *(day/month/year)*
PCT/US 99/25091	25/10/1999	26/10/1998
Applicant		
GENZYME CORPORATION et al.		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 3 sheets.

[X] It is also accompanied by a copy of each prior art document cited in this report.

1. **Basis of the report**

   a. With regard to the **language**, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

   [ ] the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

   b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :

   [X] contained in the international application in written form.

   [X] filed together with the international application in computer readable form.

   [ ] furnished subsequently to this Authority in written form.

   [ ] furnished subsequently to this Authority in computer readble form.

   [ ] the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.

   [ ] the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. [X] **Certain claims were found unsearchable** (See Box I).

3. [ ] **Unity of invention is lacking** (see Box II).

4. With regard to the **title,**

   [X] the text is approved as submitted by the applicant.

   [ ] the text has been established by this Authority to read as follows:

5. With regard to the **abstract,**

   [X] the text is approved as submitted by the applicant.

   [ ] the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

6. The figure of the **drawings** to be published with the abstract is Figure No. _____

   [ ] as suggested by the applicant.

   [ ] because the applicant failed to suggest a figure.

   [ ] because this figure better characterizes the invention.

   [X] None of the figures.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 99/25091

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. [X] Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

Remark: Although claims 23-29 partially (as far as an in vivo method is concernerd) are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

2. [ ] Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. [ ] As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

[redacted] International Application No
[redacted]/US 99/25091

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C07K16/28 C07K14/47 C12N5/12 A61K39/395 A61K38/17
C12N15/866 C12N5/10 G01N33/577 G01N33/68 C12N15/11
A61P13/12

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C07K C12N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 95 34573 A (BRIGHAM AND WOMEN'S HOSPITAL) 21 December 1995 (1995-12-21) claims 1-35	1-29
X	VAN ADELSBERG J ET AL: "Polycystin expression is temporally and spatially regulated during renal development." AMERICAN JOURNAL OF PHYSIOLOGY, (1997) 272 F602-F609, XP000892142 page F602, line 37 - line 48	1-3,6
A	PALSSON R ET AL: "Characterization and cell distribution of polycystin, the product of autosomal dominant polycystc kidney disease gene 1." MOLECULAR MEDICINE, (1996) 2 702-11, XP000892141 abstract	1-29

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
20 April 2000	11/05/2000

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL – 2280 HV Rijswijk Tel. (+31–70) 340–2040, Tx. 31 651 epo nl, Fax: (+31–70) 340–3016	Le Flao, K

1

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No
/US 99/25091

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9534573 A	21-12-1995	AU 2766195 A	05-01-1996
		US 5891628 A	06-04-1999